# TARGETING MAST CELLS AS A VIABLE THERAPEUTIC OPTION IN ENDOMETRIOSIS

# \*David A. Hart

Department of Surgery, McCaig Institute, University of Calgary, Calgary, Alberta, Canada. \*Correspondence to hartd@ucalgary.ca

**Disclosure:** The author has declared no conflicts of interest.

**Acknowledgements:** The author thanks the Institute for Gender and Health of the Canadian Institutes for Health Research for past funding, Alberta Health Services for current support, and Dr Kevin A. Hildebrand for his critical review of the manuscript. **Received:** 03.03.17 **Accepted:** 05.07.17

**Citation:** EMJ Repro Health. 2017;3[1]:76-83.

# ABSTRACT

Endometriosis is a chronic condition that affects ~10% of young women worldwide. Pain and infertility are the two most common features of the disease. The condition appears to be sex hormone-dependent, although a subset of females with the condition still experience symptoms post-menopause. The aetiology of endometriosis induction still remains elusive, and surgery to remove the lesions often fails to cure the condition, as the lesions often reappear. The lesions contain stromal cells, blood vessels, nerves, and numerous mast cells. In some respects, endometrial lesions resemble a chronic fibrotic scar-like tissue that does not resolve. Studies in other fibrotic abnormal healing conditions have revealed that targeting mast cells, as a central component of what is called a 'neural-mast cell-fibroblast' axis, by repurposing asthma drugs can prevent induction of the abnormal healing phenotype. Given the similarities between conditions with abnormal healing phenotypes and endometrial lesions, it is postulated that taking a similar approach to target endometrial lesion mast cells could exert a benefit for patients with endometriosis. This review also outlines approaches to assess the likelihood that targeting mast cells could lead to clinical trials using such 'repurposed' mast cell targeted drugs.

<u>Keywords:</u> Mast cells, mast cell stabilisers, endometriosis, cell biology of endometriosis, neuroinflammatory pathways, biochemistry of endometriosis.

## PREAMBLE

This review is intended to focus on the rationale and feasibility of targeting mast cells in endometriosis, and discussing selected findings in the literature related to this focus. Due to space limitations, it is not intended to be an exhaustive review of all aspects of endometriosis. Thus, the reader is encouraged to read the large number of recent reviews in the literature which address other aspects of this complex disease to complement the discussion below.

## THE CLINICAL PROBLEM

Endometriosis is an inflammatory condition that affects ~10% of young women of reproductive age.<sup>1</sup> The disease appears to be sex hormone-dependent as symptoms can vary across the menstrual cycle, with pain as a prominent symptom. The condition is also frequently accompanied by dysmenorrhoea and infertility, with the presence of pelvic abdominal lesions. Conservative treatment with anti-inflammatory drugs is often ineffective, and surgical removal of the lesions becomes a viable alternative.<sup>2</sup> However, after surgery, the condition can reappear with pain and associated lesions. As the condition is chronic, it can be accompanied by epigenetic changes that may complicate the effectiveness of treatment.<sup>3,4</sup>

The disease is characterised by the growth of endometrial elements outside of the uterine cavity. Why this occurs is currently unknown, and what factors predispose or contribute to this very common condition in a subpopulation of younger females is not yet evident. It may result from retrograde menstruation,<sup>5,6</sup> with attachment and growth of endometrial tissue in this environment. The tissue associated with endometriosis contains blood vessels, nerves, mast cells, myofibroblasts, and macrophages,<sup>7-10</sup> and, thus, appears to be a fibrotic lesion that in some respects resembles a progressive scar. As endometriosis tissue can be influenced by sex hormones,<sup>11-13</sup> and possibly neuroinflammatory elements,14 it may resemble abnormal scarring such as occurs following a burn injury (hypertrophic scar), or following a traumatic elbow injury leading to a joint contracture.<sup>15,16</sup> Thus, the initiating events may differ but the cellular and molecular interactions leading to fibrotic progression may share a number of commonalities. Interestingly, endometriosis has also been linked with risk for other chronic diseases such as asthma,<sup>17,18</sup> in which mast cells also play a central role.

