

# TREATMENT OF HIV-ASSOCIATED BURKITT LYMPHOMA

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## ABSTRACT

Burkitt lymphoma (BL) is a highly aggressive B-cell malignancy, occurring with increased frequency among patients infected with HIV. For several years, the immunocompromised state of HIV-positive patients was advocated as a sufficient reason to avoid the intensive chemotherapeutic regimens used in HIV-negative BL. However, with the introduction of the highly active antiretroviral therapy (HAART), the subsequent improvement of the immunological state of HIV-positive patients, and the disappointing results of less intensive schedules, investigators began to apply the same chemotherapeutic regimens used as a gold standard in HIV-negative non-Hodgkin lymphoma (NHL), including the use of rituximab. Despite promising results of different schedules in early-phase studies, agreement on the treatment of HIV-positive BL is still lacking, and further trials are needed to define a standard of care. Moreover, new treatment frontiers need to focus on improving the outcome for patients with advanced immunosuppression, unfavourable prognostic features- such as advanced stages and high International Prognostic Index (IPI) scores - and for those with adverse tumour biology.

This paper aims to revise the main epidemiological and physiopathological features of HIV-positive BL, to summarise the most relevant steps in the treatment of affected patients, and to elucidate the role of HAART in allowing HIV-positive patients to be managed with the therapeutic strategies currently used in HIV-negative patients with BL.

**Keywords:** Burkitt lymphoma, AIDS, HIV, rituximab, chemoimmunotherapy, myc.

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## INTRODUCTION

Burkitt lymphoma (BL) is an aggressive B-cell non-Hodgkin lymphoma (NHL) with a high cellular turnover and a cell-doubling time of 24-48 hours. The World Health Organization's (WHO) classification of haematological malignancies describes three clinical and epidemiologic variants of BL: endemic, sporadic and HIV-associated BL (HIV-BL).<sup>1</sup> BL accounts for up to 20% of lymphoproliferative malignancies diagnosed in HIV-positive subjects. In Western countries, symptoms related to BL are often the first clinical clues of AIDS onset.<sup>2</sup> The untamed replication kinetics of cancer cells translates into the clinical presentation of the lymphoma, with an extensive extranodal involvement, and high mortality rate when

treated with conventional chemotherapy regimens used in patients with diffuse large B-cell lymphoma (DLBCL). For this reason, several chemotherapeutic regimens for patients with HIV-BL have been developed in the last few years. These combinations often include the anti-CD20 monoclonal antibody rituximab, resulting in combinations with excellent efficacy, but important limitations in feasibility and tolerability.

This paper is focused on the different chemotherapeutic regimens used to treat HIV-BL, the impact of the introduction of the highly active antiretroviral therapy (HAART), the role of rituximab and other important drugs, and the rapport between toxicity and efficacy of each chemoimmunotherapy

regimen. Evidence from different trials concerning the introduction of the HAART and the use of rituximab in the chemotherapeutic regimens, which still lack unanimous consent, will be presented as well.

## EPIDEMIOLOGY AND RISK FACTORS

HAART, widely used in clinical practice from 1996, has dramatically changed the natural history of HIV infection and AIDS development and has significantly improved the outcomes of HIV-positive patients in terms of host immune response, reduction in opportunistic infections, and incidence of HIV-related NHL (HIV-NHL). Before HAART, the incidence of NHL in HIV-positive patients was 60-200 times higher than in HIV-negative patients;<sup>3</sup> the incidence of HIV-NHL decreased from 13.6 per 100,000 person-years before 1996 to only 1.8 per 100,000 person-years between 2002 and 2006.<sup>4</sup> Nevertheless, the incidence HIV-BL has not been affected by HAART with progressive rates increasing.<sup>5</sup>

The risk of development of BL in HIV-positive patients is 15-fold higher than HIV-negative ones.<sup>6</sup> The mean age of onset is 38 years old, and males are more likely to be affected. Conversely to the incidence of other HIV-NHL, which increases with age, the incidence of HIV-BL shows three different age-related peaks: infancy, adulthood and old age. This trimodal trend has been reported also in the HIV-negative population,<sup>7</sup> and underlies the possibility that age itself, instead of age-related level of immunosuppression, could play a role in the development of HIV-BL. A different relative risk for the development of HIV-BL as related to the way of HIV transmission, has failed to be demonstrated.

## PATHOGENESIS

BL development is strongly related to the overexpression of *c-myc* oncogene. The gene encoding for MYC, a cell-cycle regulator, is located on the long arm of chromosome 8. There are three types of balanced translocation, common to all the clinical variants of BL, involving the *c-myc* locus: the t(8;14)(q24;q32), which involves immunoglobulin heavy chain (IGH) genes located in 14q32 and is detected in 70-80% of BL patients; the t(2;8)(p12;q24), which involves IG kappa-light chain locus in the 2p12, and is detected in 15% of BL; and the t(8;22)(q24;q11), which involves IG lambda-light chain genes in the 22q11 and is detected in 5% of BL patients. In all cases, MYC locus is downstream to IG gene enhancers, causing gene overexpression.<sup>8</sup> Expression of MYC in normal

B-lymphocytes induces cell apoptosis by a p53-dependent pathway. In malignant B-cells, apoptosis is prevented by mutations in TP53 tumour suppressor gene. Moreover, p53-independent pathways, such as overexpression in B-cell lymphoma 2 (Bcl-2), due to down-regulation of Bim protein, inhibits apoptosis.<sup>9</sup> However, in HIV-positive patients who are not diagnosed with HIV-BL translocation in *c-myc* locus are frequent, which suggests that this mechanism alone is not enough to induce neoplastic proliferation of B-lymphocytes. The translocation seems to be enhanced when the enzyme activation-induced cytidine deaminase (AID) is over-expressed. AID is involved in the IGH class switch.<sup>7</sup> Enzyme induction is favoured by chronic antigenic stimulation of B-lymphocytes, or by signalling, which enhances enzyme over-expression itself.

As in other lymphomas, relative risk increases with high HIV viral load and with the duration of exposure to such elevated viral load.<sup>10,11</sup> These features suggest a pathogenic role of HIV in BL, even though molecular mechanisms of the potential oncogenic effect of HIV have never been demonstrated.<sup>12</sup> Putative mechanisms regard the effect of elevated viral loads inducing a chronic antigenic stimulus for B-cells and cytokines production that may enhance the activity of the AID, promoting *c-myc* translocation and survival signalling to mutated B-lymphocytes.<sup>10</sup> Moreover, one of the HIV envelope proteins, the CD40 ligand, is able to activate AID itself,<sup>13</sup> and Trans-Activator of Transcription (TAT) protein is able to promote B-lymphocytes cell cycle with two different mechanisms. On one hand, TAT enhances the activation of cellular genes, such as IL-6 and IL-10; on the other, TAT inhibits Rb2/p130 protein, a tumour suppressor gene involved in halting cell cycle from G0/G1.<sup>14,15</sup>

Risk for the development of HIV-BL increases with CD4-positive cell count: the majority of patients have  $\geq 250/\mu\text{L}$  CD4, while the risk lowers when CD4 are  $< 50/\mu\text{L}$ , with only 15% of cases in patients showing CD4  $< 100/\mu\text{L}$ . This is in contrast with other HIV-NHL where the relative risk increases with lowering of CD4.<sup>16</sup> An adequate number of functional CD4 cells seems to be necessary for HIV-BL development and, paradoxically, a competent immune system may have a causative role.<sup>17</sup> CD4+ lymphocytes may be involved in survival signalling to B-lymphocytes of germinal centres, carrying *c-myc* translocation, and in preventing cell death. On the contrary, low CD4 counts may fail to protect aberrant B-lymphocytes, promoting the initiation

