

More than FOLFOX and FOLFIRI: The Management of Metastatic Colorectal Cancer in the Era of Precision Oncology

**EDITOR'S
PICK**

My chosen article for the Editor's Pick in this issue is 'More than FOLFOX and FOLFIRI: Management of Metastatic Colorectal Cancer in the Era of Precision Oncology' by Jácome and Johnson. The paper discusses the current landscape and standard of care for metastatic colorectal cancer (mCRC), a disease known for its heterogeneity and poor prognosis. Enhancements in molecular biology in relation to oncology have allowed the identification of specific molecular subtypes and novel therapeutic targets. In the review, Jacome and Johnson describe the current and emerging predictive biomarkers in mCRC and present landmark clinical trials that have allowed for evolving and improving precision in therapeutic management of the disease. Promising findings with targeted therapies offer the possibility of a new era of precision oncology and personalised treatments sustaining hope for patients with mCRC.

Authors: Alexandre A. Jácome,¹ Benny Johnson²

1. Department of Gastrointestinal Medical Oncology, Oncoclínicas, Belo Horizonte, Minas Gerais, Brazil
2. Department of Gastrointestinal Medical Oncology, Division of Cancer Medicine, The University of Texas MD Anderson Cancer Center, Houston, Texas, USA

*Correspondence to bjohnson6@mdanderson.org

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Abstract

Metastatic colorectal cancer (mCRC) is a markedly heterogeneous disease, which portends a poor prognosis, with an estimated 5-year overall survival rate of approximately 15%. The standard of care of systemic therapy remains fluoropyrimidine-based chemotherapy, with modest results, despite improvements with the combination with anti-angiogenics and anti-epidermal growth factor receptor therapy.

Significant advances in cancer therapy have been observed in the past two decades. The enhanced appreciation of molecular biology in oncology has allowed for the identification of specific molecular subtypes and novel therapeutic targets. Nevertheless, meaningful precision-based advancements in the therapeutic options for mCRC have been challenging and slow to realisation. Comprehensive molecular profiling and circulating tumour DNA highlight a heterogeneous disease at the genomic, epigenomic, and transcriptomic levels, and with a low frequency of actionable alterations.

In the present review, the authors describe the current and emerging predictive biomarkers in mCRC, as well as present landmark clinical trials that have allowed for evolving precision in the therapeutic management. The understanding of the benefit of immune checkpoint inhibitors in patients with high microsatellite instability cancer and in those with *POLE* mutations or high tumour mutational burden, the combination of BRAF with epidermal growth factor receptor inhibition in *BRAF* V600-mutated patients, the use of allele-specific *KRAS* G12C inhibitors, the promising findings of dual anti-HER2 therapy in *HER2*-positive mCRC, and the possibility to offer targeted therapy for patients harbouring gene fusions *NTRK/ALK/ROS1* have ushered in a new era of precision oncology for mCRC, providing personalised treatments and sustaining hope for patients affected by this challenging disease.

INTRODUCTION

Colorectal cancer (CRC) is the second most common cause of cancer-related death in the USA.¹ Despite improvements in screening rates and in the overall survival (OS) of patients with localised and advanced disease over the past few decades, the 5-year OS of patients with metastatic disease is still extremely poor and estimated to be approximately 15%.^{2,3}

Significant advances in cancer therapy have been observed in the past two decades. The enhanced appreciation of molecular biology in oncology has allowed for the identification of specific molecular subtypes and novel therapeutic targets. This era of precision oncology allows for the development of biomarker-guided therapeutics and has markedly transformed the landscape of cancer treatment. Precision medicine represents a paradigm shift in oncology, moving from a histology-based chemotherapy to include genome-specific targeted therapy, which has promoted ongoing discovery for novel biomarkers in all malignancies.

Nevertheless, the emergence of precision oncology has drastically improved the management of CRC. Comprehensive molecular profiling confirms a markedly heterogeneous disease at the genomic, epigenomic, and transcriptomic levels, but currently with low frequency of actionable alterations. For more than a decade, personalised therapy in CRC was restricted to the identification of *RAS* mutations, which are predictive markers of resistance to anti-epidermal growth factor receptor (EGFR) monoclonal antibodies. Only recently, clinical

trials addressing novel genome-guided personalised therapies in specific molecular subtypes of CRC have been successfully completed, expanding the clinical relevance of precision oncology in CRC.

In this review, the authors describe the current and emerging predictive biomarkers in metastatic CRC (mCRC), as well as present landmark clinical trials that have allowed for evolving precision in the management of this heterogeneous disease.

ANTI-EPIDERMAL GROWTH FACTOR RECEPTOR MONOCLONAL ANTIBODIES IN METASTATIC COLORECTAL CANCER IN THE PAST TWO DECADES: PRECISION THAT NEEDS FURTHER REFINEMENT

For almost two decades, the clinical applicability of precision in mCRC has been limited to the use of the anti-EGFR monoclonal antibodies cetuximab and panitumumab for *RAS* wild-type disease. The knowledge accumulated over the past 20 years has demonstrated that the benefit offered by these monoclonal antibodies is restricted to a smaller subset of mCRC than initially proposed (Figure 1). Pure predictive biomarkers reflecting patient specific sensitivity to anti-EGFR monoclonal antibodies remains a developing area in mCRC. Interestingly, evidence accumulated over the past few years suggest that not only expanded *RAS* mutations such as *NRAS* and *HRAS* may confer additional resistance to cetuximab or panitumumab but also *BRAF*, *PI3KCA*, *HER2*, *MET*, *PTEN*, and *AKT1* abnormalities, as well as *NTRK/ROS1/ALK/RET* rearrangements.⁴⁻⁹ Furthermore, recent studies have consistently demonstrated that patients with right-sided tumours derive lower, if any, benefit from that therapy.^{10,11}

