



# Immunotherapeutic Strategies Based on CAR-T Cells in Hepatocellular Carcinoma

Author:	*Negar Hemmati <sup>1,2</sup>  1. Department of Microbiology and Immunology, University of Tehran, Iran 2. Shiraz University of Medical Sciences, Iran *Correspondence to <a href="mailto:Negar.hemmati@ut.ac.ir">Negar.hemmati@ut.ac.ir</a>
Disclosure:	The author has declared no conflicts of interest.
Received:	19.05.25
Accepted:	22.10.25
Keywords:	Chimeric antigen receptors (CAR), hepatocellular carcinoma (HCC), immunotherapy, T lymphocytes, tumour microenvironment (TME).
Citation:	EMJ Oncol. 2025;13[1]:140-153. <a href="https://doi.org/10.33590/emjoncol/TABT3057">https://doi.org/10.33590/emjoncol/TABT3057</a>

## Abstract

Hepatocellular carcinoma (HCC) is the sixth leading cause of cancer-related mortality worldwide. Despite the availability of therapeutic options such as surgical resection, radiofrequency ablation, molecular-targeted agents, and liver transplantation, HCC shows a poor prognosis and limited responsiveness to conventional treatments. The tumour immune microenvironment (TME) influences key processes in HCC, including selection pressure on tumour cells, immune evasion, tumour evolution, treatment resistance, and recurrence. Among immune components within the TME, T cells, dominant among tumour-infiltrating lymphocytes (TIL), exert both suppressive and promotive effects on tumour growth. Thus, T cell-mediated immune responses are fundamental to cancer surveillance and elimination. Research highlights the crucial role of TILs in HCC prognosis, pathogenesis, and immunotherapy. Subpopulations such as Foxp3+ regulatory T cells, CD8+ cytotoxic T cells, and CD3+/CD4+ helper T cells show complex and often contrasting roles. However, the TME often induces T cell exhaustion or dysfunction, facilitating tumour progression and immune evasion. Understanding immune dysregulation is vital for improving anti-tumour immunity and refining T cell function. This review examines TIL subpopulation roles in HCC, emphasising their plasticity and therapeutic relevance. It also covers emerging T cell-based immunotherapies, especially TIL-based adoptive transfer and CAR-T cell therapy, both showing promise in preclinical and early clinical trials. These novel approaches offer new hope for enhancing immune-driven tumour eradication and improving HCC outcomes.

## Key Points

1. Hepatocellular carcinoma remains a leading cause of cancer related mortality worldwide, and its immunosuppressive tumour microenvironment limits the efficacy of conventional treatments, highlighting the need for advanced immune based therapeutic strategies.

2. This narrative review explores recent advances in T cell-based immunotherapies, including tumour-infiltrating lymphocyte and chimeric antigen receptor-T cell approaches, and their emerging role in treating hepatocellular carcinoma.

3. Integrating chimeric antigen receptor-T or tumour-infiltrating lymphocyte therapies with immune checkpoint blockade and microenvironment modulation may enhance antitumour efficacy, offering new personalised and combinatorial treatment avenues for patients with advanced hepatocellular carcinoma.