**Table 1. Diagnosis and staging of HIV-BL.**

DIAGNOSIS							
<b>HISTOLOGY</b>	The cells seem to be molded and the cytoplasm is deeply basophilic with squared-off cytoplasmic margins. A 'starry sky' appearance is due to scattered tangible body-laden macrophages that contain apoptotic tumour cells. <sup>79</sup> There are three histologic variants, all of them having very high mitotic rates, with a Ki67 proliferation index close to 100%, and with a high cellular turnover suggested by increased apoptosis.						
	<table border="1"> <tr> <td><b><i>BL with plasmacytoid differentiation</i></b></td> <td> <ul style="list-style-type: none"> <li>- Most common in HIV-BL</li> <li>- It is associated with EBV in 50% to 70% of cases</li> <li>- It is characterised by medium-sized cells with abundant basophilic cytoplasm and eccentric nuclei</li> </ul> </td> </tr> <tr> <td><b><i>BL classic</i></b></td> <td> <ul style="list-style-type: none"> <li>- Accounts for 30% of HIV-BL and it resembles endemic BL</li> <li>- It is associated with EBV in 30% of cases</li> <li>- It is characterised by intermediate-size cells containing coarse chromatin and prominent basophilic nucleoli</li> </ul> </td> </tr> <tr> <td><b><i>Atypical BL</i></b></td> <td> <ul style="list-style-type: none"> <li>- It is characterised by cells with greater nuclear pleomorphism and fewer but more prominent nuclei</li> </ul> </td> </tr> </table>	<b><i>BL with plasmacytoid differentiation</i></b>	<ul style="list-style-type: none"> <li>- Most common in HIV-BL</li> <li>- It is associated with EBV in 50% to 70% of cases</li> <li>- It is characterised by medium-sized cells with abundant basophilic cytoplasm and eccentric nuclei</li> </ul>	<b><i>BL classic</i></b>	<ul style="list-style-type: none"> <li>- Accounts for 30% of HIV-BL and it resembles endemic BL</li> <li>- It is associated with EBV in 30% of cases</li> <li>- It is characterised by intermediate-size cells containing coarse chromatin and prominent basophilic nucleoli</li> </ul>	<b><i>Atypical BL</i></b>	<ul style="list-style-type: none"> <li>- It is characterised by cells with greater nuclear pleomorphism and fewer but more prominent nuclei</li> </ul>
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<b>IMMUNOPHENOTYPE</b>	<ul style="list-style-type: none"> <li>- Mature B-cell with germinal centre cell differentiation and expression of surface immunoglobulins (sIgs) with light chain restriction</li> <li>- Positive for: CD19, CD20, CD22, CD79a, CD10, Bcl-6</li> <li>- Negative for: CD5, CD23, terminal deoxynucleotidyl transferase (TdT)</li> <li>- Bcl-2: usually negative; however, Bcl-2 can be expressed in 10% to 20% of cases<sup>80</sup></li> </ul>						
<b>CYTOGENETICS</b>	Karyotypic analysis of neoplastic cells are aimed to identify <i>c-myc</i> translocation, by fluorescence in situ hybridisation. Three patterns can be observed.						
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<b>STAGING</b>	<ul style="list-style-type: none"> <li>- For adult patients Cotswold-modified An Arbor Staging system is used<sup>83</sup></li> <li>- For paediatric patients St Jude/Murphy Staging system is used (stage IV defined by CNS or bone-marrow involvement)<sup>84</sup></li> </ul>						
<b>PHYSICAL EXAMINATION</b>	<ul style="list-style-type: none"> <li>- Evaluation of performance status are mandatory</li> </ul>						
<b>BLOOD CHEMISTRY</b>	<ul style="list-style-type: none"> <li>- Full blood count (CD4 count), complete biochemical profile, serum lactate dehydrogenase and uric acid (to assess tumour turnover), viral infections assessment (EBV, HIV viral load, hepatitis B/C viruses)</li> </ul>						
<b>INSTRUMENTAL EXAMS</b>	<ul style="list-style-type: none"> <li>- <sup>18</sup>F-Fluorodeoxyglucose (<sup>18</sup>F-FDG) positron emission tomography/computed tomography (PET/CT),<sup>85,86</sup> enhanced total body CT scan</li> <li>- Bone marrow aspirate and biopsy (including flow cytometry and cytogenetic evaluations)</li> <li>- Cerebrospinal fluid sampling by lumbar puncture, with cytologic examination, flow cytometry, physico-chemical examination</li> <li>- Brain MRI (if CNS involvement is suspected or confirmed)</li> <li>- Echocardiography</li> <li>- Some exams should be indicated in the case of clinical or biochemical suspicion of organ involvement, for instance endoscopic studies</li> </ul>						

of cell death programs. Moreover, with a lower CD4 number, fewer B-lymphocytes are activated and *c-myc* translocation rate lowers.<sup>7</sup>

Epstein-Barr virus (EBV) was detected in <40% of cases of HIV-BL and in ~100% of cases of endemic BL.<sup>18</sup> This peculiarity seems to lessen the causative role of this virus in this malignancy. One possible explanation is that neoplastic changes occur in highly proliferating B-lymphocytes in the germinal centre in EBV-negative BL, while in EBV-positive cases, they involve memory B-cells, whose survival is promoted by viral infection itself.<sup>19</sup> EBV is able to promote lymphocytes oncogenic transformation, not only with a chronic antigenic stimulation and the consequent cytokines deregulation, but also by means of specific protein production. Epstein-Barr nuclear antigen 1 (EBNA-1) protein allows EBV latent persistence in B-cells genome, increases genetic instability of B-lymphocytes, and could enhance those mutations involved in the selection of the neoplastic clone.<sup>20</sup> Moreover, production of latent membrane protein 1 (LMP-1) promotes both proliferative signalling, via the activation of NF- $\kappa$ B pathway, and anti-apoptotic stimulation via BCL-2 overexpression in B-cells carrying *c-myc* translocation.<sup>21</sup>

## CLINICAL PRESENTATION, DIAGNOSIS, STAGING

HIV-BL is a highly aggressive disease, usually presenting with systemic symptoms, advanced stage disease, as well as extensive extranodal involvement. With respect to sporadic BL, HIV-BL shows more common bone marrow infiltration (46% versus 20%), bulky disease (54% versus 13%), less common abdominal lesions (46% versus 91%), rarer leukaemic dissemination, and more frequent meningeal involvement (38% versus 14%), the latter being asymptomatic in 25% of cases.<sup>22</sup>

In the absence of pathognomonic features, diagnosis is formulated on the basis of histologic, immunophenotypic and cytogenetic investigations on the excision biopsy of an involved organ, preferring a lymph node (Table 1). Once the diagnosis is obtained, following the WHO criteria,<sup>1</sup> staging must be performed (Table 1).

## TREATMENT

### General Background

BL is a highly chemosensitive malignancy, achieving high response rates with available chemotherapy

combinations. However, patients who reach complete remission (CR) must be closely monitored, as most relapses occur within the first year<sup>23</sup> and are uncommon after 2 years.<sup>24</sup> While some studies had demonstrated the prognostic role of the International Prognostic Index (IPI), and CD4 cell count and complex karyotype,<sup>25</sup> a therapeutic decision is not usually based on these variables. Most patients are managed with intensified chemoimmunotherapy combinations independently of IPI risk and extension of disease. Only Central Nervous System (CNS) involvement can change therapeutic choice in HIV-BL. Patients with HIV-BL should be managed with modern chemotherapy (see below) in cancer centres with appropriate expertise. This is an important issue considering that failure to achieve complete remission (CR) after first-line chemotherapy is a definitively negative prognostic event.<sup>25</sup> Importantly, the management of these patients is complex, mostly due to the high rates or co-morbidity, co-infections and treatment-related events, as well as the necessity to prevent complications and to manage symptoms. Prevention and treatment of tumour lysis syndrome with alkaline infusions and allopurinole/rasburicase administration, and appropriate G-CSF use are strongly encouraged. While actively treated patients must undergo antifungal prophylaxis with fluconazole, antiviral prophylaxis with acyclovir and prophylaxis for *Pneumocystis jiroveci* with trimethoprim-sulfamethoxazole. During CT, nadir antibiotic prophylaxis with a quinolone is appropriated.

Due to the high frequency of CNS involvement, both at presentation and relapse, the use of intravenous drugs able to penetrate the blood brain barrier and to achieve therapeutic concentrations in the CNS, as well as drug delivery by intrathecal route, are required.

HAART is a relevant component of the treatment of HIV-NHL, and BL is no exception. Before 1996, no standardised treatment was available for HIV-BL, since all HIV-NHL types were managed with the same strategy. Low-dose and less-intensive combinations led to a poor outcome in patients with HIV-BL; CHOP (cyclophosphamide, doxorubicin, vincristine and prednisone) regimen was associated with a CR rate of 30-50% and a median survival of 6-9 months,<sup>26,27</sup> and 58% and 18 months, respectively, with the CDE infusion combination.<sup>28</sup> Near 60% of deaths were due to AIDS-related events and not to lymphoma. As a consequence, less intensive approaches have been proposed (m-BACOD), with a median survival



**Table 2. Schedule of Hyper-CVAD, PETHEMA-LAL3/07-GMALL regimen (± Rituximab), CODOX-M/IVAC standard e/o modified, DA-EPOCH (± Rituximab).**

Cyclophosphamide (CTX), vincristine (VCN), doxorubicin (DOXO), dexamethasone (DEX), methotrexate (MTX), cytarabine (Ara-C), Prednisone (PDN), ifosfamide (IFO), teniposide (VM26), rituximab (R), vindesine (VND), etoposide (VP16), therapy intra-thecal (TIT), intravenous (i.v.), continuous infusion (civ.), orally (o.), days (d), hours (h), patients (pts), chemotherapy (CT), absolute neutrophil count (ANC).

- a) Ara-C to 1 g/m<sup>2</sup> if pts>60 y.
- b) MTX 0.5 g/m<sup>2</sup> if pts>50 y.
- c) Two additional doses of rituximab were administered 3 and 6 weeks after the last C cycle.
- d) Depending on centre.
- e) Maximum dose 750 mg/m<sup>2</sup>.
- f) VP16 reduce by 25% if ANC nadir<500/μL or platelets<25,000/μL (both for at least 3 d) If this occurs during a cycle in which no CTX was given or creatinine clearance<40 mL/min.
- g) DOXO reduce by 25% if ANC nadir<500/μL or platelets<25,000/μL (both for at least 3 d) if this occurs during a cycle in which no CTX was given or direct bilirubin>2.5 mg/dl.
- h) VCN reduce to 0.3 mg/m<sup>2</sup>/day if constipation or unable to walk on heels; reduce to 25% if direct bilirubin>2.5 mg/dl or creatinine clearance<40 mL/min; discontinue if difficulty ambulating.
- i) R discontinue if skin/mucositis attributed to R.