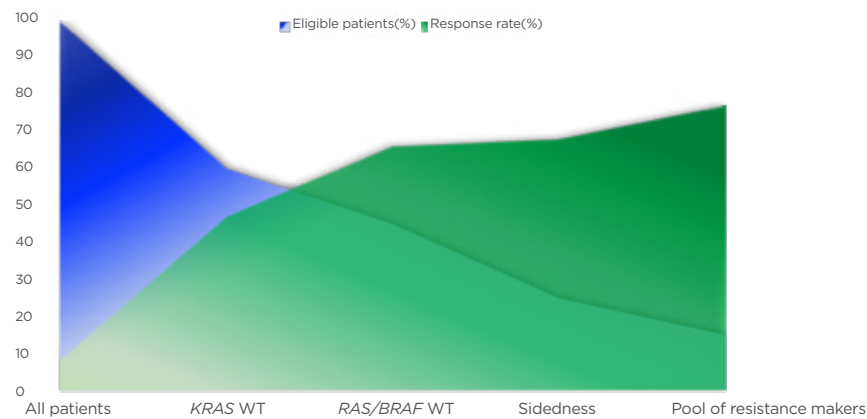


Figure 1: Negative hyper-selection of patients with metastatic colorectal cancer to anti-epidermal growth factor receptor monoclonal antibodies.

The graph shows the ascending response rate (y-axis, in green) of anti-EGFR therapy in mCRC based on patient selection by biomarkers (*RAS* and *BRAF*), sidedness, and a pool of resistance markers such as *BRAF*, *PI3KCA*, *HER2*, *MET*, *PTEN*, and *AKT1* abnormalities, as well as *NTRK/ROS1/ALK/RET* rearrangements, which lead to a descending rate of eligible patients for targeted therapy (x axis, in blue). Since there are no predictive biomarkers of sensitivity for anti-EGFR therapy, the patient selection based on predictive markers of resistance may be denominated as negative hyper-selection.

EGFR: epidermal growth factor receptor; mCRC: metastatic colorectal cancer; WT: wild-type.

Hence, the estimated rate of patients with mCRC who are actually sensitive to cetuximab or panitumumab is lower than approximately 15%.⁹ The current recommendation to use anti-EGFR monoclonal antibodies in patients with left-sided tumours and *RAS/BRAF* wild-type status still is an incipient and imprecise clinical applicability of precision medicine in the systemic therapy of CRC. Therefore, the identification of refined biomarkers and novel targeted therapies are urgently needed (Figure 2; Table 1).

NOVEL THERAPEUTIC TARGETS IN COLORECTAL CANCER

MSI-H CRC

It is estimated that approximately 5% of patients with mCRC harbour high-frequency microsatellite instability (MSI-H), which might originate from two mechanisms: somatic hypermethylation of the *MLH1* gene promoter, commonly associated with *BRAF* V600E mutation; or point mutation of one of the mismatch repair genes, mainly *MLH1* and *MSH2*.^{12,13} Patients with MSI-H CRC compose a subgroup with distinct molecular and

clinical characteristics. Typically, they present a younger median age at diagnosis, with tumours predominantly located at the proximal colon, commonly with lymphocyte infiltration, and with a higher median number of tumour mutational burden (TMB).¹³ In addition, they have lower sensitivity to chemotherapy compared with patients classified as microsatellite stable (MSS), and, more importantly, they tend to be sensitive to immunotherapeutic approaches, such as immune checkpoint inhibitors (ICIs).

Encouraging data from Phase I and II clinical trials¹⁴⁻¹⁹ prompted the conception of the Phase III KEYNOTE-177 study, which compared the efficacy of standard chemotherapy (doublets plus anti-vascular endothelial growth factor or anti-EGFR) with pembrolizumab in 307 treatment-naïve patients with MSI-H mCRC.²⁰ One of the primary endpoints, progression-free survival, was met: 8.2 months in the chemotherapy group versus 16.5 months in the immunotherapy group (hazard ratio [HR]: 0.60; 95% confidence interval [CI]: 0.45-0.80; $p=0.0002$). Likewise, the overall response rate (ORR) was statistically higher in the immunotherapy arm: 33.1% versus 43.8%.

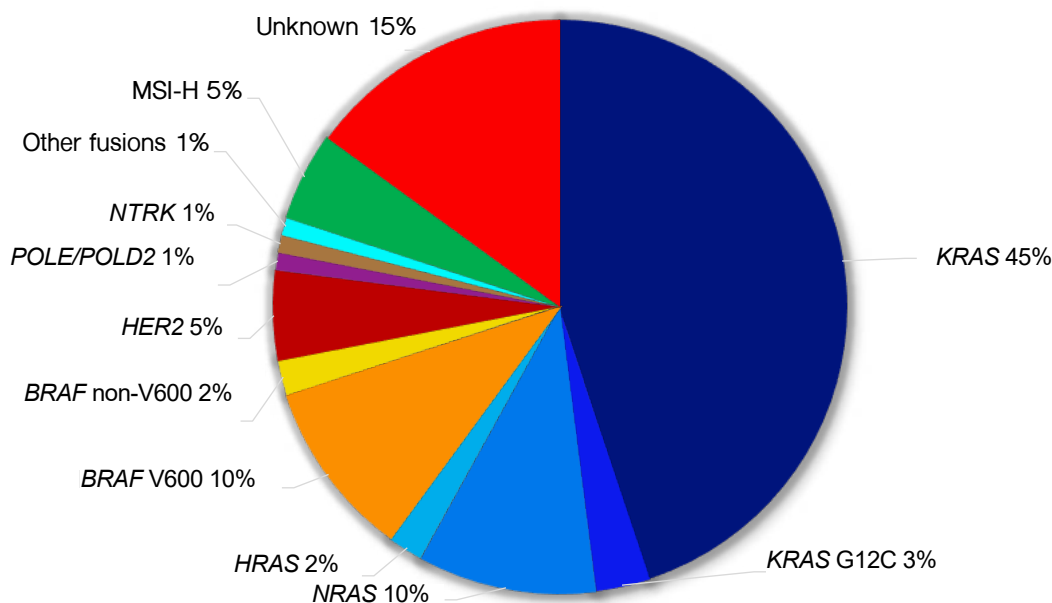


Figure 2: Molecular alterations with therapeutic implications in metastatic colorectal cancer.