## PRECLINICAL MODELS

Endometriosis can occur naturally in horses<sup>19</sup> and non-human primates,<sup>20</sup> and can be induced in rats by auto-transplantation of uterine tissue to the abdomen, or in nude mice by transplantation of human endometrial tissue.<sup>21,22</sup> Such models can provide some insights into aspects of endometriosis development and progression, as well as insights into potential interventions, but, thus far, the aetiology of endometriosis in humans and in non-human primates remains undefined.

## ROLE OF SEX HORMONES IN ENDOMETRIOSIS

As discussed above, endometriosis is a sex hormone-mediated inflammatory disease. With regard to endometriosis, oestrogen has been implicated in the activity of macrophages,<sup>10</sup> fibroblasts and myofibroblasts,<sup>13</sup> nerves,<sup>10,12</sup> and mast cells.<sup>7,23</sup> It is also clear that oestrogen is a regulator of inflammatory processes, and activation of inflammatory cells can contribute to nerve fibre-mediated pain.<sup>14</sup> In addition, sex hormones, such as oestrogen, are known to contribute to wound healing in response to injury.<sup>24,25</sup> Both oestrogen and progesterone have been implicated in mast cell degranulation.<sup>26</sup>

After menopause, inflammatory processes decline, wound healing is compromised, and endometriosis in most patients becomes latent.<sup>5,27</sup> However, 2–4% of women still continue to experience endometriosis after menopause.<sup>5</sup> Therefore, either alternative mechanisms could potentially allow the disease to continue in this subpopulation (e.g. due to epigenetic modifications<sup>4,28</sup>) or local production of oestrogen could continue to contribute to disease activity in the post-menopausal state.

While endometriosis is considered oestrogendependent, in part based on menstrual cycle dependency, a role for progesterone is also likely.<sup>29</sup> Furthermore, the impact of sex hormones could be a secondary sequela of hormone level changes (e.g. water retention with increased turgor of endometrial lesions contributing to pain).

# REGULATION OF PROTEINASE EXPRESSION IN ENDOMETRIOSIS

## **Role of Sex Hormones and Their Receptors**

Proteinases such as tissue factor,<sup>30</sup> elements of the fibrinolytic system,<sup>31</sup> cathepsins,<sup>32</sup> mast cell tryptase and chymase,<sup>33</sup> and matrix metalloproteinases (MMP)<sup>34</sup> have all been implicated in endometriosis. As these enzymes can facilitate extracellular matrix turnover, activate pro-enzymes and other molecules, and serve other functions related to inflammation, coagulation, collagen deposition, and angiogenesis, they are likely central to endometriosis progression. In this regard, their role is likely not different from their roles in other fibrogenic processes and wound healing.

Much research has focussed on the MMP, their expression in endometriosis, and their regulation by progesterone<sup>35,36</sup> and oestrogen.<sup>34</sup> Progesterone appears to limit MMP expression,<sup>35,36</sup> while oestrogen is reported to enhance MMP-9 expression,<sup>37</sup> and MMP-9 levels are enhanced in endometriosis.<sup>38</sup> As oestrogen functions primarily via estrogen receptor (ER)-alpha and ER-beta in cells, receptors known to be expressed in endometrial tissue,<sup>39,40</sup> this response is potentially contributing to the symptoms of endometriosis and its progression. However, the role of oestrogen receptors in endometriosis is somewhat controversial,<sup>39,40</sup> with ER-beta a more prominent variant in endometriosis than normal tissue.<sup>39</sup>

Interestingly, oestrogen receptors in the absence of oestrogen can also regulate expression of some MMP<sup>41-43</sup> and these include MMP-1 and MMP-13. The addition of oestrogen actually depressed expression rather than enhancing expression. ER-beta was more effective than ER-alpha in many of these responses, and, interestingly, genetic variants of the *MMP-1* promoter region were also differentially regulated. Some of these same variants, as well as those for *MMP-2*, *7*, and *9* were found to be associated with risk of endometriosis.<sup>44</sup> Intriguingly, splice variants of ER-beta that do not bind oestrogen were also effective in upregulating expression of *MMP-1*,<sup>45</sup> and splice variants have also been detected in endometriosis tissues.<sup>46</sup>