<b>HYPER-CVAD REGIMEN<sup>49</sup></b>											
<ul style="list-style-type: none"> <li>- <b>8 courses</b> of alternating intensive CT every 3 weeks</li> <li>- <b>TIT:</b> 12 mg of MTX d 2 and 100 mg of Ara-C d 7 every cycle</li> <li>- <b>HAART</b> during CT is recommended</li> </ul>											
Odd-numbered Courses (1-3-5-7)						Even-numbered Courses (2-4-6-8)					
<b>CTX</b>	300 mg/m <sup>2</sup>	i.v. twice	d 1-3			<b>MTX</b>	1000 mg/m <sup>2</sup>	i.v. over 24 h	d 1		
<b>VCN</b>	2 mg	i.v.	d 4, 11								
<b>DOXO</b>	50 mg/m <sup>2</sup>	i.v.	d 4			<b>Ara-C<sup>a</sup></b>	3000 mg/m <sup>2</sup>	i.v. twice	d 2-3		
<b>DEX</b>	40 mg/d	i.v. or o.	d 1-4 and d 11-14								
<b>PETHEMA REGIMEN<sup>53</sup></b>											
<ul style="list-style-type: none"> <li>- <b>8 courses</b> of alternating intensive CT every 3 weeks</li> <li>- <b>Pre-phase</b> (to prevent lysis syndrome) <b>CTX</b> 200 mg/m<sup>2</sup> i.v. and <b>PDN</b> 60 mg/m<sup>2</sup> i.v. d 1-5</li> <li>- <b>TIT:</b> MTX 12 mg, Ara-C 30 mg, hydrocortisone 20 mg d 1 and 5 of each cycle</li> <li>- <b>HAART</b> during CT is recommended</li> </ul>											
Odd-numbered A-cycles (induction and 2-4-6)						Even-numbered B-cycles (1-3-5-7)					
<b>VCN</b>	2 mg	i.v.	d 1			<b>VCN</b>	2 mg	i.v.	d 1		
<b>MTX<sup>b</sup></b>	3000 mg/m <sup>2</sup>	i.v. over 24 h	d 1			<b>MTX</b>	3000 mg/m <sup>2</sup>	i.v. over 24 h	d 1		
<b>IFO</b>	800 mg/m <sup>2</sup>	i.v.	d 1-5			<b>CTX</b>	200 mg/m <sup>2</sup>	i.v.	d 1-5		
<b>DEX</b>	10 mg/d	i.v. or o.	d 1-5			<b>DEX</b>	10 mg/d	i.v. or o.	d 1-5		
<b>VM26</b>	100 mg/m <sup>2</sup>	i.v.	d 4-5			<b>DOXO</b>	25 mg/m <sup>2</sup>	i.v.	d 4-5		
<b>Ara-C</b>	150 mg/m <sup>2</sup>	i.v. twice	d 4-5								
<b>PETHEMA REGIMEN + R<sup>54</sup></b>											
<ul style="list-style-type: none"> <li>- <b>6 courses</b> of alternating intensive CT every 4 weeks</li> <li>- <b>Pre-phase</b> (to prevent lysis syndrome) <b>CTX</b> 200 mg/m<sup>2</sup> i.v. and <b>PDN</b> 60 mg/m<sup>2</sup> i.v. d 1-5</li> <li>- <b>TIT:</b> MTX 15 mg, Ara-C 40 mg, DEX 20 mg d 2 and 6 of each A e B cycles</li> <li>- <b>HAART</b> during CT is recommended</li> </ul>											
A-cycles (1-4)				B-cycles (2-5)				C-cycles (3-6) <sup>c</sup>			
<b>R</b>	375 mg/m <sup>2</sup>	i.v.	d 1	<b>R</b>	375 mg/m <sup>2</sup>	i.v.	d 1	<b>R</b>	375 mg/m <sup>2</sup>	i.v.	d 1
<b>VCN</b>	2 mg	i.v.	d 2	<b>VCN</b>	2 mg	i.v.	d 2	<b>VND</b>	3 mg/m <sup>2</sup> (max. 5 mg)	i.v.	d 2

<b>MTX</b>	1500 mg/m <sup>2</sup>	i.v. over 24 h	d 2	<b>MTX</b>	1500 mg/m <sup>2</sup>	i.v. over 24 h	d 2	<b>MTX</b>	1500 mg/m <sup>2</sup>	i.v. over 24 h	d 2
<b>IFO</b>	800 mg/m <sup>2</sup>	i.v.	d 2-6	<b>CTX</b>	200 mg/m <sup>2</sup>	i.v.	d 2-6	<b>DEX</b>	10 mg/d	i.v.	d 2-6
<b>VM26</b>	100 mg/m <sup>2</sup>	i.v.	d 5-6	<b>DEX</b>	10 mg/d	i.v.	d 2-6	<b>VP16</b>	250 mg/m <sup>2</sup>	i.v.	d 5-6
<b>Ara-C</b>	150 mg/m <sup>2</sup>	i.v. twice	d 5-6	<b>DOXO</b>	25 mg/m <sup>2</sup>	i.v.	d 4-5	<b>Ara-C</b>	2000 mg/m <sup>2</sup>	i.v. twice	d 6

### CODOX-M/IVAC REGIMEN<sup>55</sup>

- low risk pts received 3 cycles of CODOX-M
- high risk pts received alternating cycles of CODOX-M, IVAC, CODOX-M, IVAC
- HAART during CT is recommended

CODOX-M				IVAC			
<b>CTX</b>	800 mg/m <sup>2</sup>	i.v.	d 1	<b>IFO</b>	1500 mg/m <sup>2</sup>	i.v.	d 1-5
<b>CTX</b>	200 mg/m <sup>2</sup>	i.v.	d 2-5	<b>VP16</b>	60 mg/m <sup>2</sup>	i.v.	d 1-5
<b>DOXO</b>	40 mg/m <sup>2</sup>	i.v.	d 1	<b>Ara-C</b>	2000 mg/m <sup>2</sup>	i.v. twice	d 1-2
<b>VCN</b>	1.5 mg/m <sup>2</sup>	i.v.	d 1, 8, 15				
<b>MTX</b>	6720 mg/m <sup>2</sup>	i.v. over 24 h	d 10				
TIT Ara-C 70 mg d 1, 3 MTX 12 mg d 15				TIT MTX 12 mg d 5			

### CODOX-M/IVAC REGIMEN Modified

<b>Noy et al.<sup>56</sup></b>	R 375 mg/m <sup>2</sup> d 1, <b>CTX</b> 800 mg/m <sup>2</sup> twice d 1-2, <b>VCN</b> 2 mg cap, <b>MTX</b> 3000 mg/m <sup>2</sup>
<b>Montoto et al.<sup>58</sup></b>	<b>VCN</b> 1.5 mg/m <sup>2</sup> (cap 2 mg), <b>MTX</b> 3000 mg/m <sup>2</sup> for pts>65 y <b>MTX</b> decreased to 1000 mg/m <sup>2</sup> , <b>IFO</b> decreased to 1000 mg/m <sup>2</sup> , <b>Ara-C</b> decreased to 1000 mg/m <sup>2</sup>
<b>Barnes et al.<sup>60</sup></b>	R 375 mg/m <sup>2</sup> d 1, <b>CTX</b> 800 mg/m <sup>2</sup> twice d 1-2, <b>DOXO</b> 50 mg/m <sup>2</sup> , <b>VCN</b> 2 mg cap, <b>MTX</b> 3000 mg/m <sup>2</sup> , <b>TIT Ara-C</b> 50 mg
<b>Rodrigo et al.<sup>57</sup></b>	<b>CTX</b> 800 mg/m <sup>2</sup> twice d1-2, <b>DOXO</b> 50 mg/m <sup>2</sup> , <b>VCN</b> 2 mg cap, <b>R</b> 375 mg/m <sup>2</sup> d 8 (of CODOX-M) and d 4 (of IVAC), <b>MTX</b> 3000 mg/m <sup>2</sup> or standard <sup>(d)</sup> , <b>TIT Ara-C</b> 50 mg (during CODOX-M) and <b>MTX</b> 12 mg (twice during IVAC) or standard <sup>(d)</sup>

### DA-EPOCH REGIMEN<sup>65</sup>

- 6 courses every 3 weeks
- TIT total 8 MTX 12 mg d 1,5 of cycles 3 through 6

<b>Oral therapy d 1-5</b>	<b>PDN</b> 60 mg/m <sup>2</sup> /d
<b>Infused agents (civ. of 96 h) d 1-4</b>	<b>VP16<sup>f</sup></b> 50 mg/m <sup>2</sup> /d, <b>DOXO<sup>g</sup></b> 10 mg/m <sup>2</sup> /d, <b>VCN<sup>h</sup></b> 0.4 mg/m <sup>2</sup> /d
<b>Bolus agents d 5</b>	
<b>CTX</b> First cycle	After cycle 1 (DA-CTX <sup>e</sup> )
if CD4cells≥100/μL <b>CTX</b> 375 mg/m <sup>2</sup>	if nadir ANC>500/μL or platelets>25,000/μL <b>CTX</b> ↑187 mg above previous cycle
If CD4cells<100/μL <b>CTX</b> 187 mg/m <sup>2</sup>	if nadir ANC<500/μL or platelets<25,000/μL <b>CTX</b> ↓187 mg above previous cycle

### DA-EPOCH-R REGIMEN<sup>47</sup>

Same schedule DA-EPOCH with additional R 375<sup>i</sup> mg/m<sup>2</sup>

<b>Arm A:</b> before each EPOCH cycle	<b>Arm B:</b> weekly times 6 weeks after EPOCH completed
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of 8 months, and ~10% of patients alive at 2 years.<sup>29-31</sup> The introduction of HAART has contributed to significant improvement of host immune response, reduced the risk of opportunistic infection,<sup>32-35</sup> and allowed HIV-positive patients to be treated with the same strategies used in HIV-negative patients,<sup>36</sup> often with similar results.<sup>37</sup> The risks and benefits of continuing HAART during chemotherapy have been variably interpreted. Some physicians are concerned by the uncontrolled HIV replication that may worsen immune function, whereas others are concerned by the risk of adverse effects of HAART on efficacy. Pharmacokinetic analyses have showed a 1.5-fold reduction in cyclophosphamide clearance in patients treated with CHOP plus HAART, while no changes in doxorubicin clearance.<sup>38</sup> CD4 increase during chemotherapy has raised the concern that HAART protects T-cells from chemotherapy. In HIV-BL patients treated with DA-EPOCH and SC-EPOCH-RR, HAART has been suspended for ~7 weeks in most cases,<sup>39,40</sup> without a relevant increase of infections, and with only a transient increase of HIV viral loads and decrease of CD4 cells. There is some concern over the use of protease inhibitors (PIs) during chemotherapy as these drugs induce cytochrome P450 isoenzymes 3A (CYP 3A) inhibition. However, data from clinical trials suggest similar clinical outcome when protease inhibitor-containing regimen are compared with non-PI-based regimen, but possibly at the expense of greater myelotoxicity.<sup>41</sup> Available evidence seems to suggest that HAART interruption during short-term chemotherapy is irrelevant from a clinical point of view, while longer treatments should request continued HAART assumption.