## INTRODUCTION

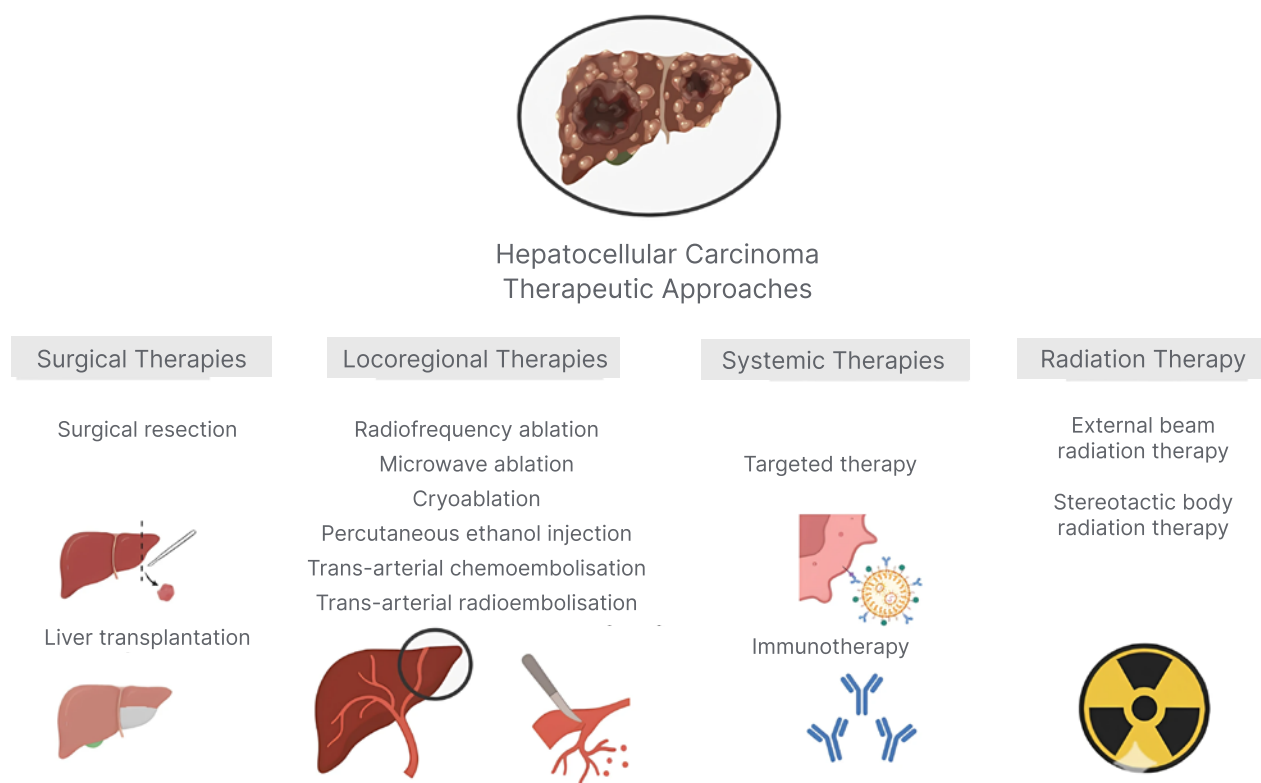
Hepatocellular carcinoma (HCC) is the sixth leading cause of cancer-related mortality worldwide.<sup>1</sup> Despite therapeutic modalities such as surgical resection, radiofrequency ablation, molecular-targeted agents, and liver transplantation being available, HCC is frequently associated with a poor prognosis and limited responsiveness to conventional interventions.<sup>2</sup> Therapeutic modalities vary based on the tumour characteristics, hepatic activity, and physiological condition of the patients.<sup>3</sup> To prevent liver parenchyma damage, various mechanisms function to inherently avert undesirable immune responses elicited by contact with microbial antigens and conserved molecular motifs termed danger or pathogen-associated molecular patterns, rendering the liver a predominantly immune-suppressive microenvironment.<sup>4</sup> The liver immune microenvironment exhibits operational heterogeneity, characterised by the diverse roles of stromal cells such as liver sinusoidal endothelial cells, hepatic stellate cells, liver resident macrophages (Kupffer cells), and various components of the adaptive immune response, such as CD4+ and CD8+ T lymphocytes and natural killer (NK) cells (Figure 1).<sup>5</sup>

Immunotherapy treatment methods developed in recent years, such as monoclonal antibodies that simultaneously suppress the programmed cell death protein 1 (PD-1) axis and cytotoxic T lymphocyte-associated protein 4 (CTLA-4), could stimulate anti-tumour immunity.<sup>6,7</sup> Furthermore, locoregional treatments such as trans-arterial chemoembolisation and ablation produce cell death locally and promote CD8+ cell infiltration into the tumour microenvironment, which supports the use of combination PD-1 blockers.<sup>8</sup> In

order to circumvent immune escape and trigger anti-tumour responses, autologous T cell transplantation also entails *ex vivo* activation of mixed T cell/NK cells through cytokine-induced stimulation and reinfusion into the patient. Additionally, *ex vivo* stimulation of dendritic cells can be used with anti-tumour vaccines against immunodominant peptides of oncofetal proteins to enhance efficient antigen presentation (Figure 2).

T cells are an important part of the immune system, and cancer-specific T lymphocytes can experience a gradual decrease in functional activity once subjected to continuous antigenic stimulus. This decrease exhibits several characteristics, such as persistent generation of restrictive receptors, reduced cytokine production, modified metabolic characteristics, inadequate memory re-establishment, and unique transcriptional and epigenomic characteristics, a dynamic phenomenon commonly referred to as T cell exhaustion.<sup>9</sup> The present emphasis of immunotherapy is on reversing T cell depletion, seeking to mitigate the detrimental impacts of continuous antigenic induction and offering a pathway for the therapy of HCC. T cells undergo terminal differentiation during cancer progression due to sustained antigenic induction, resulting in T cell exhaustion.<sup>10</sup> Naive CD8+ T cells can differentiate into various degrees of exhaustion, characterised by diminishing working and proliferative abilities, which can eventually suffer from overstimulation-induced cell death.<sup>11,12</sup> A thorough understanding of the procedure and its fundamental processes is essential to successfully preventing T cell exhaustion. It is necessary to relocate T lymphocytes from a defective developmental pathway before they reach terminal exhaustion.<sup>13</sup> In recent