## MAST CELLS IN ENDOMETRIOSIS

Mast cells, often thought of in the context of allergies or asthma, are present in most tissues throughout the body, including connective tissues and many organs.<sup>7,15,16</sup> Mast cells are also very prevalent in normal and abnormal healing environments, with elevated numbers, in abnormal healing conditions.<sup>7,15,16</sup> Mast cells are very prevalent in endometriosis tissue, and many of them appear to be activated and degranulated.47-49 In addition, many of the mast cells were localised very close to neural elements,<sup>49</sup> and, as such, could play an active role in neuroinflammatory processes.7,15,16 We have previously seen neural elements ending in normal dense connective tissues very close to a mast cell, implying that nerve-mast cell co-localisation may also play a role in normal tissue functioning as well.7,15,16

Mast cells and their products could contribute to several features of endometriosis. Mast cell tryptase can activate protease activated receptors (PAR), particularly PAR-2.50 PAR are known to be expressed in endometrium and endometriosis.<sup>51</sup> Activation of PAR-2 on cells may participate in pain processing<sup>52</sup> and angiogenesis.<sup>53</sup> Mast cell tryptase can also activate myofibroblasts and contribute to fibrosis.<sup>15,16</sup> Histamine released from mast cells has been studied for decades and has multiple activities, including enhancing tissue oedema and many other effects. Activated mast cells can also release a number of pro-inflammatory cytokines, mediators, and growth factors, and thus, can have a very potent and varied impact on a target tissue, particularly one that appears to be abnormally regulated, as in endometriosis. The enhanced presence of mast cells, many of which appear to be activated, in endometriosis tissue has led to the proposal that mast cells should be targeted with drug interventions to assist in controlling endometriosis progression and symptoms.7,23,54,55 Based on the findings that activated mast cells are present in endometrial tissues, oestrogen/progesterone can influence mast cells, mast cells can release many biologically active molecules, and mast cell numbers

are increased in endometrial tissues, this cell may be an excellent cell to target in endometriosis, alone or in combination with other targets.

#### MAST CELL STABILISERS

Mast cell stabilisers are drugs which inhibit or prevent mast cell degranulation. These include two drugs, ketotifen and sodium cromoglycate, that have a long history of use in the treatment of asthma. While ketotifen is now off patent protection, it has been used safely for >40 years in adult and paediatric populations. It is not totally targeted specifically for mast cells, and is also reported to inhibit degranulation of polymorphonuclear leukocytes.<sup>17,18</sup> Thus, with the development of new potential indications (e.g. endometriosis), there may be an impetus for industry to develop newer versions that may be more specific for mast cells, or more active in specific diseases or conditions.

## USE OF MAST CELL STABILISERS IN ABNORMAL FIBROPROLIFERATIVE CONDITIONS

Abnormal healing follows induction of skin injuries in the red Duroc pig<sup>56,57</sup> and following trauma and immobilisation of a knee injury in a rabbit model.<sup>58</sup> The former exhibits characteristics of hypertrophic scarring and also fibrogenic scarring.<sup>59</sup> In the red Duroc pig, this abnormal scarring response appears to have a genetic component.<sup>60</sup> The rabbit model exhibits cellular and molecular characteristics of joint contractures that occur in a subset of humans who experience an elbow injury.<sup>15,16</sup>

In these two models, elevated numbers of mast cells, nerves, and myofibroblasts have been observed leading to the conclusion that the cells contribute to fibroproliferative dysfunction. These cells have been postulated to form a cellular 'axis' (Figure 1) in which the mast cell plays a central role.<sup>7,15,16</sup> In this proposed axis, the fibroblasts and myofibroblasts are the effector cells which release fibrotic molecules, such as collagens, and contract the fibrotic matrix. Neuropeptides from nerves or, in the case of endometriosis, possibly sex hormones, impact mast cells, which then release molecules that enhance the activity of fibroblasts and myofibroblasts. Thus, the mast cells function as accelerators or amplifiers of the fibrotic environment. This axis is postulated to be

dysfunctional in the two models, but whether the initiator of the dysfunction is only due to the nerves (e.g. neuroinflammation), or some other cell type or stimulus, remains to be determined. Treatment of skin wounds on red Duroc pigs with the asthma drug ketotifen from the time of injury prevented development of the abnormal fibrogenic response to injury and led to a more typical healing response.<sup>57</sup>

If treatment was delayed until 28 days post-injury, the drug was without effect. Stopping the drug treatment after epithelisation of the wounds did not lead to a reactivation of the abnormal fibrogenic response. Thus, this mast cell stabiliser appeared to exert its influence early in the response to injury. Interestingly, treatment with ketotifen led to a decline in detectable nerves, mast cells, and myofibroblasts in the skin scar tissue, which likely also indicated that the postulated axis was not unidirectional and interfering with mast cell degranulation impacted other elements of this axis. Finally, treatment of skin wounds with ketotifen in Yorkshire pigs that healed normally was without effect and did not influence the number of cells in the axis that could be detected. Thus, the drug did not appear to influence normal healing of skin wounds.