## The Role of Rituximab

The advent of rituximab, a monoclonal antibody against the B-cell antigen CD20, has significantly improved outcomes in several B-cell lymphomas, including BL.<sup>42</sup> The intense expression of CD20 in tumour cells provides a strong rationale for the use of this antibody in Burkitt-oriented chemotherapies. However, the application of rituximab to HIV-BL is controversial, mostly due to its potential risk of additional immunosuppression and increased incidence of major infectious events, which was shown in a large randomised clinical trial.<sup>43</sup> Subsequent studies have not confirmed this excessive infectious risk,<sup>44</sup> and some groups have introduced rituximab to their prior protocols to treat HIV-NHL patients, with a high CR rate and without increased toxicities.<sup>45,46</sup> A recent trial from the Aids Malignancy Consortium

(AMC) confirmed a good tolerance of the rituximab-chemotherapy combination, with or without HAART, though increased infectious deaths in patients with a CD4 count <50 cells/mL remained problematic.<sup>47</sup>

## CHOP and Rituximab-CHOP Regimens

Among the first-generation schedule, CHOP and CHOP-like regimen were extensively studied in national cooperative-group trials and have been considered the standard approach for patients with aggressive NHL in the HIV setting. Full dose CHOP combined with HAART has been associated with 48% CRR, and a 5-year EFS of 40%.<sup>38,32</sup> As expected, dose reduction of CHOP drugs have been associated with better tolerability, while efficacy remained unchanged.<sup>32</sup> In HAART era, the wide use of rituximab-CHOP (R-CHOP) regimen has been associated with remarkably improved results in HIV-positive patients with DLBCL, while survival figures in BL have remained largely disappointing. In fact, median time to progression (TTP) was 22 weeks and 157 weeks for HIV-BL and other NHL, respectively,<sup>43</sup> and survival data were not statistically different among patients with HIV-BL and patients with other lymphoma categories, especially DLBCL. The response rate for DLBCL was 81% and the response rate for HIV-BL was 73%.<sup>48</sup>

## Intensified Regimens

Hyper-CVAD was one of the first regimens assessed in HIV-BL<sup>49</sup> (Table 2); this combination was assessed in a series of HIV-associated aggressive lymphomas, with only six cases of HIV-BL. HAART has been assumed by 64% of patients, with no evidence of increased toxicity.<sup>50</sup> Overall, tolerability and efficacy in HIV-BL are similar to those obtained in HIV-negative patients.<sup>50,51</sup> There are no data on addition of rituximab to this schedule in HIV-BL. This combination has been associated with infectious events in 85% of cases, severe myelosuppression, neurotoxicity, a 15% treatment-related mortality (TRM), and only 23% of patients completed the planned treatment (eight courses) (Tables 3).

A multidrug combination derived from the German Multicenter Study Group for Adult Acute Lymphoblastic Leukemia GM-ALL B-ALL 05/93 protocol (Table 2) has been assessed in HIV-BL patients by PETHEMA Group.<sup>52,53</sup> More recently, this complex regimen was combined with rituximab and assessed in a prospective trial.<sup>54</sup> The addition of rituximab has been associated with improved tolerability and efficacy, but improvements should

**Table 3. Feasibility, tolerability and toxicity of more commonly used chemotherapy regimens.**

Patients (pts), number (N.º), days (d), chemotherapy (CT), treatment-related mortality (TRM), not reported (NR), rituximab (R).

a) Only 6 pts had HIV-BL.

b) Treatment termination due to reasons different from progression disease.

c) 14 pts with HIV-BL (other with L3ALL).

d) 16 pts with HIV-BL (other with L3ALL).

e) Toxicity expressed in number of courses.

f) 4 pts not received rituximab.

g) 7 pts with HIV-BL.

h) 8 pts with HIV-BL.

i) 16 pts with HIV-BL.

l) 11 pts with HIV-BL.

m) 10 pts with HIV-BL.

Regimen	Pts N°	Planned CT (days)	TRM (%)	Severe infections	Mycotic infections	Mucositis	Incompleted treatment <sup>b</sup> (%)
Hyper-CVAD <sup>49</sup> (prospective)	13 <sup>a</sup>	168	15	85	23	NR	77
PETHEMA-LAL3/97-GMALL <sup>53</sup> (prospective)	19 <sup>c</sup>	173	21	37	NR	32	72
PETHEMA-LAL3/97-GMALL+R <sup>54</sup> (prospective)	19 <sup>d</sup>	173	16	26 <sup>e</sup>	21	27	32
CODOX-M/IVAC <sup>55</sup> (retrospective)	8	84	13	88	NR	75	14
Modified CODOX-M/IVAC + R <sup>56</sup> (prospective)	22	84	0	32	5	9	36
Modified CODOX-M/IVAC(58) (retrospective)	30	84	10	20	NR	43	50
Modified CODOX-M/IVAC + R <sup>60</sup> (retrospective)	14 <sup>f</sup>	84	10	20	NR	NR	NR
Modified CODOX-M/IVAC + R <sup>57</sup> (retrospective)	14 <sup>f</sup>	84	0	50	NR	43	50
DA-EPOCH <sup>65</sup> (prospective)	39 <sup>g</sup>	126	0	13	NR	3	23
DA-EPOCH + R <sup>66</sup> (prospective)	23 <sup>h</sup>	126	0	16	NR	NR	NR
DA-EPOCH + R <sup>47</sup> Arm A (prospective)	51 <sup>i</sup>	126	9.8	27	NR	2	6
DA-EPOCH + R <sup>47</sup> Arm B (prospective)	55 <sup>l</sup>	189	7.3	29	NR	7	5
DA-EPOCH + R <sup>67</sup> (prospective)	29 <sup>m</sup>	126	NR	NR	NR	NR	NR
Short-term chemo-immunotherapy <sup>70</sup> (retrospective)	15	90	6.7	35	NR	NR	13.4



not be attributed exclusively to rituximab considering that supportive care was also optimised and HAART was used in all patients. Overall, infection rates were not affected by the addition of rituximab.<sup>53</sup> Importantly, there were no differences in TRM or in CR rates between HIV-negative and HIV-positive patients, showing that this schedule can safely be applied to HIV-positive patients. Results were reported together for patients with BL-HIV or mature B-cell acute lymphoblastic leukaemia (L3ALL), and schedules for these subgroups of patients were similar, but not identical. Since toxicities and outcome were not reported separately, conclusions may be misleading (Tables 3 and 4). This regimen is very long (173 days) and exhibits a high toxicity rate, with a TRM of ~20%. The use of methotrexate in every cycle resulted in increased incidence of mucositis.

CODOX-M/IVAC is the most investigated chemotherapy combination in HIV-BL patients (Table 2). It is currently with different modifications of dosage and schedule according to patient's age, co-morbidity and risk.<sup>55,24</sup> Dose adjustments, mostly of methotrexate and vincristine, were followed by an evident reduction in adverse events, with only grade 1-2 mucositis, neurotoxicity and TRM (Tables 2 and 3), and with a progressive immunological recovery and CD4+ counts.<sup>56-58</sup> CODOX-M/IVAC, combined or not with rituximab, has been assessed in a few, small series of patients with HIV-BL (Table 3 and 4), resulting in a TRM of ~15%, severe infections in ~65% of cases, mycosis infections in ~5%, and mucositis in 75%.<sup>56-60</sup> Improved outcome in recent studies can be explained by a better management of patients and also by a positive patient selection. In fact, the study population had a remarkably better immune status, with high CD4 counts (median: 375/ $\mu$ L), lower HIV viral load (median <50 copies/mL) and more limited stage of disease (only 50% had stage IV).<sup>56-58</sup> Recent studies demonstrated that HAART can be used during CODOX-M/IVAC, without additional toxicities, resulting in an acceptable TRM, similar to those reported in HIV-negative patients. Although the addition of rituximab in recent series improved outcome and prevented relapses, without an increase in infectious events (Table 3), cytopaenias, myelosuppression and nephrotoxicity remain important concerns in HIV-BL patients treated with CODOX-M/IVAC.