MSI-H: high-frequency microsatellite instability.

Table 1: Predictive biomarkers and targeted therapies in metastatic CRC.

Biomarkers	Rates*	Predictive role to systemic therapy
<i>KRAS</i> non-G12C	45%	Resistance to anti-EGFR therapy
<i>KRAS</i> G12C	3%	Resistance to anti-EGFR therapy Poor sensitivity to sotorasib or adagrasib alone
<i>NRAS</i>	10%	Resistance to anti-EGFR therapy
<i>HRAS</i>	2%	Resistance to anti-EGFR therapy
<i>BRAF</i> V600E	10%	Sensitivity to encorafenib+cetuximab±binimetinib Resistance to anti-EGFR therapy
<i>BRAF</i> non-V600E	2%	Uncertain
<i>HER2</i> amplification	5%	Sensitivity to anti- <i>HER2</i> therapies Potential resistance to anti-EGFR therapy
<i>NTRK</i> fusions	1%	Sensitivity to larotrectinib or entrectinib
<i>ALK/ROS1</i> fusions	<1%	Sensitivity to <i>ALK/ROS1</i> inhibitors
MSI-H	5%	Sensitivity to immune checkpoint inhibitors
<i>POLE/POLD2</i>	1%	Potential sensitivity to immune checkpoint inhibitors

*Approximate rates.

MSI-H: high-frequency microsatellite instability, EGFR: epidermal growth factor receptor.

Interestingly, 29.4% of the patients in the pembrolizumab group presented progressive disease compared with 12.3% in the chemotherapy group, predominantly in the first 4 months of treatment. Of the patients who presented objective response to immunotherapy, an impressive rate of 83% had ongoing responses at 24 months, compared with only 35% in the chemotherapy group. Another primary endpoint, OS, did not have mature data to be analysed.

MSI-H is not the only biomarker to explain sensitivity to ICIs. Patients with MSI-H who present with low TMB seem to have a lower probability to respond to immunotherapy.²¹ A high concordance rate is expected between MSI-H and TMB. Patients with abnormalities in DNA mismatch repair pathways, whether germline or somatic, tend to present higher number of nonsynonymous mutations, and thereby high TMB. In a study with 6,004 patients with mCRC, 5% were classified as MSI-H and 95% as MSS.²² The median TMB was significantly higher in the population with MSI-H: 46.8 mutations/Mb versus 3.6 mutations/Mb. The median TMB in the overall population was 4.5 mutations/Mb. Approximately 3% of patients with MSS were classified as high TMB, defined as ≥ 12 mutations/Mb. Variants in the mismatch repair genes, such as *MLH1*, *MSH2*, and *MSH6*, as well as in *POLE*, were significantly more common in this population of patients with high TMB and MSS relative to those with low TMB and MSS.²²

POLE

POLE encodes the catalytic subunit of DNA polymerase ϵ , which acts in the replication of the DNA strand before cell division.²³ *POLE* proofreading is an essential step in the maintenance of the integrity of the genome, which is consistent with the finding of ultra-mutated tumours in the presence of pathogenic exonuclease domain mutations, with a mean tumour mutational burden >200 mutations/Mb.^{24,25} *POLE* mutation is rarely found in malignancies, being identified in approximately 5–10% of endometrial cancer,²⁶ 1% of CRC,²⁷ and less frequently in gastric and pancreatic cancers.²⁸

POLE-mutated CRC portends a better prognosis. In a population of 6,517 patients

with CRC, 1% (66 patients) harboured the mutation, which was associated with a reduced risk of recurrence and a superior OS in a population of patients with Stage II/III CRC.²³ Patients with *POLE*-mutated CRC were younger at diagnosis, predominantly male, with a higher frequency of right-sided tumours, and with disease diagnosed at earlier stages compared with the wild-type counterparts.^{23,25,29} They also demonstrated increased CD8+ lymphocyte infiltration and expression of cytotoxic T-cell markers.²³

This immunogenic subset of CRC has been demonstrated to be highly sensitive to the ICIs. Case reports with successful experiences in the treatment of metastatic CRC have been presented in the past few years.^{31,32} Since there is a U.S. Food and Drug Administration (FDA) approval for the use of pembrolizumab for patients with metastatic disease and TMB >10 mutations/Mb,³² the use of immunotherapy should be strongly considered in the treatment of patients with *POLE*-mutated mCRC. The probability to identify these mutations is higher in early-onset CRC compared to the late-onset.³³

BRAF

BRAF V600E mutation is found in approximately 10% of patients with mCRC.^{34–37} These patients have a poorer prognosis compared with the wild-type counterparts, demonstrating lower sensitivity to the standard chemotherapeutic drugs used in CRC, with lower ORR, and shorter progression-free survival and OS.^{38–40} More commonly found in right-sided tumours, this mutation, similarly to *RAS* mutations, also denotes resistance to the anti-EGFR monoclonal antibodies.^{41,42}

The success of BRAF inhibitors in the systemic therapy of *BRAF*-mutated melanoma prompted the evaluation of these drugs in mCRC. However, Phase I data addressing the efficacy of vemurafenib in patients with *BRAF*-mutated mCRC showed poor efficacy.⁴³ Preclinical studies demonstrated that BRAF inhibition induced adaptive feedback reactivation of mitogen-activated protein kinase signalling, often mediated by EGFR activation, suggesting that the combination of BRAF inhibitor with an anti-EGFR monoclonal antibody might overcome this

therapeutic resistance.⁴⁴⁻⁴⁶ Subsequently, a Phase IB study confirmed the hypothesis, demonstrating that the combination of vemurafenib, irinotecan, and cetuximab yielded 35% of ORR in a population of 19 patients with *BRAF*-mutated mCRC.⁴⁷ Additionally, further work demonstrated the clinical activity of the combination of *BRAF* and EGFR inhibition with or without mitogen-activated protein kinase kinase (MEK) inhibition in *BRAF*-mutated mCRC.⁴⁴