#### Figure 1: Mast cells mediated inflammation and fibrosis.

Mast cells circulate as CD34-positive precursor cells and terminally differentiate in connective tissues. Both IgE-dependent and independent mechanisms can activate mast cells causing the release of preformed and newly synthesised pro-inflammatory mediators. Many of these mediators increase vascular permeability and promote the recruitment of other inflammatory cells and additional mast cell precursors. SCF is also secreted by activated fibroblasts and myofibroblasts, further potentiating mast cell recruitment and proliferation. TGF- $\beta$  is a potent fibroblast mitogen and stimulator of myofibroblast differentiation. It also impedes myofibroblast apoptosis.

bFGF: basic fibroblast growth factor; CGRP: calcitonin gene-related peptide; CTGF: connective tissue growth factor; IgE: immunoglobulin E; NGF: nerve growth factor; PDGF: platelet-derived growth factor; SCF: stem cell factor; TGF-β: transforming growth factor beta; TNF-a: tumor necrosis factor alpha; VIP: vasoactive intestinal peptide.

Adapted from Monument et al.<sup>16</sup>

#### Table 1: Potential clinical pathway to confirming mast cells as a therapeutic target in endometriosis.

#### Phase 1: Amass evidence for mast cell activation in patients with endometriosis

Assess mast cell tryptase in serum of age matched individuals with and without established endometriosis at three defined points during their menstrual cycle using a validated ELISA.<sup>63</sup> If indeed mast cell degranulation is involved in endometriosis activity and symptoms across the menstrual cycle, the ELISA results should parallel symptoms.

As surgery is recommended for many patients with endometriosis, one could assess serum mast cell tryptase levels before and post-surgery during specific points in the menstrual cycle to address the question of whether serum tryptase levels can be used as a biomarker for a return of the endometriosis.

Histologic assessment of the nerve-mast cell-myofibroblast axis (Figure 1) in tissue obtained at the time of surgery, using immunolocalisation protocols developed for assessing human joint contracture tissues.<sup>58</sup>

*In vitro* explant studies using tissue obtained at the time of surgery. Such tissues could be incubated plus or minus ketotifen and supernatant levels of tryptase assessed using an ELISA.<sup>63</sup> A search of the literature did not reveal whether such surgery is preferentially performed at specific times in the menstrual cycle of patients, but certainly this may impact the results of the studies outlined.

Phase 2: Short-term ketotifen trial in endometriosis patients scheduled for surgery

Patients would be randomly assigned to a ketotifen arm or a sham solution arm of the protocol (oral dosing of ketotifen or sham solution) in a blinded fashion. Daily diaries for symptoms would be maintained for the 3 months prior to surgery, as well as serum levels for mast cell tryptase (blood draws once per month at the optimal timing). Tissue obtained at the time of subsequent surgery could be assessed for nerves, mast cells, myofibroblasts, and macrophage subsets, by immunolocalisation techniques, as well as RNA isolated from the tissue by RT-qPCR, and incubation of culture supernatants for detection of relevant proteins by established array technologies.

**Phase 3: Large scale clinical trials of ketotifen (single-centre and multicentre)** These trials could take many designs, optimal designs determined by clinical experts to yield the most information and maintain patient safety.

Ketotifen pilot trial in patients: pain assessments (diary) for 3 months prior to drug or sham initiation and then daily for 3 months. Washout for 3 months and then cross over for 3 months.

Ketotifen implementation trial: Cohort of patients to receive ketotifen or sham for 12 months and then stop and follow for a defined period of time to determine whether any improvements revert following cessation of treatment.

Large-scale RCTs: Double-blind multicentre trials

ELISA: enzyme-linked immunosorbent assay; RCT: randomised controlled trial; RT-qPCR: quantitative reverse transcription polymerase chain reaction.