**Figure 1: Current therapeutic modalities for hepatocellular carcinoma.**



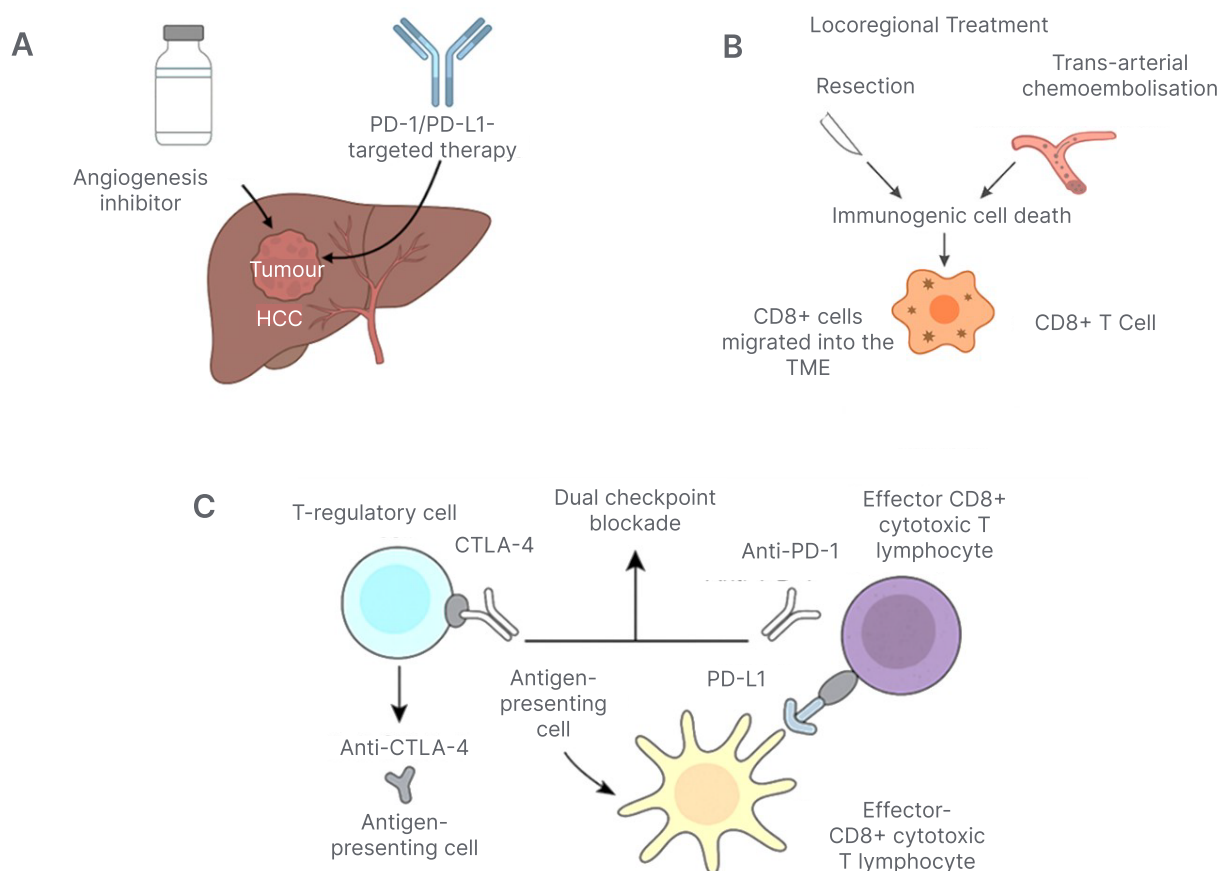
years, the efficacy of PD-1/L1 targeted therapy using monoclonal antibodies has led to promising outcomes in solid tumours, such as HCC, and prompted growing efforts to focus on immunotherapy to reactivate T cell function for tumour treatment.<sup>14</sup>

In this light, adoptive cell transfer, like tumour-infiltrating lymphocytes (TIL), and chimeric antigen receptor (CAR)-T cell therapy, not only modulates immune equilibrium within the tumour microenvironment but has also demonstrated significant efficacy in treating various malignancies, thereby heightening clinical researchers' interest in this area.<sup>15</sup> Nonetheless, the use of both adoptive cell treatment and immune-targeted therapies will bring about the challenge of T cell exhaustion.<sup>16-18</sup> This study will outline the significant T cell subsets found in HCC and examine existing T cell-based immunotherapies in HCC, focusing on TILs and CAR-T cell-based therapies.

## SUBTYPES OF TUMOUR-INFILTRATING T CELLS IN HEPATOCELLULAR CARCINOMA

### CD8+ Cytotoxic T Lymphocytes

CD8+ cytotoxic T lymphocytes are crucial for mediating antigen-specific cytotoxicity through tumour-associated antigens presented by antigen-presenting cells, including dendritic cells. The cytotoxic T cell receptor (TCR) binds to the tumour antigen presented by major histocompatibility complex class I, and becomes mature. Then CD8+ mature T lymphocytes induce cytotoxicity by immediate destruction of neoplastic cells and the release of enzymes along with destructive mediators, including perforin/granzyme B, interferon gamma (IFN $\gamma$ ), and TNF $\alpha$ .<sup>19</sup> Perforin disrupts the integrity of the tumour cell membrane, while granzyme B triggers apoptosis through caspase activation, via interaction with Fas ligand expressed by CD8+ T lymphocytes with Fas receptors on cancerous cells, and initiates the caspase signalling axis. TNF induces inflammatory responses and