These promising findings elicited the conception of the Phase III BEACON study, a three-arm clinical trial that explored the combination of *BRAF* and EGFR inhibition with MEK inhibition. A total of 665 patients with *BRAF* V600E-mutated mCRC who had been submitted to at least one previous line of systemic therapy were randomised to one of three arms: the triplet-regimen composed of encorafenib plus binimetinib plus cetuximab; the doublet-regimen with encorafenib plus cetuximab; and the control arm with irinotecan-based regimens (folinic acid, fluorouracil, and irinotecan; or irinotecan) plus cetuximab.⁴⁸ The primary end points were OS and ORR in the triplet-regimen arm compared to the control arm. Updated survival results showed a median OS of 9.3 months in the triplet arm versus 5.9 months in the control arm (HR: 0.60; 95% CI: 0.47-0.75).⁴⁹ The ORR was 27%, 20%, and 2% in the triplet, doublet, and control arms, respectively. A comparison of the median OS in the doublet arm (9.3 months) with the control arm, a secondary endpoint, also favoured the *BRAF* inhibitor (HR: 0.61; 95% CI: 0.48-0.77). There was no statistically significant difference between the triplet and doublet arms in OS: 9.3 months in both groups (HR: 0.95; 95% CI: 0.74-1.21).⁴⁹ Grade ≥ 3 adverse events were found in 66%, 58%, and 64% of the patients in the triplet, doublet, and control arms, respectively. Based on BEACON data, the FDA has approved the combination of encorafenib plus cetuximab for the treatment of patients with mCRC and a *BRAF* V600E mutation with at least one prior systemic therapy.⁵⁰

The clinical relevance of non-V600 *BRAF* mutations has not yet been fully elucidated. These mutations have been found in approximately 2% of patients with mCRC, of which the D594N (Class III) and G469A (Class II) mutations seem to be the most frequent.⁵¹

The patients who harbour these atypical *BRAF* mutations seem to present similar prognosis compared to the wild-type counterparts. Unlike V600E, these atypical mutations are mostly identified in left-sided tumours, and younger male patients.⁵¹ In addition, most of them are MSS and *RAS* mutations are not mutually exclusive in this context, occurring in approximately one-third of non-V600 patients.^{51,52} The predictive value of these mutations for the deployment of anti-EGFR monoclonal antibodies is not yet entirely clear, and appears to differ according to the underlying *BRAF* class. Class II mutations appear to be resistant while Class III are sensitive, although with limited duration.⁵³⁻⁵⁵ Furthermore, non-V600 *BRAF* mutations might be involved in the development of adaptive resistance to EGFR inhibition.⁵¹ These distinct class-specific biochemical and functional properties highlight the importance to decipher the unique biology of atypical *BRAF* mutations in order to promote novel clinical trial design and ultimately offer effective therapeutic options for patients.

KRAS G12C

KRAS mutations are the most common activating genetic mutations in solid tumours, mainly in pancreatic cancer, non-small cell lung cancer, and CRC, where they are estimated to be found in approximately 45% of tumours.^{37,56,57} Right-sided tumours present a higher percentage of *KRAS* mutations compared with their left-sided counterparts, mainly in the cecum, where approximately 70% of the tumours harbour the mutation.⁵⁶ For decades, *KRAS* mutations have not been deemed as actionable, but, together with other *RAS* mutations, they predict resistance to anti-EGFR monoclonal antibodies in CRC.^{58,59}

Despite years of research focus, targeting *KRAS* has been an elusive goal in cancer therapy since the mutated protein has high affinity for guanosine triphosphate or guanosine diphosphate and has no binding pocket.⁶⁰ In addition, inhibition of the downstream effectors in the mitogen-activated protein kinase pathway (*BRAF*-*MEK*-*ERK*) has proven ineffective in clinical trials.⁶⁰

The codons 12 and 13 in exon 2 are the most commonly altered in *KRAS* mutations, occurring

in approximately 30% and 10%, respectively, of the patients with mCRC.⁵⁷ The amino acid changes p.G12D, p.G12V, and p.G13D are the most frequent of these codons in CRC, found in approximately 13%, 10%, and 9% of the patients, respectively.⁵⁷ The oncoprotein KRAS p.G12C is found in 1–3% of patients with mCRC.^{57,61} The substitution of glycine for cysteine at position 12 results in a predominantly guanosine triphosphate-bound KRAS protein, the active form, favouring proliferation and survival of tumour cells.^{62,63}

Recently, the isoform *KRAS* G12C has demonstrated to be targetable by a covalent allele-specific inhibitor. Sotorasib (AMG510) is a small molecule that specifically and irreversibly inhibits *KRAS* G12C in its inactive guanosine diphosphate-bound state through an interaction with one of its pockets.⁶¹ It was evaluated in a Phase I trial with 129 previously treated patients with advanced solid tumours harbouring the *KRAS* G12C mutation.⁶¹ In the overall trial population, of the 42 patients with CRC, only 3 (7%) presented partial response, but 28 (67%) experienced stable disease. On the other hand, 32% of the 59 patients with non-small cell lung cancer had partial response, and 56% showed stable disease. Adagrasib (MRTX849) is another *KRAS* G12C inhibitor under therapeutic development and it has shown promising efficacy in preclinical studies and preliminary clinical findings.⁶⁴ Additional Phase I/II clinical trials are currently evaluating the efficacy of adagrasib in *KRAS* G12C-mutated malignancies, and an ongoing Phase III clinical trial is comparing the efficacy of adagrasib in combination with cetuximab versus chemotherapy in the second-line setting for patients with mCRC and *KRAS* G12C mutation (NCT04793958).⁶⁵