Similar studies in the rabbit model of post-traumatic joint contractures using ketotifen treatment also led to improvement in the healing process and significant declines in the extent of the joint contractures (~50%), as well as cells of the nervemast cell-myofibroblast axis.<sup>61,62</sup> The suggestion that ketotifen was likely inhibiting mast cell degranulation, arose from recent studies demonstrating that serum levels of mast cell tryptase were significantly depressed in the treated rabbits.<sup>63</sup>

The mast cell stabiliser sodium cromoglycate has also shown effectiveness in a rat model of experimental endometriosis.<sup>55</sup> In this model, transplantation of endometrial tissue to the abdominal wall followed by a 2-week treatment regimen with sodium cromoglycate led to significant declines in activated mast cells, as well as tissue levels of mast cell tryptase and serum levels of tumour necrosis factor alpha. However, the size of the lesions was not apparently influenced by

the treatment. As myofibroblasts have also been identified in endometriosis lesions,<sup>9</sup> as well as nerves,<sup>49</sup> it would be of interest to assess the influence of mast cell stabiliser treatment of the transplanted tissues on levels of these two cells as well, particularly since mast cells and nerve elements have been detected in close proximity in human endometriosis tissues.<sup>49</sup> Interestingly, close proximity of nerves and mast cells have been noted in unrelated tissues, and it appeared that neuropeptides released from nerves could impact the mast cells (Figure 1), leading to an amplified impact on the target tissue.

From our studies using ketotifen,<sup>57,61,62</sup> and those of Zhu et al.<sup>55</sup> with sodium cromoglycate, inhibition of mast cell degranulation may be a viable direction to explore. In contrast, D'Cruz and Uckun<sup>54</sup> proposed using a janus kinase (JNK) 3 inhibitor, JANEX-1, and recently a JNK inhibitor has been reported to cause regression of endometriotic lesions in rodent models.<sup>64</sup> While targeting mast cells using these approaches has not yet led to clinical trials in endometriosis, a clinical trial is currently underway using ketotifen to prevent joint contracture development following traumatic elbow injuries.<sup>65</sup>

## Are There Commonalities in Dysregulated Fibrogenic Responses to Inflammation and Injuries?

Joint contracture development following a trauma to the elbow occurs in 10–15% of those injured.<sup>15,16</sup> Development of a contracture is accompanied by a fibrotic response in which the healing process is dysregulated and does not proceed through to maturation and remodelling of the scar. In many cases, the patients undergo surgery to release the contracture, but often with only partial recovery.<sup>15,16</sup>

Similarly, development of a hypertrophic scar after a severe burn injury is also a fibrotic response to the injury, with excessive matrix deposition, myofibroblasts, and mast cells. For many patients, the hypertrophic scars have to be surgically removed. Interestingly, female sex is also a risk factor for hypertrophic scar development.<sup>66</sup> The abnormal fibrogenic skin wound healing, in the previously discussed porcine models, also exhibits some similarities with hypertrophic scarring.

Given these similarities, perhaps much of the uniqueness of endometriosis is associated with where the abnormal fibrotic tissue is located, its sex hormone dependency, and the sequelae of the fibrosis, rather than any intrinsic uniqueness in the cellular aspects of the dysregulated processes contributing to progression and chronicity. Thus, some of what has been learned from these other diseases or conditions, as well as from the preclinical models discussed, could provide insights into focussing research directions forward with mast cells as a target.

## Future Directions for Assessment of Efficacy and Implementation of Mast Cell Stabilisers in Patients with Endometriosis

To undertake a clinical trial of mast cell stabilisers, such as ketotifen or sodium cromoglycate, in populations of young females with endometriosis, investigators must proceed with caution as many of these individuals are of reproductive age, and mast cells have been implicated in normal ovulation events and others related to reproduction in some species.<sup>67</sup> However, the complexity of endometriosis in patient populations could be approached in

phases to strengthen the link between mast cell activity and symptoms and then identify those best suited to participate in pilot trials using the mast cell stabilisers (Table 1).

One of the current gaps in endometriosis research is a lack of good biomarkers, such as serum components, that can be used to monitor disease activity, assess the impact of interventions, or assess the return of disease activity following surgery prior to overt symptoms being evident. Serum mast cell tryptase levels, as assessed by an enzyme-linked immunosorbent assay<sup>63</sup> could contribute to filling this gap.