**Figure 2: Integrated immunotherapeutic strategies combining checkpoint blockade and locoregional therapies.**

**A)** Schematic depiction of the combination of angiogenesis inhibitors and PD-1/PD-L1-targeted therapy. **B)** Locoregional treatments, including resection and trans-arterial chemoembolisation, serve as locoregional stimulants of immunogenic cell death and promote CD8+ cells to migrate into the tumour microenvironment, hence justifying the use of a combination PD-1 blocker. **C)** Concurrent blocking of CTLA-4 and the PD-1 pathway with monoclonal antibodies. The impact of dual checkpoint on T-cell immune reconstitution is shown, with CTL-4 primarily influencing T-regulatory cells and antigen-presenting cells, while PD-1 affects effector CD8+ cytotoxic lymphocytes. CTLA-4: cytotoxic T-lymphocyte-associated protein 4; PD-1: programmed death-1; PD-L1: programmed death-ligand 1; TME: tumour microenvironment.

apoptosis by activating caspase-mediated signalling pathways in tumour cells.<sup>20-22</sup>

Clinically, considerably higher levels of CD8+ T cells are found in HCC, particularly within or around neoplastic tissue, identifying them as key components of tumour-infiltrating lymphocytes for monitoring HCC development.<sup>23</sup> Cytotoxic lymphocytes, along with an elevated CD4+/CD8+ ratio, are correlated with a reduced risk of relapse following hepatic transplantation.<sup>24</sup> The effectiveness of anti-tumour immunological response requires not only antigen presentation and T cell priming, but also efficient infiltration and persistence

of effector T cells within the tumour parenchyma. Immune-desert phenotype, immune-excluded phenotype, and inflamed phenotype are three tumour penetration types based on pathology investigations. The amount and quality of invading T cells both enhance anti-cancer effectiveness.<sup>25,26</sup>

Previous research has proved that the proportion of infiltrating CD8+ T cells is considerably lower in HCC tissues compared to non-cancerous liver tissue.<sup>27</sup> Furthermore, the immunosuppressive function of regulatory T cells (Tregs) and CD8+ T cells within the tumour microenvironment often exhibits functional

defects, which are evident through exhaustion and immune tolerance. This pathologic state entails reduced cytotoxic activity and elevated expression of inhibitory immune checkpoint receptors, such as PD-1, LAG-3, TIM-3, and CTLA-4. The loss of CD8<sup>+</sup> T cells is also promoted by the immunosuppressive activities of Tregs, which together compromise the effectiveness of the anti-tumour immune response. Inside the tumour microenvironment, CTLs may undergo activation-induced cell death, a process which is induced by chronic antigen stimulation and inflammatory stress.<sup>12</sup> Empirical evidence based on murine models of HCC has demonstrated increased rates of apoptosis in intratumoral CD8<sup>+</sup> T cells, thus substantiating the belief that the tumour microenvironment is actively engaged in facilitating T cell dysfunction and consequent loss of cells. Collectively, these findings demonstrate that CD8<sup>+</sup> T cells are crucial for the mediation of anti-tumour immunity; their quantity, quality, and longevity in the TME ultimately determine their therapeutic potential in HCC.<sup>28-30</sup>

### Regulatory T Cells

Regulatory T cells are a distinct subset of CD4<sup>+</sup> T lymphocytes that play a critical role in the induction of immune tolerance and are critical in maintaining immune homeostasis. In the tumour microenvironment of HCC, the existence of Tregs facilitates immune evasion by cancer cells by suppressing effective anti-tumour immune responses.<sup>31</sup> In tumour tissues, Tregs are generally divided into two major subsets: natural Tregs and inducible Tregs. Natural Tregs with the CD4<sup>+</sup>CD25<sup>+</sup>FOXP3<sup>+</sup> phenotype are the prominent subset and play a critical role in the maintenance of immunological homeostasis through the regulation of peripheral inflammation.<sup>32,33</sup> These cells possess the capacity to exert immune suppressive function in the tumour via the Fas-FasL pathway, and importantly, induce apoptosis in natural killer cells and CD8<sup>+</sup> cytotoxic T lymphocytes in a granzyme B/perforin-dependent process.<sup>34,35</sup> Inducible Tregs, that may or may not express FOXP3, perform immunosuppressive activities by mainly secreting anti-inflammatory

cytokines IL-10 and transforming growth factor-TGF- $\beta$  (TGF- $\beta$ ), and by producing adenosine. All these events serve to inhibit the activation and functionality of effector B and T lymphocytes following contact with an antigen. While Tregs are essential under normal circumstances to prevent autoimmunity and regulate inflammatory responses, their inappropriate expansion and activity in the TME of HCC facilitate tumour immune evasion.<sup>36</sup> The immunosuppressive role of Tregs operates via multiple mechanisms, including the release of inhibitory cytokines, direct elimination of effector T cells through granzyme and perforin pathways, and the alteration of dendritic cell differentiation and their antigen-presenting ability.<sup>37</sup> The mechanism above entails the regulatory signalling mediated by inhibitory receptors like CTLA-4 and LAG-3.<sup>38</sup> Surprisingly, the concurrent activation of both the TCR and IL-2 receptor is central to the survival of Tregs and their immunosuppressive functions.<sup>39</sup> In HCC, Tregs predominantly undermine anti-tumour immunity through the inhibition of CD8<sup>+</sup> T cell activities. Mechanistic studies have also shown that Tregs derived from HCC can downregulate the co-stimulatory molecules CD80 and CD86 on DCs in a CTLA-4-dependent manner, dampening antigen presentation and inhibiting effector T cell responses further.<sup>40,41</sup>