Interestingly, patients with *KRAS* G12C-mutated mCRC seem to present poorer clinical outcomes compared with the patients with *KRAS* non-G12C mutations.^{66,67} A recent single-institutional study identified 187 patients with *KRAS* G12C from an original population of 4,685 patients with mCRC.⁶⁶ When compared to a cohort of 720 patients with *KRAS* non-G12C mutations, these 187 patients had shorter OS, excluding patients who had undergone metastasectomy: 21.2 months versus 31.6 months ($p=0.003$). Another cohort of 839 patients with mCRC also found an inferior OS in G12C population compared with the non-G12C: 25.9 months versus 35.8

months (HR: 1.55; 95% CI: 1.08–2.24; $p=0.018$), which was confirmed by multivariate analysis (HR: 1.81; 95% CI: 1.20–2.70; $p=0.04$).⁶⁷ Correlative findings also demonstrated that this subgroup of patients with mCRC show a distinct mutational profile, with higher rates of *APC* co-mutations compared with the patients without the G12C mutation, but lower rates of *BRAF*, *ERBB4*, *NRAS*, and *TP53* co-mutations.

The reasons for the different efficacy of the *KRAS* G12C inhibitor according to the tissue of origin are not clear.⁶¹ Ongoing translational studies will be crucial in the understanding of the probable intrinsic resistance of *KRAS* G12C-mutated mCRC to the *KRAS* G12C inhibitors as monotherapy, and for the design of clinical trials evaluating the combination of these inhibitors with other therapeutic strategies.

HER2

Comprehensive molecular characterisation of CRC and the greater availability of next-generation sequencing in tumour genomic profiling have demonstrated that *HER2* amplification is found in approximately 5–10% of patients with *RAS* wild-type mCRC.^{7,8,27} This molecular abnormality is predominantly identified in patients with *RAS/BRAF* wild-type patients, who may harbour *HER2* amplification primarily or secondarily as a mechanism of resistance to anti-EGFR therapy.^{58,68–70} Exploratory analyses suggest that patients with *RAS* wild-type who harbour *HER2* amplification derive lower, if any, benefit from anti-EGFR therapy.^{7,8} Based on the successful experiences of anti-*HER2* therapy in *HER2*-amplified breast and gastric cancers, the identification of this molecular abnormality in mCRC prompted the evaluation of anti-*HER2* therapy in clinical trials.

HERACLES was the first clinical trial addressing the efficacy of anti-*HER2* therapy in *HER2*-positive mCRC.⁷¹ This proof-of-concept Phase II study was comprised of 27 treatment-refractory patients, of which 30% presented an objective response to the combination of trastuzumab plus lapatinib, and an additional 44% had stable disease. The subsequent MyPathway Phase II study showed that 32% of the 57 heavily pre-treated patients with *HER2*-positive mCRC had objective response to the

combination of trastuzumab plus pertuzumab.⁷² The Phase II TAPUR basket trial evaluated the same combination of anti-HER2 therapy in 28 previously treated patients with *HER2*-positive mCRC, and 14% of the patients demonstrated objective response.⁷³ Impressive findings were demonstrated by the preliminary results of the Phase II MOUNTAINEER study, which evaluated the combination of tucatinib plus trastuzumab. Of the 22 evaluable previously treated patients with *HER2*-positive mCRC, 55% presented objective response.⁷⁴ Likewise, promising data have also been presented by DESTINY-CRC01 trial, which showed 45% of ORR with trastuzumab deruxtecan in 53 patients with previously treated *HER2*-positive mCRC.⁷⁵ Taken together, these initial clinical trials have demonstrated that *HER2* is a viable therapeutic target in mCRC, with encouraging efficacy data of the dual anti-HER2 therapy in treatment refractory patients. However, the FDA has not yet approved anti-HER2 therapies for mCRC in the USA. The results of the ongoing randomised clinical trial SWOG1613 evaluating the combination of trastuzumab plus pertuzumab in *RAS/BRAF* wild-type patients are eagerly awaited (NCT03365882),⁷⁶ as well as the results of the Phase II DESTINY-CRC02 trial, with more data of trastuzumab deruxtecan in mCRC (NCT04744831).⁷⁷

NTRK*, *ALK*, and *ROS1

NTRK are genes that encode the tropomyosin receptor kinase (Trk) family, which is comprised of three transmembrane proteins, TrkA, TrkB, and TrkC receptors, which are encoded by the *NTRK1*, *NTRK2*, and *NTRK3* genes, respectively.⁷⁸ The signal transduction pathways activated by these receptors are associated with proliferation, differentiation, and survival in normal and neoplastic neuronal cells.⁷⁹ Gene fusions of the *NTRK* are the main molecular abnormalities with known oncogenic and transforming potential.⁸⁰

Based on a study with 408 patients with CRC, it is estimated a prevalence rate of 0.5% of this gene fusion.⁸¹ Efficacy of larotrectinib in *NTRK* fusion-positive mCRC was demonstrated in a basket trial with 55 patients with solid malignancies, of which four had mCRC.⁸² Three patients presented tumour shrinkage and one had stable disease. Entrectinib, a pan-Trk inhibitor, has also been demonstrated to be effective in this subset of patients.⁸³ Other gene fusions, such as those involving *ALK* and *ROS1*, are rarely found in mCRC, but, once present, they portend a poorer prognosis.^{84,85} Targeted therapies, including entrectinib, have been effective in this subgroup of patients with mCRC.⁸⁶

CONCLUSIONS

Meaningful precision-based advancements in the therapeutic options for mCRC have been challenging and slow to realisation. Comprehensive molecular profiling and circulating tumour DNA highlights a markedly heterogeneous disease at the genomic, epigenomic, and transcriptomic levels; however, to date, they only reflect a low frequency of actionable alterations. For almost two decades, clinical applicability of precision oncology in mCRC was limited to the identification of *RAS* mutations as predictive biomarkers of resistance to the use of anti-EGFR monoclonal antibodies. However, novel therapeutic targets have emerged in recent years, refining the landscape of systemic therapy of the disease. The benefit of ICIs in patients with MSI-H and in those with *POLE* mutations or high TMB, the combination of BRAF with EGFR inhibition in patients with *BRAF* V600 mutations, the advent of allele-specific KRAS G12C inhibitors, and the promising findings of dual anti-HER2 therapy in *HER2*-positive mCRC cases have ushered in a new era of precision oncology for mCRC, providing personalised treatments and sustaining hope for patients affected by this disease.