## Infusion Regimens

Infusion chemotherapy regimens exhibit a high activity in lymphoma patients, even when they were previously exposed to the cytostatics administered

as an intravenous bolus, suggesting a schedule-dependent effect in favour of the infusional administration of certain cytotoxic agents in patients with lymphoid neoplasms.<sup>61</sup> This was the rationale for the use of infusional regimens in the treatment of HIV-NHL. CDE regimen, consisting of a 96-hour continuous intravenous infusion of cyclophosphamide (800 mg/m<sup>2</sup>), doxorubicin (50 mg/m<sup>2</sup>), and etoposide (240 mg/m<sup>2</sup>) was the first assessed combination in HIV-BL, which was associated with a CRR of 45%, a 2-year FFS of 36% and 2-year OS of 43%, with significantly better tolerability and efficacy in patients treated in the HAART era.<sup>62</sup> Pooled results of three phase II trials addressing activity and tolerability of rituximab+CDE regimen have showed encouraging results, with a 70% CRR and a 2-year OS of 64%, but with increased risk for life-threatening infection (TRM= 8%).<sup>63</sup> These results were significantly poorer among patients with BL.

Another group has reported encouraging results with a 96-hour infusion regimen of doxorubicin, etoposide, and vincristine used in conjunction with intravenous bolus cyclophosphamide plus oral prednisone (EPOCH regimen) (Table 2). This regimen has been developed on the base of *in vitro* studies showing that tumour cells are relatively less resistant to prolonged low concentration exposure to the natural product-derived agents vincristine, doxorubicin, and etoposide, compared with brief higher concentration exposure.<sup>64</sup> In a first US National Cancer Institute (NCI) trial,<sup>65</sup> EPOCH therapy resulted in a 74% CRR and a 72% survival rate at a median follow-up of 53 months) in 39 patients with HIV-NHL (Table 3 and 4). The dose-adjustment strategy (DA-EPOCH) was implemented to reduce haematopoietic toxicity, and HAART was withheld during the study to avoid an increased risk of haematological toxicity with chemotherapy. HIV-BL patients enrolled were very few, and toxicities and subgroup analyses were not performed. Importantly, all deaths were related to CNS involvement, which was expected since DA-EPOCH does not include drugs that penetrate blood brain barrier, like methotrexate or cytarabine.

The addition of rituximab to DA-EPOCH (DA-EPOCH-R) has been associated with promising preliminary results in a prospective trial including 23 patients with BL (8 of them were HIV-positive), half of them advanced stage; at a median follow-up of 27 months, CR and OS rates were 100%, without toxic death.<sup>66</sup> A randomised phase II trial has demonstrated that concurrent rituximab plus infusion of EPOCH is

**Table 4. Activity and efficacy of more commonly used chemotherapy regimens.**

Rituximab (R), years (y), complete response (CR), 2-year-Progression Free Survival (2-y-PFS), 2-year-Overall-Survival (2-y-OS), Disease Free Survival (DFS), not reported (NR).

a) Percentuale relativa a pts in stadio III e IV.

b) This is 1-year-OS.

c) This is 3-years-EFS.

d) This is 3-years-OS.

e) Dati generali riguardanti l'intera popolazione (80 pts) non solo i 14 HIV-BL.

f) This is 3-years-PFS.

g) Median HIV viral load log<sub>10</sub>.

h) PFS and OS at 53 months.

Regimen	Median age y (range)	Stage IV (%)	Median CD4/ $\mu$ L	Median viral load copies/mL	Median follow-up (months)	CR (%)	2-y-PFS (%)	2-y-OS (%)
Hyper-CVAD <sup>49</sup> (prospective)	43 (32-55)	31	77	32,000	29	92	DFS:52	48
PETHEMA-LAL3/97-GMALL <sup>53</sup> (prospective)	41 (23-65)	57 <sup>a</sup>	420	400,000	31	68	DFS:71	46
CODOX-M/IVAC <sup>55</sup> (retrospective)	41 (19-61)	88	149	6,357	34	63	EFS:60	NR
PETHEMA-LAL3/97-GMALL+R <sup>54</sup> (prospective)	39 (29-54)	42 <sup>a</sup>	NR	NR	22	84	DFS:87	73
Modified CODOX-M/IVAC + R <sup>56</sup> (prospective)	40 (19-55)	NR	290	15,600	17	NR	NR	85.7 <sup>b</sup>
Modified CODOX-M/IVAC <sup>58</sup> (retrospective)	38 (28-69)	70 <sup>a</sup>	171	96,000	22	70	75 <sup>c</sup>	52 <sup>d</sup>
Modified CODOX-M/IVAC + R <sup>60</sup> (retrospective)	46 (17-78)	73 <sup>e</sup>	237	22,604	NR	93	68 <sup>f</sup>	68 <sup>d</sup>
Modified CODOX-M/IVAC + R <sup>57</sup> (retrospective)	46 (32-56)	50	375	<50	12	86	ND	83
DA-EPOCH <sup>65</sup> (prospective)	40 (31-57)	67 <sup>a</sup>	198	4.4 <sup>g</sup>	53	74	73 <sup>h</sup>	60 <sup>h</sup>
DA-EPOCH + R <sup>66</sup> (prospective)	31 (18-66)	52	NR	NR	27	100	100	100
DA-EPOCH + R <sup>47</sup> Arm A (prospective)	44	84 <sup>a</sup>	181	NR	30	63	66	70
DA-EPOCH + R <sup>47</sup> Arm B (prospective)	43	75 <sup>a</sup>	194	NR	30	82	63	67
DA-EPOCH + R <sup>67</sup> (prospective)	35 (16-88)	59 <sup>a</sup>	NR	NR	57	NR	NR	100
Short-term chemo-immunotherapy <sup>70</sup> (retrospective)	42 (27-63)	87 <sup>a</sup>	248	23.640	25	80	73	73

**Table 5. Short-term chemoimmunotherapy proposed by GICAT (Gruppo Italiano Cooperativo AIDS e Tumori).**

Cyclophosphamide (CTX), methotrexate (MTX), cytarabine (ara-C), vincristine (VCN), doxorubicin (DOXO), rituximab (R), etoposide (VP16), methylprednisolone (MP), intravenous (i.v.), therapy intra-thecal (TIT), carmustine (BCNU), melphalan (M), autologous stem cell transplant (ASCT).

<b>REGIMEN: Short-term chemoimmunotherapy<sup>70</sup></b>	
<b>Schedule</b>	<b>Therapeutic programme</b>
<p><b>Induction</b></p> <p>-2; -1 <b>MP</b> 0.5 - 1 mg/Kg/d i.v.</p> <p>0 <b>MP</b> 0.5 - 1 mg/Kg/d i.v.</p> <p><b>CTX</b> 500 mg/m<sup>2</sup> over 1 h infusion</p> <p><b>VCR</b> 2 mg total dose i.v. bolus</p> <p>1 <b>MP</b> 0.5 - 1 mg/kg/d i.v.</p> <p><b>CTX</b> 500 mg/m<sup>2</sup> over 1-h infusion</p> <p>2 <b>R</b> 375 mg/m<sup>2</sup></p> <p>5 <b>MTX</b> 12 mg + <b>Ara-C</b> 50 mg + steroids, i.t.</p> <p>7 <b>MTX</b> 3 g/m<sup>2</sup> i.v. over 6 h + leucovorin rescue</p> <p>14 <b>R</b> 375 mg/m<sup>2</sup></p> <p>15 <b>VP16</b> 250 mg/m<sup>2</sup> every 12 h</p> <p>19 <b>MTX</b> 12 mg + <b>Ara-C</b> 50 mg + steroids, i.t.</p> <p>21 <b>MTX</b> 3 g/m<sup>2</sup> i.v. over 6 h + leucovorin rescue</p> <p>29 <b>R</b> 375 mg/m<sup>2</sup></p> <p><b>DOXO</b> 50 mg/m<sup>2</sup> i.v. bolus</p> <p>33 <b>MTX</b> 12 mg + <b>Ara-C</b> 50 mg + steroids, i.t.</p> <p>36 <b>R</b> 375 mg/m<sup>2</sup></p> <p><b>VCN</b> 2 mg total dose i.v. bolus</p> <p><b>Consolidation</b></p> <p>50-51 <b>Ara-C</b> 2 g/m<sup>2</sup> in a 3-h infusion, twice a day (every 12 h)</p> <p>52 <b>R</b> 375 mg/m<sup>2</sup></p> <p>60 <b>R</b> 375 mg/m<sup>2</sup></p>	<p><b>After induction:</b></p> <p>If CR, high dose Ara-C and R based consolidation phase</p> <p>If PR, high dose Ara-C and R based consolidation phase followed by BEAM (BCNU, vp16, Ara-C, M) plus ASCT</p> <p>If SD/PD intensification phase, followed by BEAM plus ASCT</p> <p><b>At the end of CT</b></p> <p>If initial bulky disease or residual PET-positive single lesion were administered 36 Gy involved field irradiation</p>

associated with improved outcome with respect to sequential administration of rituximab and EPOCH in 106 patients with HIV-NHL.<sup>47</sup> This trial included only 27 HIV-BL patients, but activity in these patients was encouraging with both combinations, with a CRR of 63% in concurrent arm and 82% in sequential arm. Recently, the efficacy of DA-EPOCH-R was assessed in patients with *myc*-related aggressive-B-cell lymphomas; preliminary results of 29 patients with BL (10 HIV-positive) showed a 100% OS at a median follow-up of 57 months<sup>67</sup> (Table 3 and 4).