Clinical observations have revealed significantly increased percentages of CD4<sup>+</sup>CD25<sup>+</sup> Tregs in peripheral blood as well as liver tissues of patients with HCC compared to healthy controls. Such an increase is linked with disease progression and poor prognosis. Additionally, a high intratumoural Treg-to-CTL ratio has been identified as a prognostic factor, where higher ratios are correlated with poorer overall survival and disease-free survival.<sup>42</sup> In a study of 19 patients with advanced HCC, lower frequencies of PD-1<sup>+</sup> and FOXP3<sup>+</sup> Tregs were correlated with increased survival after sorafenib treatment, suggesting their utility as predictive biomarkers. Additionally, evidence exists that Treg accumulation can be causally responsible for HCC recurrence following liver transplantation through a CXCL10–



CXCR3 chemokine axis. In preclinical models, depletion of Tregs using monoclonal antibodies against CD25 has been effective at preventing HCC progression, highlighting a potential therapeutic avenue that involves reprogramming the immunosuppressive TME.<sup>43,44</sup>

### Th1 and Th2 Helper T Cells

T helper cells are a heterogeneous population of CD4<sup>+</sup> T lymphocytes that orchestrate immune responses by secreting various cytokine profiles.<sup>45,46</sup> Th1 and Th2 cells are two principal subsets among them, which have different cytokine secretion patterns and immunological roles. Th1 cells are mainly responsible for producing IFN $\gamma$ , IL-2, IL-22, and TNF $\beta$ , whereas Th2 cells are characterised by the secretion of IL-4, IL-5, IL-9, IL-10, and IL-13.<sup>47,48</sup> These polarised subsets play complementary but often opposite functions: Th1 cells play a central role in inducing cell-mediated immunity and delayed-type hypersensitivity, whereas Th2 cells induce humoral immune responses and are involved in regulating the production of antibodies.<sup>49-51</sup> In HCC, the functional distinction between Th1 and Th2 cells is crucial for determining the nature of the TME and the course of the disease. Th1 cells, on recognising peptide-major histocompatibility complex class II complexes and interacting with co-stimulatory molecules, produce pro-inflammatory cytokines like IFN $\gamma$ , which are accountable for the recruitment and activation of CD8<sup>+</sup> CTLs in tumour tissue.<sup>52,53</sup> This Th1-induced immune activation enables productive anti-tumour immunity. The shift from Th1-dominant to Th2-dominant cytokine milieu is frequently observed in HCC and signifies the setup of an immunosuppressive TME.<sup>54,55</sup> While Th1-associated cytokines IL-1 $\alpha$ , IL-1 $\beta$ , IL-2, and IFN $\gamma$  are typically associated with good clinical outcomes and improved prognosis, overexpression of Th2 cytokines IL-4, IL-5, and IL-10 is correlated with more invasive tumour behaviour and poor prognosis. This Th1-to-Th2 transition has been regarded as a hallmark of immune evasion in HCC.<sup>56,57</sup> Moreover, increased circulating Th2 cells have been associated with progressed HCC and decreased efficacy of treatment options available. Research demonstrated

that individuals with lower Treg levels at baseline exhibit an improved Th1/Th2 ratio prior to trans-arterial chemoembolisation, a widely applied locoregional therapy for HCC. Importantly, a higher Th1/Th2 ratio was linked to more intense anti-tumour immunity and prolonged survival rates. Although cancer vaccines targeting helper T cells have reported promising activity in a number of malignancies, the specific contribution of Th1 responses to promoting effector T cell activation in HCC remains ill-defined. Further research is required to establish the therapeutic utility of adjusting the Th1/Th2 balance as a strategy to reverse immunosuppression and enhance anti-tumour immunity in HCC.<sup>58</sup>