References

1. Siegel RL et al. Cancer statistics, 2020. *CA Cancer J Clin.* 2020;70(1):7-30.
2. Noone A et al. SEER cancer statistics, 1975–2015. 2018. Available at: https://seer.cancer.gov/archive/csr/1975_2015/. Last accessed: 26 august 2021.
3. Siegel RL et al. Colorectal cancer statistics, 2020. *CA Cancer J Clin.* 2020;70(3):145-64
4. Morano F et al. Negative hyperselection of patients with *RAS* and *BRAF* wild-type metastatic colorectal cancer who received panitumumab-based maintenance therapy. *J Clin Oncol.* 2019;37(33):3099-110.
5. Di Nicolantonio F et al. Wild-type *BRAF* is required for response to panitumumab or cetuximab in metastatic colorectal cancer. *J Clin Oncol.* 2008;26(35):5705-12.

6. Loupakis F et al. *KRAS* codon 61, 146 and *BRAF* mutations predict resistance to cetuximab plus irinotecan in *KRAS* codon 12 and 13 wild-type metastatic colorectal cancer. *Br J Cancer*. 2009;101(4):715-21.
7. Raghav K et al. Validation of *HER2* amplification as a predictive biomarker for anti-epidermal growth factor receptor antibody therapy in metastatic colorectal cancer. *JCO Prec Oncol*. 2019;3:1-13.
8. Martin V et al. *HER2* gene copy number status may influence clinical efficacy to anti-EGFR monoclonal antibodies in metastatic colorectal cancer patients. *Br J Cancer*. 2013;108(3):668-75.
9. Bardelli A, Siena S. Molecular mechanisms of resistance to cetuximab and panitumumab in colorectal cancer. *J Clin Oncol*. 2010;28(7):1254-61.
10. Venook AP et al. Effect of first-line chemotherapy combined with cetuximab or bevacizumab on overall survival in patients with *KRAS* wild-type advanced or metastatic colorectal cancer: a randomized clinical trial. *JAMA*. 2017;317(23):2392-401.
11. Arnold D et al. Prognostic and predictive value of primary tumour side in patients with *RAS* wild-type metastatic colorectal cancer treated with chemotherapy and EGFR directed antibodies in six randomized trials. *Ann Oncol*. 2017;28(8):1713-29.
12. Boland CR et al. A National Cancer Institute Workshop on microsatellite instability for cancer detection and familial predisposition: development of international criteria for the determination of microsatellite instability in colorectal cancer. *Cancer Res*. 1998;58(22):5248-57.
13. Sinicrope FA. Lynch Syndrome-associated colorectal cancer. *N Engl J Med*. 2018;379(8):764-73.
14. Overman MJ et al. Durable clinical benefit with nivolumab plus ipilimumab in DNA mismatch repair-deficient/microsatellite instability-high metastatic colorectal cancer. *J Clin Oncol*. 2018;36(8):773-79.
15. Overman MJ et al. Nivolumab in patients with metastatic DNA mismatch repair-deficient or microsatellite instability-high colorectal cancer (CheckMate 142): an open-label, multicentre, Phase 2 study. *Lancet Oncol*. 2017;18(9):1182-91.
16. Overman MJ et al. Nivolumab ± ipilimumab in treatment (tx) of patients (pts) with metastatic colorectal cancer (mCRC) with and without high microsatellite instability (MSI-H): CheckMate-142 interim results. *J Clin Oncol*. 2016;34(Suppl 15):3501.
17. Le DT et al. PD-1 blockade in tumors with mismatch-repair deficiency. *N Engl J Med*. 2015;372(26):2509-20.
18. Brahmer JR et al. Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. *N Engl J Med*. 2012;366(26):2455-65.
19. Topalian SL et al. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *N Engl J Med*. 2012;366(26):2443-54.
20. Andre T et al. Pembrolizumab versus chemotherapy for microsatellite instability-high/mismatch repair deficient metastatic colorectal cancer: the Phase 3 KEYNOTE-177 Study. *J Clin Oncol*. 2020;38(Suppl 18):LBA4.
21. Schrock AB et al. Tumor mutational burden is predictive of response to immune checkpoint inhibitors in MSI-high metastatic colorectal cancer. *Ann Oncol*. Jul 2019;30(7):1096-103.