Finally, it is clear that epigenetic changes occur during progression of endometriosis.<sup>3,4,28</sup> These changes (DNA methylation, histone modifications, and alterations to miRNA profiles) can lead to alterations in cell responsiveness to interventions. In some chronic diseases (e.g. rheumatoid arthritis), these changes can occur in fibroblasts and other cells. Thus, endometrial lesions early in the disease may respond differently to targeting mast cells than those with more advanced disease. Such factors may need to be considered when assessing effectiveness.

#### CONCLUSIONS

Based on the above discussion, there is considerable circumstantial evidence to support the use of mast cell targeted drug interventions in the treatment of endometriosis. There is an advantage for using known drugs such as ketotifen and sodium cromoglycate, both with long track records of use for the treatment of asthma in a variety of populations. Thus, their safety and efficacy is well documented. However, repurposing these drugs is a viable approach, but one that will require a systematic analysis in patient populations (Table 1). Given the large population of females with endometriosis (5-15% of females worldwide), and the impact of the condition on these young women, new approaches that could impact their quality of life should be entertained. However, this approach using mast cell targeted drugs may address the how of the disease, but not why the disease occurs, so it would only be a stop-gap approach until new information arises as to why it develops, and what is unique about the subset of women who experience the disease (e.g. genetics, epigenetics, exposure to environmental stimuli, and stochastic events).

#### REFERENCES

1. Greene AD et al. Endometriosis: where are we and where are we going? Reproduction. 2016;152(3):R63-78.

2. Bedaiwy MA et al. New developments in the medical treatment of endometriosis. Fertil Steril. 2017;107(3):555-65.

3. Borghese B et al. Recent insights on the genetics and epigenetics of endometriosis. Clin Genet. 2017;91(2): 254-64.

4. Koukoura O et al. DNA methylation in endometriosis (Review). Mol Med Res. 2016;13(4):2939-48.

5. Bendon CL, Becker CM. Potential mechanisms of postmenopausal endometriosis. Maturitas. 2012;72(3): 214-9.

6 Vercellini P et al. Endometriosis: pathogenesis and treatment. Nat Rev Endocrinol. 2014;10(4):261-75.

7. Hart DA. Curbing inflammation in multiple sclerosis and endometriosis: should mast cells be targeted? Int J Inflam. 2015;2015:452095.

8. McKinnon B et al. Endometriosisassociated nerve fibers, peritoneal fluid cytokine concentrations, and pain in endometriotic lesions from different locations. Fertil Steril. 2012;97(2):373-80.

9. Ibrahim MG et al. Abdominal wall endometriosis: myofibroblasts as a possible evidence of metaplasia: a case report. Gynecol Obstet Invest. 2017;82(1): 90-101.

10. Greaves E et al. Estradiol is a critical mediator of macrophage-nerve cross talk in peritoneal endometriosis. Am J Pathol. 2015;185(8):2286-97.

11. Bedaiwy MA et al. Medical Management of endometriosis in patients with chronic pelvic pain. Semin Reprod Med. 2017;35(1):38-53.

12. Liang Y, Yao S. Potential role of estrogen in maintaining the imbalanced sympathetic and senory innervation in endometriosis. Mol Cell Endocrinol. 2016; 424:42-9.

13. Da Costa e Silva Rde C et al. Estrogen signaling in the proliferative endometrium: implications in endometriosis. Rev Assoc Med Bras. 2016; 62(1):72-7.

14. McKinnon BD et al. Inflammation and nerve fiber interaction in endometriotic pain. Trends Endocrinol Metab. 2015; 26(1):1-10.

15. Monument MJ et al. Posttraumatic elbow contractures: targeting neuroinflammatory fibrogenic mechanisms. J Orthop Sci. 2013;18(6): 869-77.

16. Monument MJ et al. Neuroinflammatory

mechanisms of connective tissue fibrosis: targeting neurogenic and mast cell contributions. Adv Wound Care. 2015; 4(3):137-51.

17. Kvaskoff M et al. Endometriosis: a high-risk population for major chronic diseases? Hum Reprod Update. 2015: 21(4):500-16.

18. Fourquet J et al. Characteristics of women with endometriosis from the USA and Puerto Rico. J Endometr Pelvic Pain Disord. 2015:7(4):129-35.