## CRITICAL SIGNALLING PATHWAYS MODULATING TUMOUR-INFILTRATING LYMPHOCYTES IN HEPATOCELLULAR CARCINOMA

### TGF $\beta$ Signalling

The TGF $\beta$  family includes TGF $\beta$ s, activins, inhibins, bone morphogenetic proteins, and growth and differentiation factors.<sup>59,60</sup> The initiation of this axis begins with the attachment of ligands to the extracellular domains of TGF $\beta$  Type I and Type II receptors (T $\beta$ RI and T $\beta$ RII), stimulating both suppressor of mothers against decapentaplegic homolog (SMAD)-reliant and SMAD-non-reliant axis to elicit the downstream pathway. The TGF $\beta$  axis is implicated in almost every phase of tumorigenesis in HCC.<sup>61</sup> The TME exhibited elevated levels of TGF $\beta$  in both cancer cells and several immunological cells. In the first phase of HCC, TGF $\beta$  inhibited the growth of pre-malignant hepatocytes. However, in the developed phase of HCC, it facilitated cancer growth by modulating immunological cells, including Tregs, CTLs, TAMs, and NKs.<sup>62</sup> TGF $\beta$  may up-regulate FOXP3 in CD4<sup>+</sup>CD25-naïve T cells by promoting the SMAD-reliant axis, facilitating the development of Tregs. Tregs from peripheral blood penetrated HCC tumour tissues, thereby inhibiting the therapeutic efficacy of CTLs. Moreover, HCC cells demonstrate elevated levels of TGF $\beta$ , thus underlying the upregulation of PD-1 levels on CD8<sup>+</sup> CTLs. CTLs subsequently attach to PD-L1 on malignant cells and

antigen-presenting cells, resulting in CTL exhaustion.<sup>63,64</sup> TGF $\beta$  activation may regulate the innate immune system by inhibiting NK cells directly and facilitating M2 macrophage polarisation to promote immune evasion. TGF $\beta$  signalling pathways were extensively implicated in the control of TILs, ultimately facilitating HCC progression.<sup>61,62</sup>

### JAK/STAT3 Signalling

The STAT protein family has seven members, including STAT1, STAT2, STAT3, STAT4, STAT5A, STAT5B, and STAT6. STAT3 has been the subject of several studies concerning HCC management. It was believed to be triggered swiftly and temporarily in normal cells by various mediators, including IL-6/IL-10, IFNs, and growth factors like vascular endothelial growth factor. Nonetheless, it was consistently and aberrantly activated in neoplastic cells.<sup>65-68</sup> Recent research indicated that phosphorylated STAT3 was present in approximately 60% of HCC samples, correlating with a worse prognosis. The STAT3 axis primarily promotes tumorigenesis by modulating dendritic cells and developing tumour-associated macrophages, NK cells, and Tregs.<sup>68,69</sup> In DCs, IL-6 released through HCC cells and hepatic carcinoma-associated fibroblasts can attach to the IL-6 receptor on DCs, activating JAK and initiating the downstream phosphorylation STAT3 axis, which inhibits T cell growth and enhances Tregs expansion.<sup>70</sup> Furthermore, the stimulation of STAT3 has been shown to suppress the activating receptor NKG2D on NK cells and its associated ligands, major histocompatibility complex class I-related protein A/B on malignant cells, hence obstructing NK cell stimulation and leading to impaired immune surveillance in HCC.<sup>71</sup> Inhibition of STAT3 signalling in HCC may reactivate NK cells to perform anti-tumour functions via modifying cytokine levels in the TME, specifically by decreasing IL-10 levels. Additionally, the IL-6-STAT3 axis promoted tumorigenesis by diminishing M1 macrophage polarisation and enhancing M2 differentiation of TAM. STAT3 activation affected many immunological cells, contributing to the progression of HCC.<sup>72,73</sup>

### Wnt/ $\beta$ -Catenin Signalling

The Wnt signalling pathway becomes active when Wnt proteins engage with their specific membrane-bound receptors, initiating a cascade of intracellular events. One key player in this pathway is  $\beta$ -catenin, a protein usually associated with cell-to-cell adhesion or found in the cytoplasm. Upon pathway activation,  $\beta$ -catenin accumulates and translocates to the nucleus, where it influences gene expression. In HCC, persistent activation of the Wnt/ $\beta$ -catenin axis has been frequently observed and is linked to immune evasion. This activation impairs the infiltration and function of tumour-infiltrating lymphocytes, especially by reducing dendritic cell presence, which plays a crucial role in antigen presentation and T cell priming. Consequently, this weakens the recruitment and cytotoxic function of CD8<sup>+</sup> T cells. Chronic stimulation of this axis also contributes to cytotoxic T lymphocyte dysfunction, ultimately fostering an immunosuppressive environment favourable to tumour progression. Overall, the aberrant Wnt/ $\beta$ -catenin signalling in HCC is a critical factor in disrupting immune surveillance and supporting tumour immune escape mechanisms (Table 1).<sup>44,74-78</sup>

## TUMOUR-INFILTRATING LYMPHOCYTE THERAPY IN HEPATOCELLULAR CARCINOMA

TIL therapy is a promising immunotherapeutic approach for HCC, leveraging the patient's own tumour-reactive T cells. These lymphocytes are isolated from the tumour, expanded *in vivo*, often with IL-2, and reintroduced after lymphodepletion to enhance their persistence and efficacy.<sup>79-81</sup> This strategy boosts the body's immune response by restoring and amplifying tumour-specific T cell activity. However, challenges remain, including difficulty in isolating sufficient TILs from the liver's immunosuppressive environment and ensuring effective tumour antigen recognition. Current research aims to optimise expansion techniques, tailor conditioning regimens, and overcome immune resistance. Continued refinement is essential to establish TIL therapy as a reliable treatment option for HCC.<sup>82-86</sup>