22. Fabrizio DA et al. Beyond microsatellite testing: assessment of tumor mutational burden identifies subsets of colorectal cancer who may respond to immune checkpoint inhibition. *J Gastrointest Oncol*. 2018;9(4):610-17.
23. Domingo E et al. Somatic *POLE* proofreading domain mutation, immune response, and prognosis in colorectal cancer: a retrospective, pooled biomarker study. *Lancet Gastroenterol Hepatol*. 2016;1(3):207-16.
24. Pursell ZF. Yeast DNA polymerase ϵ participates in leading-strand DNA replication. *Science*. 2007;317(5834):127-30.
25. Hu H et al. Ultra-mutated colorectal cancer patients with *POLE* driver mutations exhibit distinct clinical patterns. *Cancer Med*. 2021;10(1):135-42.
26. Levine DA. Integrated genomic characterization of endometrial carcinoma. *Nature*. 2013;497(7447):67-73.
27. Kandath C et al.; Cancer Genome Atlas Research Network. Comprehensive molecular characterization of human colon and rectal cancer. *Nature*. 2012;487(7407):330-7.
28. Rayner E et al. A panoply of errors: polymerase proofreading domain mutations in cancer. *Nat Rev Cancer*. 2016;16(2):71-81.
29. Hino H et al. Clinicopathological and mutational analyses of colorectal cancer with mutations in the *POLE* gene. *Cancer Med*. 2019;8(10):4587-97.
30. Gong J et al. Response to PD-1 blockade in microsatellite stable metastatic colorectal cancer harboring a *POLE* mutation. *J Natl Compr Canc Netw*. 2017;15(2):142-7.
31. Silberman R et al. Complete and prolonged response to immune checkpoint blockade in *POLE*-mutated colorectal cancer. *JCO Prec Oncol*. 2019;3:1-5.
32. Merck & Co., Inc. Highlights of prescribing information for Keytruda (pembrolizumab) injection. 2014. Available at: https://www.merck.com/product/usa/pi_circulars/k/keytruda/keytruda_pi.pdf. Last accessed: 26 August 2021.
33. Ahn S-M et al. The somatic *POLE* P286R mutation defines a unique subclass of colorectal cancer featuring hypermutation, representing a potential genomic biomarker for immunotherapy. *Oncotarget*. 2016;7(42):68638-49.
34. Yaeger R et al. Clinical sequencing defines the genomic landscape of metastatic colorectal cancer. *Cancer Cell*. 2018;33(1):125-36.
35. Barras D et al. *BRAF* V600E mutant colorectal cancer subtypes based on gene expression. *Clin Cancer Res*. 2017;23(1):104-15.
36. Bysma LC et al. Prevalence of *RAS* and *BRAF* mutations in metastatic colorectal cancer patients by tumor sidedness: a systematic review and meta-analysis. *Cancer Med*. 2020;9(3):1044-57.
37. AACR Project GENIE Consortium. AACR Project Genie: powering precision medicine through an international consortium. *Cancer Discov*. 2017;7(8):818-31.
38. Clarke CN, Kopetz ES. *BRAF* mutant colorectal cancer as a distinct subset of colorectal cancer: clinical characteristics, clinical behavior, and response to targeted therapies. *J Gastrointest Oncol*. 2015;6(6):660-7.
39. Tran B et al. Impact of *BRAF* mutation and microsatellite instability on the pattern of metastatic spread and prognosis in metastatic colorectal cancer. *Cancer*. 2011;117(20):4623-32.
40. Loupakis F et al. Initial therapy with FOLFOXIRI and bevacizumab for metastatic colorectal cancer. *N Engl J Med*. 2014;371(17):1609-18.
41. Douillard JY et al. Panitumumab-FOLFOX4 treatment and *RAS* mutations in colorectal cancer. *N Engl J Med*. 2013;369(11):1023-34.
42. Pietrantonio F et al. Predictive role of *BRAF* mutations in patients with advanced colorectal cancer receiving cetuximab and panitumumab: a meta-analysis. *Eur J Cancer*. 2015;51(5):587-94.
43. Kopetz S et al. Phase II pilot study of vemurafenib in patients with metastatic *BRAF*-mutated colorectal cancer. *J Clin Oncol*. 2015;33(34):4032-8.
44. Corcoran RB et al. Combined *BRAF*, *EGFR*, and *MEK* inhibition in patients with *BRAF*^{V600E}-mutant colorectal cancer. *Cancer Discov*. 2018;8(4):428-43.
45. Prahallad A et al. Unresponsiveness of colon cancer to *BRAF* (V600E) inhibition through feedback activation of *EGFR*. *Nature*. 2012;483(7387):100-3.