### Short-Term Chemoimmunotherapy

A dose-dense, short-term chemotherapy programme including seven active drugs and intrathecal drug delivery has showed excellent activity and safety profiles in HIV-negative patients with BL in the pre-rituximab era.<sup>68</sup> This regimen, proposed by the Gruppo Italiano Cooperativo AIDS e Tumori (GICAT), has been modified to be used with maintained efficacy and improved tolerability in HIV-BL. In

particular, six doses of rituximab have been added and methotrexate dose has been reduced from 150 and 250 mg/Kg to 3 g/m<sup>2</sup>, mostly to avoid mucositis, which constitutes an important route of access for infectious agents, and one of the main causes of death in these patients.<sup>69</sup> Treatment consists of a 36-day induction phase including sequential doses of fractionated cyclophosphamide, high doses of methotrexate and cytarabine, doxorubicin, vincristine, and etoposide, rituximab and intrathecal prophylaxis/treatment (Table 7). Subsequent treatment is tailored according to the objective response to induction phase: patients in CR are referred to high-dose cytarabine-based consolidation phase (Table 7); patients in partial response are referred to consolidation followed by BEAM plus autologous stem cell transplant; patients with stable or progressive disease are referred to intensification phase, followed by BEAM+ASCT. At the end of chemoimmunotherapy, patients with initial bulky disease or with a residual PET-positive

single lesion are evaluated for 36-Gy involved-field irradiation.

This modified chemoimmunotherapy regimen in 15 consecutive HIV-BL, with excellent safety profile and efficacy.<sup>70</sup> This intensive, short-term chemoimmunotherapy regimen is fast, safe, cost-effective, and active in HIV-BL, especially in patients responsive to HAART and with adequate CD4+ cell counts. It showed tolerability and efficacy similar to those reported with the original regimen in HIV-negative patients with BL,<sup>68</sup> and its activity and efficacy are similar to those attained with more demanding and resource consuming regimen in HIV-BL, with an apparently better tolerability profile (Tables 2 and 3). In fact, this program was delivered in a shorter period (median 90 days; range 52-143), without cases of mucositis, opportunistic infections and interruption due to toxicity, with manageable haematological toxicity, only mild infectious and a single toxic death. Autologous peripheral blood stem cell (APBSC) collection was successful in 9 out of 11 patients (median:  $14 \cdot 10^6$  CD34+ cells/kg). There was a single case of G4 non-haematological toxicity (transient diarrhoea). Patients with Hepatitis B virus (HBV) or Hepatitis C virus (HCV) positivity completed the planned treatment (ASCT in three), and experienced only transient G3 increase of transaminase serum level, without significant chemotherapy delay. CRR at the end of the whole treatment was 80%; at a median follow-up of 25 months (range 9-33), 11 patients remained disease-free, with a 2-year PFS and OS of 73%. CD4+ cell count  $\geq 200$  cells/ $\mu$ L is a favourable prognostic factor.<sup>70</sup> These encouraging results will be confirmed in an ongoing multicentre prospective phase II trial called the CARMEN trial (ClinicalTrials.gov Identifier: NCT01516593).

### Salvage Therapy and Role of Stem Cell Transplant

Standard salvage treatment for patients with relapsed or refractory HIV-BL remains to be defined as there are no studies focused on this issue, and recommendations are based on retrospective studies of HIV-associated lymphomas that include a few HIV-BL patients.<sup>71-74</sup> The first choice concerns the type of treatment, based upon patients' clinical conditions: supportive care, palliative chemotherapy or high-dose chemotherapy (HDC) plus autologous stem cell transplantation (ASCT).<sup>75</sup> The latter is considered the best curative option for HIV-negative patients with relapsed disease.<sup>76</sup> In the absence of significant differences, in terms of OS and PFS

between HIV negative and HIV positive patients, this schedule is considered to be feasible even in HIV-related lymphomas.<sup>72</sup> In most patients, data is based on the outcome after HDC/ASCT of the entire HIV-positive population, not exclusively on HIV-BL.<sup>71-74</sup> In other case series, HDC/ASCT led to very poor outcome in HIV-BL, sometimes due to inefficiency of induction chemotherapy,<sup>75</sup> or to early deaths after ASCT.<sup>77</sup> Ferreri et al. demonstrated a very promising outcome with BEAM + ASCT after induction therapy in patients who achieved CR, resulting in 5 CR beyond the 6 CR obtained with induction therapy alone.<sup>70</sup> There are no data on allo-SCT in relapsed HIV BL.<sup>78</sup>

## CONCLUSIONS

The introduction of HAART allowed the treatment of HIV-BL patients with the same intensive schedules proved to be curative in HIV-negative BL. Since there are no randomised trials comparing different first-line schedules (Table 4), gold standard regimen for HIV-BL is still debated. Available literature is mostly constituted by small retrospective and prospective studies considering patients with different lymphoma categories, other than HIV-BL. Thus, analysis and comparison of outcomes and toxicities is very difficult, potentially leading to unreliable conclusions. Adding rituximab on various schedules demonstrated good efficacy and tolerability compared to chemotherapy alone. The use of improved supportive therapy and antimicrobial prophylaxis significantly reduced adverse events, improving outcome. New de-escalated regimens could produce the same positive results obtained by more intensive and resource consuming combinations, with a lower risk of severe toxicity.<sup>70</sup> These innovative regimens should be assessed in prospective trials aimed also to identify prognostic markers to establish a risk-tailored overall strategy.



## REFERENCES

1. Jaffe ES. The 2008 WHO classification of lymphomas: implications for clinical practice and translational research. *Hematology Am Soc Hematol Educ Program*. 2009;(1):523-31.
2. Blinder VS, Chadburn A, Furman RR, Mathew S, Leonard JP. Improving outcomes for patients with Burkitt lymphoma and HIV. *AIDS Patient Care STDS*. 2008;22(3):175-87.
3. Stebbing J, Gazzard B, Mandalia S, Teague A, Waterston A, Marvin V et al. Antiretroviral treatment regimens and immune parameters in the prevention of systemic AIDS-related non-Hodgkin's lymphoma. *J Clin Oncol*. 2004;22(11):2177-83.
4. Polesel J, Clifford GM, Rickenbach M, Dal Maso L, Battegay M, Bouchardy C et al. Non-Hodgkin lymphoma incidence in the Swiss HIV Cohort Study before and after highly active antiretroviral therapy. *AIDS*. 2008;22(2):301-6.
5. Diamond C, Taylor TH, Aboumrad T, Anton-Culver H. Changes in acquired immunodeficiency syndrome-related non-Hodgkin lymphoma in the era of highly active antiretroviral therapy: incidence, presentation, treatment, and survival. *Cancer*. 2006;106(1):128-35.
6. Engels EA, Biggar RJ, Hall HI, Cross H, Crutchfield A, Finch JL et al. Cancer risk in people infected with human immunodeficiency virus in the United States. *Int J Cancer*. 2008;123(1):187-94.
7. Guech-Ongey M, Simard EP, Anderson WF, Engels EA, Bhatia K, Devesa SS et al. AIDS-related Burkitt lymphoma in the United States: what do age and CD4 lymphocyte patterns tell us about etiology and/or biology? *Blood*. 2010;116(25):5600-4.
8. Bertrand S, Berger R, Philip T, Bernheim A, Bryon PA, Bertoglio J et al. Variant translocation in a non endemic case of Burkitt's lymphoma: t (8;22) in an Epstein-Barr virus negative tumour and in a derived cell line. *Eur J Cancer*. 1981;17(5):577-84.
9. Egle A, Harris AW, Bouillet P, Cory S. Bim is a suppressor of Myc-induced mouse B cell leukemia. *Proc Natl Acad Sci USA*. 2004;101(16):6164-9.
10. Engels EA, Pfeiffer RM, Landgren O, Moore RD. Immunologic and virologic predictors of AIDS-related non-hodgkin lymphoma in the highly active antiretroviral therapy era. *J Acquir Immune Defic Syndr*. 2010;54(1):78-84.
11. Zoufaly A, Stellbrink HJ, Heiden MA, Kollan C, Hoffmann C, van Lunzen J et al. Cumulative HIV viremia during highly active antiretroviral therapy is a strong predictor of AIDS-related lymphoma. *J Infect Dis*. 2009;200(1):79-87.
12. Jarrett RF. Viruses and lymphoma/leukaemia. *J Pathol*. 2006;208(2):176-86.
13. Epeldegui M, Thapa DR, De la Cruz J, Kitchen S, Zack JA, Martinez-Maza O. CD40 ligand (CD154) incorporated into HIV virions induces activation-induced cytidine deaminase (AID) expression in human B-lymphocytes. *PLoS One*. 2010;5(7):e11448.
14. Herndier BG, Shiramizu BT, Jewett NE, Aldape KD, Reyes GR, McGrath MS. Acquired immunodeficiency syndrome-associated T-cell lymphoma: evidence for human immunodeficiency virus type 1-associated T-cell transformation. *Blood*. 1992;79(7):1768-74.
15. Bellan C, Lazzi S, De Falco G, Nyongo A, Giordano A, Leoncini L. Burkitt's lymphoma: new insights into molecular pathogenesis. *J Clin Pathol*. 2003;56(3):188-92.
16. Gabarre J, Raphael M, Lepage E, Martin A, Oksenhendler E, Xerri L et al. Human immunodeficiency virus-related lymphoma: relation between clinical features and histologic subtypes. *Am J Med*. 2001;111(9):704-11.
17. Smith SM. AIDS-related BL and CD4 count: a clue? *Blood*. 2010;116(25):5435-6.
18. Knowles DM. Etiology and pathogenesis of AIDS-related non-Hodgkin's lymphoma. *Hematol Oncol Clin North Am*. 2003;17(3):785-820.
19. Bellan C, Lazzi S, Hummel M, Palumbo N, de Santi M, Amato T et al. Immunoglobulin gene analysis reveals 2 distinct cells of origin for EBV-positive and EBV-negative Burkitt lymphomas. *Blood*. 2005;106(3):1031-6.
20. Young LS, Rickinson AB. Epstein-Barr virus: 40 years on. *Nat Rev Cancer*. 2004;4(10):757-68.
21. Gaidano G, Capello D, Carbone A. The molecular basis of acquired immunodeficiency syndrome-related lymphomagenesis. *Semin Oncol*. 2000;27(4):431-41.
22. Just PA, Fieschi C, Baillet G, Galicier L, Oksenhendler E, Moretti JL. 18F-fluorodeoxyglucose positron emission tomography/computed tomography in AIDS-related Burkitt lymphoma. *AIDS Patient Care STDS*. 2008;22(9):695-700.
23. Ostronoff M, Soussain C, Zambon E, Ibrahim A, Bosq J, Bayle C et al. Burkitt's lymphoma in adults: a retrospective study of 46 cases. *Nouv Rev Fr Hematol*. 1992;34(5):389-97.
24. Magrath I, Adde M, Shad A, Venzon D, Seibel N, Gootenberg J et al. Adults and children with small non-cleaved-cell lymphoma have a similar excellent outcome when treated with the same chemotherapy regimen. *J Clin Oncol*. 1996;14(3):925-34.
25. Bower M, Stebbing J. AIDS-associated malignancies. *Cancer Chemother Biol Response Modif*. 2005;22:687-706.
26. Levine AM. Acquired immunodeficiency syndrome-related lymphoma. *Blood*. 1992;80(1):8-20.
27. Sparano JA. Treatment of AIDS-related lymphomas. *Curr Opin Oncol*. 1995;7(5):442-9.
28. Sparano JA, Wiernik PH, Hu X, Sarta C, Schwartz EL, Soeiro R et al. Pilot trial of infusional cyclophosphamide, doxorubicin, and etoposide plus didanosine and filgrastim in patients with human immunodeficiency virus-associated non-Hodgkin's lymphoma. *J Clin Oncol*. 1996;14(11):3026-35.
29. Levine AM, Wernz JC, Kaplan L, Rodman N, Cohen P, Metroka C et al. Low-dose chemotherapy with central nervous system prophylaxis and zidovudine maintenance in AIDS-related lymphoma. A prospective multi-institutional trial. *JAMA*. 1991;266(1):84-8.
30. Kaplan LD, Straus DJ, Testa MA, Von Roenn J, Dezube BJ, Cooley TP et al. Low-dose compared with standard-dose m-BACOD chemotherapy for non-Hodgkin's lymphoma associated with human immunodeficiency virus infection. National Institute of Allergy and Infectious Diseases AIDS Clinical Trials Group. *N Engl J Med*. 1997;336(23):1641-8.
31. Gisselbrecht C, Oksenhendler E, Tirelli U, Lepage E, Gabarre J, Farcet JP et al. Human immunodeficiency virus-related lymphoma treatment with intensive combination chemotherapy. French-Italian Cooperative Group. *Am J Med*. 1993;95(2):188-96.
32. Mounier N, Spina M, Gabarre J, Raphael M, Rizzardini G, Golfier JB et al. AIDS-related non-Hodgkin lymphoma: final analysis of 485 patients treated with risk-adapted intensive chemotherapy. *Blood*. 2006;107(10):3832-40.
33. Bower M, Gazzard B, Mandalia S, Newsom-Davis T, Thirlwell C, Dhillon T et al. A prognostic index for systemic AIDS-related non-Hodgkin lymphoma treated in the era of highly active antiretroviral therapy. *Ann Intern Med*. 2005;143(4):265-73.
34. Hans CP, Weisenburger DD, Greiner TC, Gascoyne RD, Delabie J, Ott G et al. Confirmation of the molecular classification of diffuse large B-cell lymphoma by immunohistochemistry