**Table 1: Critical immunoregulatory signalling pathways affecting tumour-infiltrating lymphocytes in hepatocellular carcinoma.**

Signalling Pathway	Ligand Source	Receptor	Intracellular Pathway (in Cell Type)	Immune Cells Affected	Immunological Outcome	References
TGFβ	HCC cells, immune cells in TME	TβRI/TβRII	SMAD-dependent and independent in T cells, NKs, and macrophages	Tregs, CTLs, NKs, TAMs	↑ Tregs via FOXP3 upregulation, ↓ NK cell cytotoxicity, ↑ PD-1 expression on CTLs, ↑ M2 macrophage polarisation	(63-66)
IL-6/STAT3	HCC cells, hCAFs	IL-6R	JAK → STAT3 in DCs, macrophages, NKs	DCs, Tregs, NKs, TAMs	↑ Tregs through DC modulation, ↓ CD8+ T cell priming, ↓ NKD2G receptor on NK cells, ↑ M2 macrophages	(67-74)
Wnt/β-Catenin	HCC cells	Frizzled/ LRP5/6	β-catenin nuclear translocation in tumour cells	DCs, CD8+ T cells (indirectly)	↓ DC recruitment and antigen presentation, ↓ CD8+ T cell infiltration, ↑ and immune evasion	(46, 76-80)

CTL: cytotoxic T lymphocyte; DC: dendritic cell; hCAF: hepatic cancer-associated fibroblast; HCC: hepatocellular carcinoma; NK: natural killer; PD-1: programmed death-1; TAM: tumour-associated macrophage; TGFβ: transforming growth factor-β; TME: tumour microenvironment; Tregs: regulatory T cells.

RECENT ADVANCES AND CLINICAL APPLICATIONS

Recent advances in tumour-infiltrating lymphocyte therapy have positioned it as a promising immunotherapeutic option for HCC. Early-phase clinical trials, such as the Phase I trial of BST02, a TIL-derived product, have demonstrated encouraging outcomes in patients with advanced HCC. Notably, combining TILs with immune checkpoint inhibitors, particularly PD-1/PD-L1 blockade, appears to overcome the immunosuppressive liver microenvironment, leading to more durable antitumor responses compared to TIL monotherapy.<sup>87-90</sup> To further improve efficacy, researchers have employed adjunct strategies such as IL-2 to enhance TIL proliferation and function. Pre-treatment lymphodepletion with agents like cyclophosphamide has also shown benefit by facilitating TIL engraftment and expansion. However, the tumour microenvironment, characterised by high PD-L1 expression and suppressive cytokines, continues to limit therapeutic success. To address this,

novel approaches, including oncolytic virotherapy and lymphodepleting regimens, are being evaluated for their synergistic potential to reprogram the TME.<sup>91,92</sup> In parallel, TIL engineering strategies such as CAR expression have shown promise in preclinical models, offering increased tumour specificity. Phase II trials report that combination regimens remain safe and tolerable, even in patients with chronic hepatitis B, although tumour regression is modest in this subgroup. Overall, the future of TIL therapy in HCC lies in personalised approaches informed by immune profiling and the strategic integration of combination therapies. These advancements may ultimately establish TILs as a foundational component of immunotherapy in liver cancer (Table 2).<sup>25,93,94</sup>

BIOMARKERS AND PREDICTIVE INDICATORS

Biomarker identification is pivotal for optimising patient selection and improving



**Table 2: Recent studies on tumour-infiltrating lymphocytes in hepatocellular carcinoma.**

Study/Strategy	Description	Phase	References
TIL+Checkpoint Inhibitors (Atezolizumab+Bevacizumab)	Combination of TIL therapy with PD-1/PD-L1 inhibitors to reverse T cell exhaustion in the tumour microenvironment	Phase II	(89-92, 95)
BST02 Trial	TIL-derived product (BST02) for patients with HCC	Phase I	(85, 88, 96)
TIL+IL-2 Cytokine Support	Using IL-2 to improve the survival and function of TILs	Preclinical and Observational	(81, 82, 84, 85)
CAR-T in HCC	Engineering T cells to express chimeric antigen receptors targeting tumour antigens	Preclinical/Early Clinical	(93, 94, 97-99)
Oncolytic Virus+ Lymphodepletion+TIL	Modifying the tumour microenvironment using oncolytic viruses and chemotherapy to enhance TIL efficacy	Under Investigation	(83, 84, 94, 100)
TIL+ICIs in Patients with Chronic HBV	Assessing the safety and effectiveness of TIL+checkpoint inhibitors in patients with chronic hepatitis B	Phase II	(90, 91, 101)
Personalised TIL Therapy Based on Immune Profile	Tailored TIL treatments based on individual patient immune signatures	In Development	(101-104)

HBV: hepatitis B virus; HCC: hepatocellular carcinoma; PD-1: programmed death-1; PD-L1: programmed death-ligand 1; TIL: tumour-infiltrating lymphocytes.