46. Mao M et al. Resistance to BRAF inhibition in BRAF-mutant colon cancer can be overcome with PI3K inhibition or demethylating agents. *Clin Cancer Res.* 2013;19(3):657-67.
47. Hong DS et al. Phase IB study of vemurafenib in combination with irinotecan and cetuximab in patients with metastatic colorectal cancer with *BRAF*^{V600E} mutation. *Cancer Discov.* 2016;6(12):1352-65.
48. Kopetz S et al. Encorafenib, binimetinib, and cetuximab in *BRAF* V600E-mutated colorectal cancer. *N Engl J Med.* 2019;381(17):1632-43.
49. Kopetz S et al. Encorafenib plus cetuximab with or without binimetinib for *BRAF* V600E metastatic colorectal cancer: Updated survival results from a randomized, three-arm, Phase III study versus choice of either irinotecan or FOLFIRI plus cetuximab (BEACON CRC). *J Clin Oncol.* 2020;38(Suppl 4):8.
50. Braftovi. (encorafenib). Array BioPharma Inc. Boulder. CO.2020.
51. Johnson B et al. Atypical, non-V600 BRAF mutations as a potential mechanism of resistance to EGFR inhibition in metastatic colorectal cancer. *JCO Precis Oncol.* 2019;3:1-10.
52. Kotani D et al. Clinicopathological features, efficacy of anti-EGFR therapy, and survival outcomes in patients with *BRAF* non-V600 mutated metastatic colorectal cancer. *J Clin Oncol.* 2019;37(Suppl 4):659.
53. Fontana E, Valeri N. Class(y) dissection of *BRAF* heterogeneity: beyond non-V600. *Clin Cancer Res.* 2019;25(23):6896-8.
54. Wang Y et al. Activity of EGFR antibody in non-V600 *BRAF* mutant metastatic colorectal cancer. *Ann Oncol.* 2019;30(1):147-49.
55. Yaeger R et al. Response to anti-EGFR therapy in patients with *BRAF* non-V600-mutant metastatic colorectal cancer. *Clin Cancer Res.* 2019;25(23):7089-97.
56. Loree JM et al. Classifying colorectal cancer by tumor location rather than sidedness highlights a continuum in mutation profiles and consensus molecular subtypes. *Clin Cancer Res.* 2018;24(5):1062-72.
57. Vaughn CP et al. Frequency of *KRAS*, *BRAF*, and *NRAS* mutations in colorectal cancer. *Genes Chromosomes Cancer.* 2011;50(5):307-12.
58. Misale S et al. Emergence of *KRAS* mutations and acquired resistance to anti-EGFR therapy in colorectal cancer. *Nature.* 2012;486(7404):532-6.
59. Sorich M et al. Extended *RAS* mutations and anti-EGFR monoclonal antibody survival benefit in metastatic colorectal cancer: a meta-analysis of randomized, controlled trials. *Ann Oncol.* 2015;26(1):13-21.
60. Moore AR et al. RAS-targeted therapies: is the undruggable drugged? *Nat Rev Drug Discov.* 2020;19(8):533-52.
61. Hong DS et al. *KRAS*^{G12C} Inhibition with sotorasib in advanced solid tumors. *N Engl J Med.* 2020;383(13):1207-17.
62. Ostrem JM et al. K-Ras(G12C) inhibitors allosterically control GTP affinity and effector interactions. *Nature.* 2013;503(7477):548-51.
63. Kargbo RB. Inhibitors of G12C mutant Ras proteins for the treatment of cancers. *ACS Med Chem Lett.* 2018;10(1):10-11.
64. Hallin J et al. The *KRAS*^{G12C} inhibitor MRTX849 provides insight toward therapeutic susceptibility of *KRAS*-mutant cancers in mouse models and patients. *Cancer Discov.* 2020;10(1):54-71.
65. Mirati Therapeutics Inc. Phase 3 study of MRTX849 with cetuximab vs chemotherapy in patients with advanced colorectal cancer with *KRAS* G12C mutation (KRYSTAL-10). NCT04793958. <https://clinicaltrials.gov/ct2/show/NCT04793958>.
66. Henry JT et al. Comprehensive clinical and molecular characterization of *KRAS*^{G12C}-mutant colorectal cancer. *JCO Precis Oncol.* 2021;5:613-21.
67. Schirripa M et al. *KRAS* G12C metastatic colorectal cancer: specific features of a new emerging target population. *Clin Colorectal Cancer.* 2020;19(3):219-25.
68. Pietrantonio F et al. Heterogeneity of acquired resistance to anti-EGFR monoclonal antibodies in patients with metastatic colorectal cancer. *Clin Cancer Res.* 2017;23(10):2414-22.
69. Diaz Jr LA et al. The molecular evolution of acquired resistance to targeted EGFR blockade in colorectal cancers. *Nature.* 2012;486(7404):537-40.
70. Misale S et al. Resistance to anti-EGFR therapy in colorectal cancer: from heterogeneity to convergent evolution. *Cancer Discov.* 2014;4(11):1269-80.
71. Sartore-Bianchi A et al. Dual-targeted therapy with trastuzumab and lapatinib in treatment-refractory, *KRAS* codon 12/13 wild-type, HER2-positive metastatic colorectal cancer (HERACLES): a proof-of-concept, multicentre, open-label, Phase 2 trial. *Lancet Oncol.* 2016;17(6):738-46.
72. Meric-Bernstam F et al. Pertuzumab plus trastuzumab for *HER2*-amplified metastatic colorectal cancer (MyPathway): an updated report from a multicentre, open-label, Phase 2a, multiple basket study. *Lancet Oncol.* 2019;20(4):518-30.
73. Gupta R et al. Pertuzumab plus trastuzumab (P+ T) in patients (Pts) with colorectal cancer (CRC) with *ERBB2* amplification or overexpression: results from the TAPUR Study. *J Clin Oncol.* 2020;38(Suppl 4):132.
74. Strickler J et al. Trastuzumab and tucatinib for the treatment of *HER2* amplified metastatic colorectal cancer (mCRC): initial results from the MOUNTAINEER trial. *Ann Oncol.* 2019;30(Suppl 5):v200.
75. Siena S et al. Trastuzumab deruxtecan (DS-8201) in patients with *HER2*-expressing metastatic colorectal cancer (DESTINY-CRC01): a multicentre, open-label, Phase 2 trial. *Lancet Oncol.* 2021;22(6):779-89.
76. Southwest Oncology Group. S1613, trastuzumab and pertuzumab or cetuximab and irinotecan hydrochloride in treating patients with locally advanced or metastatic *HER2/Neu* amplified colorectal cancer that cannot be removed by surgery. NCT03365882. <https://clinicaltrials.gov/ct2/show/NCT03365882>.
77. Daiichi Sankyo, Inc. Trastuzumab deruxtecan in participants with *HER2*-overexpressing advanced or metastatic colorectal cancer (DESTINY-CRC02). NCT04744831. <https://clinicaltrials.gov/ct2/show/NCT04744831>.
78. Amatu A et al. *NTRK* gene fusions as novel targets of cancer therapy across multiple tumour types. *ESMO Open.* 2016;1(2):e000023.
79. Nakagawara A. Trk receptor tyrosine kinases: a bridge between cancer and neural development. *Cancer Lett.* 2001;169(2):107-14.
80. Vaishnavi A et al. TRKing down an old oncogene in a new era of targeted therapy. *Cancer Discov.* 2015;5(1):25-34.
81. Créancier L et al. Chromosomal rearrangements involving the *NTRK1* gene in colorectal carcinoma. *Cancer Lett.* 2015;365(1):107-11.
82. Drilon A et al. Efficacy of larotrectinib in *TRK* fusion-positive cancers in adults and children. *N Engl J Med.* 2018;378(8):731-9.
83. Sartore-Bianchi A et al. Sensitivity to entrectinib associated with a novel *LMNA-NTRK1* gene fusion in metastatic colorectal cancer. *J Natl Cancer Inst.* 2016;108(1):d1jv306.
84. Pietrantonio F et al. *ALK*, *ROS1*, and *NTRK* rearrangements in metastatic colorectal cancer. *J Natl Cancer Inst.* 2017;109(12):d1jx089.
85. Yakirevich E et al. Oncogenic *ALK* fusion in rare and aggressive subtype of colorectal adenocarcinoma as a potential therapeutic target. *Clin Cancer Res.* 2016;22(15):3831-40.
86. Amatu A et al. Novel *CAD-ALK* gene rearrangement is druggable by entrectinib in colorectal cancer. *Br J Cancer.* 2015;113(12):1730-4.