19. Rebordao MR et al. Physiopathologic mechanisms involved in mare endometriosis. Reprod Domest Anim. 2014;49(suppl 4):82-7.

20. Yamanaka A et al. Primate model research for endometriosis. Tohoku J Exp Med. 2012;226(2):95-9.

21. Grummer R. Animal models in endometriosis research. Hum Reprod Update. 2006;12(5):641-9.

22. Tirado-Gonzalez I et al. Endometriosis research: animal models for the study of a complex disease. J Reprod Immunol. 2010;86(2):141-7.

23. Binda MM et al. Targeting mast cells: a new way to treat endometriosis. Expert Opin Ther Targets. 2017;21(1):67-75.

24. Ashcroft GS, Ashworth JJ. Potential role of estrogens in wound healing. Am J Clin Dermatol. 2003;4(11):737-43.

25. Gilliver SC, Ashcroft GS. Sex steroids and cutaneous wound healing: the contrasting influences of estrogens and androgens. Climacteric. 2007;10(4): 276-88.

26. Zierau O et al. Role of female sex hormones, estradiol and progesterone, in mast cell behavior. Front Immunol. 2012: 3:169.

27. Inceboz U. Endometriosis after menopause. Women's Health. 2015;11(5): 711-5.

28. Koike N et al. Epigenetic dysregulation of endometriosis susceptibility genes (Review). Mol Med Rep. 2015;12(2):1611-6.

29. Patel BG et al. Progesterone resistance in endometriosis: origins, consequences and interventions. Acta Obstet Gynecol Scand. 2017:96(6): 623-32.

30. Krikun G et al. Endometriosis and tissue factor. Ann N Y Acad Sci. 2008; 127(1):101-5.

31. Zorio E et al. Fibrinolysis: the key to new pathogenetic mechanisms. Curr Med Chem. 2008;15(9):923-9.

32. Porter KM et al. Cathepsin Protease inhibition reduces endometriosis lesion establishment. Reprod Sci. 2016; 23(5):623-9. 33. Paula R Jr et al. The intricate role of mast cell proteases and the annexin A1-FPR1 system in abdominal wall endometriosis. J Mol Histol. 2015;46(1): 33-43.

34. Pitsos M, Kanakas N. The role of matrix metalloproteinase in the pathogenesis of endometriosis. Reprod Sci. 2009;16(8):717-26.

35. Osteen KG et al. Progesteronemediated endometrial maturation limits matrix metalloproteinase (MMP) expression in an inflammatory-like environment: a regulatory system altered in endometriosis. Ann N Y Acad Sci. 2002: 955:37-47.

36. Monckedieck V et al. Progestins inhibit expression of MMPs and of angiogenic factors in human ectopic endometrial lesions in a mouse model. Mol Hum Reprod. 2009;15(10):633-43.

37. Zhang L et al. Intracellular Wnt/ beta-catenin signaling underlying 17beta-estradiol-induced matrix metalloproteinase 9 expression in human endometriosis. Biol Reprod. 2016; 94(3):70.

38. Liu H et al. Correlation between matrix metalloproteinase-9 and endometriosis. Int J Clin Exp Pathol. 2015;8(10):13399-404.

39. Bulun SE et al. Role of estrogen receptor- $\beta$  in endometriosis. Semin Reprod Med. 2012;30(1):39-45.

40. Shao R et al. The elusive and controversial roles of estrogen and progesterone receptors in human endometriosis. Am J Transl Res. 2014; 6(2):104-13.

41. Achari Y et al. Polymorphisms in the promoter regions for human MMP-1 and MMP-13 lead to differential responses to alpha and beta isoforms of estrogen receptors and their ligand in vitro. Biochim Biophys Acta. 2008;1782(6): 391-400.

42. Achari Y et al. Distinct roles for AF-1 and -2 of ER-alpha in regulation of MMP-13 promoter activity. Biochim Biophys Acta. 2009;1792(3):211-20.

43. Lu et al. Evidence that estrogen receptor beta enhances MMP-13 promoter activity in HIG-82 cells and that this enhancement can be influenced by ligands and involves specific promoter sites. Biochem Cell Biol. 2007;85(3): 326-36.