- using a tissue microarray. *Blood*. 2004;103(1):275-82.
35. Lim ST, Karim R, Tulpule A, Nathwani BN, Levine AM. Prognostic factors in HIV-related diffuse large-cell lymphoma: before versus after highly active antiretroviral therapy. *J Clin Oncol*. 2005;23(33):8477-82.
36. Bower M, Collins S, Cottrill C, Cwynarski K, Montoto S, Nelson M et al. British HIV Association guidelines for HIV-associated malignancies 2008. *HIV Med*. 2008;9(6):336-88.
37. Kaplan LD. HIV-associated lymphoma. *Best Pract Res Clin Haematol*. 2012;25(1):101-17.
38. Ratner L, Lee J, Tang S, Redden D, Hamzeh F, Herndier B et al. Chemotherapy for human immunodeficiency virus-associated non-Hodgkin's lymphoma in combination with highly active antiretroviral therapy. *J Clin Oncol*. 2001;19(8):2171-8.
39. Little RF, Pittaluga S, Grant N, Steinberg SM, Kavlick MF, Mitsuya H et al. Highly effective treatment of acquired immunodeficiency syndrome-related lymphoma with dose-adjusted EPOCH: impact of antiretroviral therapy suspension and tumor biology. *Blood*. 2003;101(12):4653-9.
40. Dunleavy K, Little RF, Pittaluga S, Grant N, Wayne AS, Carrasquillo JA et al. The role of tumor histogenesis, FDG-PET, and short-course EPOCH with dose-dense rituximab (SC-EPOCH-RR) in HIV-associated diffuse large B-cell lymphoma. *Blood*. 2010;115(15):3017-24.
41. Bower M, McCall-Peat N, Ryan N, Davies L, Young AM, Gupta S et al. Protease inhibitors potentiate chemotherapy-induced neutropenia. *Blood*. 2004;104(9):2943-6.
42. Thomas DA, Faderl S, O'Brien S, Bueso-Ramos C, Cortes J, Garcia-Manero G et al. Chemoimmunotherapy with hyper-CVAD plus rituximab for the treatment of adult Burkitt and Burkitt-type lymphoma or acute lymphoblastic leukemia. *Cancer*. 2006;106(7):1569-80.
43. Kaplan LD, Lee JY, Ambinder RF, Sparano JA, Cesarman E, Chadburn A et al. Rituximab does not improve clinical outcome in a randomized phase 3 trial of CHOP with or without rituximab in patients with HIV-associated non-Hodgkin lymphoma: AIDS-Malignancies Consortium Trial 010. *Blood*. 2005;106(5):1538-43.
44. Ribera JM, Oriol A, Morgades M, Gonzalez-Barca E, Miralles P, Lopez-Guillermo A et al. Safety and efficacy of cyclophosphamide, adriamycin, vincristine, prednisone and rituximab in patients with human immunodeficiency virus-associated diffuse large B-cell lymphoma: results of a phase II trial. *Br J Haematol*. 2008;140(4):411-9.
45. Boue F, Gabarre J, Gisselbrecht C, Reynes J, Cheret A, Bonnet F et al. Phase II trial of CHOP plus rituximab in patients with HIV-associated non-Hodgkin's lymphoma. *J Clin Oncol*. 2006;24(25):4123-8.
46. Spina M, Jaeger U, Sparano JA, Talamini R, Simonelli C, Michieli M et al. Rituximab plus infusional cyclophosphamide, doxorubicin, and etoposide in HIV-associated non-Hodgkin lymphoma: pooled results from 3 phase 2 trials. *Blood*. 2005;105(5):1891-7.
47. Sparano JA, Lee JY, Kaplan LD, Levine AM, Ramos JC, Ambinder RF et al. Rituximab plus concurrent infusional EPOCH chemotherapy is highly effective in HIV-associated B-cell non-Hodgkin lymphoma. *Blood*. 2010;115(15):3008-16.
48. Boue F, Gabarre J, Gisselbrecht C, Reynes J, Cheret A, Bonnet F et al. Phase II trial of CHOP plus rituximab in patients with HIV-associated non-Hodgkin's lymphoma. *J Clin Oncol*. 2006;24(25):4123-8.
49. Cortes J, Thomas D, Rios A, Koller C, O'Brien S, Jeha S et al. Hyperfractionated cyclophosphamide, vincristine, doxorubicin, and dexamethasone and highly active antiretroviral therapy for patients with acquired immunodeficiency syndrome-related Burkitt lymphoma/leukemia. *Cancer*. 2002;94(5):1492-9.
50. Thomas DA, Cortes J, O'Brien S, Pierce S, Faderl S, Albitar M et al. Hyper-CVAD program in Burkitt's-type adult acute lymphoblastic leukemia. *J Clin Oncol*. 1999;17(8):2461-70.
51. Bowman WP, Shuster JJ, Cook B, Griffin T, Behm F, Pullen J et al. Improved survival for children with B-cell acute lymphoblastic leukemia and stage IV small noncleaved-cell lymphoma: a pediatric oncology group study. *J Clin Oncol*. 1996;14(4):1252-61.
52. Oriol A, Ribera JM, Esteve J, Sanz MA, Brunet S, Garcia-Boyer R et al. Lack of influence of human immunodeficiency virus infection status in the response to therapy and survival of adult patients with mature B-cell lymphoma or leukemia. Results of the PETHEMA-LAL3/97 study. *Haematologica*. 2003;88(4):445-53.
53. Oriol A, Ribera JM, Brunet S, del Potro E, Abella E, Esteve J. Highly active antiretroviral therapy and outcome of AIDS-related Burkitt's lymphoma or leukemia. Results of the PETHEMA-LAL3/97 study. *Haematologica*. 2005;90(7):990-2.
54. Oriol A, Ribera JM, Bergua J, Gimenez Mesa E, Grande C, Esteve J et al. High-dose chemotherapy and immunotherapy in adult Burkitt lymphoma: comparison of results in human immunodeficiency virus-infected and noninfected patients. *Cancer*. 2008;113(1):117-25.
55. Wang ES, Straus DJ, Teruya-Feldstein J, Qin J, Portlock C, Moskowitz C et al. Intensive chemotherapy with cyclophosphamide, doxorubicin, high-dose methotrexate/ifosfamide, etoposide, and high-dose cytarabine (CODOX-M/IVAC) for human immunodeficiency virus-associated Burkitt lymphoma. *Cancer*. 2003;98(6):1196-205.
56. Noy A, Kaplan L, Lee J. Feasibility and toxicity of a modified dose intensive R-CODOX-M/IVAC for HIV-associated Burkitt and atypical Burkitt lymphoma (BL): preliminary results of a prospective multicenter phase II trial of the AIDS Malignancy Consortium (AMC). *Blood*. (ASH Annual Meeting Abstracts) 2009;114:3673.
57. Rodrigo JA, Hicks LK, Cheung MC, Song KW, Ezzat H, Leger CS et al. HIV-Associated Burkitt Lymphoma: Good Efficacy and Tolerance of Intensive Chemotherapy Including CODOX-M/IVAC with or without Rituximab in the HAART Era. *Adv Hematol*. 2012;2012:735392.
58. Montoto S, Wilson J, Shaw K, Heath M, Wilson A, McNamara C et al. Excellent immunological recovery following CODOX-M/IVAC, an effective intensive chemotherapy for HIV-associated Burkitt's lymphoma. *AIDS*. 2010;24(6):851-6.
59. Mead GM, Barrans SL, Qian W, Walewski J, Radford JA, Wolf M et al. A prospective clinicopathologic study of dose-modified CODOX-M/IVAC in patients with sporadic Burkitt lymphoma defined using cytogenetic and immunophenotypic criteria (MRC/NCRI LY10 trial). *Blood*. 2008;112(6):2248-60.
60. Barnes JA, Lacasce AS, Feng Y, Toomey CE, Neuberg D, Michaelson JS et al. Evaluation of the addition of rituximab to CODOX-M/IVAC for Burkitt's lymphoma: a retrospective analysis. *Ann Oncol*. 2011;22(8):1859-64.
61. Sparano JA, Wiernik PH, Strack M, Leaf A, Becker N, Valentine ES. Infusional cyclophosphamide, doxorubicin, and etoposide in human immunodeficiency virus- and human T-cell leukemia virus type I-related non-Hodgkin's lymphoma: a highly active regimen. *Blood*. 1993;81(10):2810-5.
62. Sparano JA, Lee S, Chen MG, Nazeer T, Einzig A, Ambinder RF et al. Phase II trial of infusional cyclophosphamide, doxorubicin, and etoposide in patients with HIV-associated non-Hodgkin's lymphoma: an Eastern Cooperative Oncology Group Trial (E1494). *J Clin Oncol*. 2004;22(8):1491-1500.
63. Spina M, Jaeger U, Sparano JA, Talamini R, Simonelli C, Michieli M et al. Rituximab plus infusional cyclophosphamide, doxorubicin, and etoposide in HIV-associated non-Hodgkin