44. Yang H et al. Associations between various possible promoter polymorphisms of MMPs genes and endometriosis risk: a meta-analysis. Eur J Obstet Gynecol Reprod Biol. 2016; 205:174-88. 45. Thaler JD et al. Estrogen receptor beta and truncated variants enhance the expression of transfects MMP-1 promoter constructs in response to specific mechanical loading. Biol Sex Differ. 2014; 5(1):14.

46. Juhasz-Böss I et al. Endometrial expression of estrogen receptor beta and its splice variants in patients with and without endometriosis. Arch Gynecol Obstet. 2011;284(4): 885-91.

47. Kempuraj D et al. Increased numbers of activated mast cells in endometriosis lesions positive for corticotropin-releasing hormone and urocortin. Am J Reprod Immunol. 2004;52(4):267-75.

48. Sugamata M et al. Increase of activated mast cells in human endometriosis. Am J Reprod Immunol. 2005;53(3):120-5.

49. Anaf V et al. Pain, mast cells, and nerves in peritoneal, ovarian, and deep infiltrating endometriosis. Fertil Steril. 2006;86(5):1336-43.

50. Cottrell GS et al. Protease-activated receptors: the role of cell-surface proteolysis in signalling. Essays Biochem. 2002;38:169-83.

51. Osuga Y et al. Proteinase-activated receptors in the endometrium and endometriosis. Front Biosci. 2012;4: 1201-12.

52. Bao Y et al. Protease-activated receptor 2 signalling pathways: a role in pain processing. Expert Opin Ther Targets. 2014;18(1):15-27.

53. Ammendola M et al. Targeting mast cells tryptase in tumor microenvironment: a potential antiangiogenic strategy. Biomed Res Int. 2014;2014:154702.

54. D'Cruz OJ, Uckun FM. Targeting mast cells in endometriosis with janus kinase 3 inhibitor, JANEX-1. Am J Reprod Immunol. 2007;58(2):75-97.

55. Zhu LB et al. Sodium cromoglycate attenuates experimental endometriosis in rats by regulating mast cells. Zhejiang Da Xue Xue Bao Yi Xue Ban. 2015;44(3): 278-84. (In Chinese).

56. Gallant CL et al. Molecular, histologic, and gross phenotype of skin wound healing in red Duroc pigs reveals an abnormal healing phenotype of hypercontracted, hyperpigmented scarring. Wound Repair Regen. 2004; 12(3):305-19.

57. Gallant-Behm CL et al. The mast cell stabilizer ketotifen prevents development of excessive skin wound contraction and fibrosis in red Duroc pigs. Wound Repair Regen. 2008;16(2):226-33.

58. Hildebrand KA et al. Joint capsule turnover in a rabbit model of chronic joint contracture: correlation with human contractures. J Orthop Res. 2006;24(5): 1036-43.

59. Engrav LH et al. Hypertrophic scar, wound contraction and hyperhypopigmentation. J Burn Care Res. 2007; 28(4):593-7.

60. Gallant-Behm CL et al. Skin wound healing in the first generation (F1) offspring of Yorskshire and red Duroc pigs: evidence for genetic inheritance of wound phenotype. Burns. 2006;32(2): 180-93.

61. Monument M et al. The mast cell stabilizer, ketotifen fumarate, lessens contracture severity and myofibroblast hyperplasia in a rabbit model of posttraumatic joint contractures. J Bone Joint Surg. 2010;92(6):1468-77.

62. Monument et al. The mast cell stabilizer ketotifen reduces joint capsule fibrosis in a rabbit model of post-traumatic joint contractures. Inflamm Res. 2012;61(4):285-92.

63. Kopka M et al. Serum mast cell tryptase as a marker of posttraumatic joint contracture in a rabbit model. J Orthop Trauma. 2016;31(3)e86-9.

64. Palmer SS et al. Bentamapimod (JNK Inhibitor AS602801) induces regression of endometriotic lesions in animal models. Reprod Sci. 2016:23(1):11-23.

65. University of Calgary. Investigations of mechanisms and treatment in post-traumatic joint contractures (Ketotifen). NCT01902017. Available at: https://clinicaltrials.gov/ct2/show/NCT01902017.

66. Thompson CM et al. Genetic risk factors for hypertrophic scar development. J Burn Care Res. 2013; 34(5):477-82.

67. Jensen F et al. Estradiol and progesterone regulate the migration of mast cells from the periphery to the uterus and induce their maturation and degranulation. PLoS One. 2010; 5(12):e14409.