- lymphoma: pooled results from 3 phase 2 trials. *Blood*. 2005;105(5):1891-7.
64. Gutierrez M, Chabner BA, Pearson D, Steinberg SM, Jaffe ES, Cheson BD et al. Role of a doxorubicin-containing regimen in relapsed and resistant lymphomas: an 8-year follow-up study of EPOCH. *J Clin Oncol*. 2000;18(21):3633-42.
65. Little RF, Pittaluga S, Grant N, Steinberg SM, Kavlick MF, Mitsuya H et al. Highly effective treatment of acquired immunodeficiency syndrome-related lymphoma with dose-adjusted EPOCH: impact of antiretroviral therapy suspension and tumor biology. *Blood*. 2003;101(12):4653-9.
66. Dunleavy K, Little R, Pittaluga S, Grant N, Shovlin M, Steinberg S et al. A prospective study of dose-adjusted (DA) EPOCH with rituximab in adults with newly diagnosed Burkitt lymphoma: a regimen with high efficacy and low toxicity. *Annals of Oncology*. 2008;19(suppl.4):009.
67. Dunleavy K, Pittaluga S, Wayne S, Shovlin M, Johnson J, Little R et al. MYC+ aggressive lymphomas: novel therapy of untreated Burkitt lymphoma (BL) and MYC+ diffuse large B-cell lymphoma (DLBCL) with DA-EPOCH-R. *Annals of Oncology*. 2011;22:071.
68. Di Nicola M, Carlo-Stella C, Mariotti J, Devizzi L, Massimino M, Cabras A et al. High response rate and manageable toxicity with an intensive, short-term chemotherapy programme for Burkitt's lymphoma in adults. *Br J Haematol*. 2004;126(6):815-20.
69. Galicier L, Fieschi C, Borie R, Meignin V, Daniel MT, Gerard L et al. Intensive chemotherapy regimen (LMB86) for St Jude stage IV AIDS-related Burkitt lymphoma/leukemia: a prospective study. *Blood*. 2007;110(8):2846-54.
70. Ferreri AJ, Bruno Ventre M, Donadoni G, Cattaneo C, Fumagalli L, Foppoli M et al. Safety and activity of a new intensive short-term chemoimmunotherapy in HIV-positive patients with Burkitt lymphoma. *Br J Haematol*. 2012;159(2):252-5.
71. Balsalobre P, Diez-Martin JL, Re A, Michieli M, Ribera JM, Canals C et al. Autologous stem-cell transplantation in patients with HIV-related lymphoma. *J Clin Oncol*. 2009;27(13):2192-8.
72. Diez-Martin JL, Balsalobre P, Re A, Michieli M, Ribera JM, Canals C et al. Comparable survival between HIV+ and HIV- non-Hodgkin and Hodgkin lymphoma patients undergoing autologous peripheral blood stem cell transplantation. *Blood*. 2009;113(23):6011-4.
73. Krishnan A, Molina A, Zaia J, Smith D, Vasquez D, Kogut N et al. Durable remissions with autologous stem cell transplantation for high-risk HIV-associated lymphomas. *Blood*. 2005;105(2):874-8.
74. Re A, Michieli M, Casari S, Allione B, Cattaneo C, Rupolo M et al. High-dose therapy and autologous peripheral blood stem cell transplantation as salvage treatment for AIDS-related lymphoma: long-term results of the Italian Cooperative Group on AIDS and Tumors (GICAT) study with analysis of prognostic factors. *Blood*. 2009;114(7):1306-13.
75. Bayraktar UD, Ramos JC, Petrich A, Gupta N, Lensing S, Moore PC et al. Outcome of patients with relapsed/refractory acquired immune deficiency syndrome-related lymphoma diagnosed 1999-2008 and treated with curative intent in the AIDS Malignancy Consortium. *Leuk Lymphoma*. 2012;53(12):2383-9.
76. Philip T, Guglielmi C, Hagenbeek A, Somers R, Van der Lelie H, Bron D et al. Autologous bone marrow transplantation as compared with salvage chemotherapy in relapses of chemotherapy-sensitive non-Hodgkin's lymphoma. *N Engl J Med*. 1995;333(23):1540-5.
77. Gabarre J, Marcelin AG, Azar N, Choquet S, Levy V, Levy Y et al. High-dose therapy plus autologous hematopoietic stem cell transplantation for human immunodeficiency virus (HIV)-related lymphoma: results and impact on HIV disease. *Haematologica*. 2004;89(9):1100-8.
78. Gupta V, Tomblyn M, Pedersen TL, Atkins HL, Battiwalla M, Gress RE et al. Allogeneic hematopoietic cell transplantation in human immunodeficiency virus-positive patients with hematologic disorders: a report from the center for international blood and marrow transplant research. *Biol Blood Marrow Transplant*. 2009;15(7):864-71.
79. Swerdlow SH, Campo E, Harris NL, Pileri S et al. WHO classification of tumors of Haematopoietic and Lymphoid Tissues. (2008), France: IARC Press.
80. Rodig SJ, Vergilio JA, Shahsafaei A, Dorfman DM. Characteristic expression patterns of TCL1, CD38, and CD44 identify aggressive lymphomas harboring a MYC translocation. *Am J Surg Pathol*. 2008;32(1):113-22.
81. Guikema JE, de Boer C, Haralambieva E, Smit LA, van Noesel CJ, Schuurin E et al. IGH switch breakpoints in Burkitt lymphoma: exclusive involvement of noncanonical class switch recombination. *Genes Chromosomes Cancer*. 2006;45(9):808-19.
82. Lovisa F, Mussolin L, Corral L, Pillon M, Cazzaniga G, Biondi A et al. IGH and IGK gene rearrangements as PCR targets for pediatric Burkitt's lymphoma and mature B-ALL MRD analysis. *Lab Invest*. 2009;89(10):1182-6.
83. Moormeier JA, Williams SF, Golomb HM. The staging of non-Hodgkin's lymphomas. *Semin Oncol*. 1990;17(1):43-50.
84. Murphy SB. Classification, staging and end results of treatment of childhood non-Hodgkin's lymphomas: dissimilarities from lymphomas in adults. *Semin Oncol*. 1980;7(3):332-9.
85. Cheson BD. Role of functional imaging in the management of lymphoma. *J Clin Oncol*. 2011;29(14):1844-54.
86. Brust D, Polis M, Davey R, Hahn B, Bacharach S, Whatley M et al. Fluorodeoxyglucose imaging in healthy subjects with HIV infection: impact of disease stage and therapy on pattern of nodal activation. *AIDS*. 2006;20(7):985-93.