the therapeutic efficacy of TIL therapy in HCC. Accumulating evidence suggests that a high frequency of CD8<sup>+</sup> TILs within tumour tissues is associated with superior clinical outcomes, particularly when TIL therapy is combined with immune checkpoint inhibitors. Tumours enriched in CD8<sup>+</sup> T cells may demonstrate enhanced immune responsiveness, making them more amenable to TIL-based interventions. Nonetheless, the predictive roles of additional immune biomarkers, such as PD-L1 expression and TCR diversity, remain under active investigation. Furthermore, both liver functional status and HCC staging are important considerations in determining patient eligibility. Individuals with preserved liver function and early-stage disease are more likely to benefit from adoptive TIL therapy.<sup>95,101</sup> Recent clinical studies have yielded variable results regarding its therapeutic impact. A Phase I trial published in 2022 demonstrated the technical

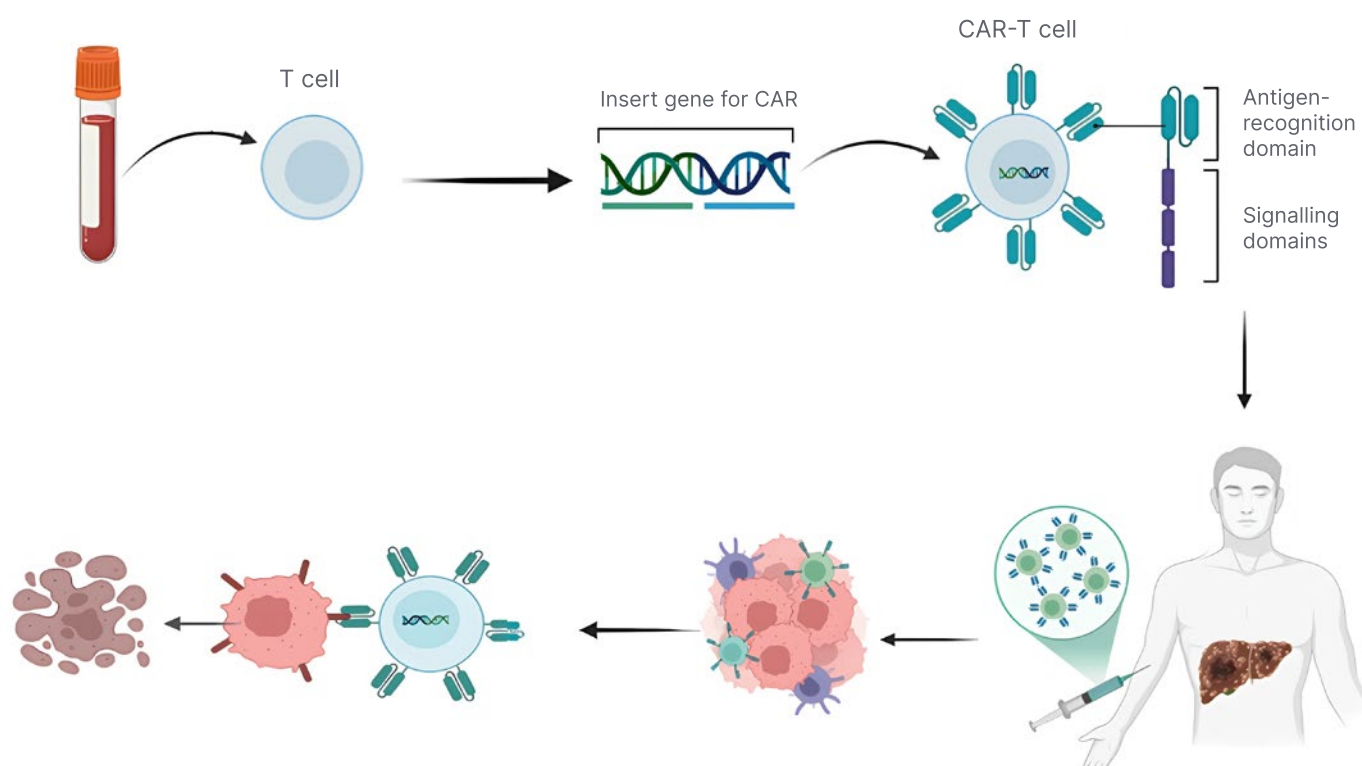
feasibility of TIL isolation, expansion, and reinfusion in patients with HCC; however, clinical responses were limited, particularly in heavily pre-treated individuals.<sup>102</sup> More recent investigations have evaluated the combination of TIL therapy with PD-1 inhibitors, showing promising early efficacy, especially in tumours with high PD-L1 expression.<sup>96,103</sup> These findings underscore the need for improved patient stratification strategies and potentially the incorporation of additional combination therapies to enhance response. Moreover, TME-induced immunosuppression continues to be a major barrier to TIL functionality in HCC. Observational studies confirm that overcoming the TME's suppressive effects is critical for maximising TIL efficacy, aligning with previous research emphasising the central role of the TME in modulating immune responses in liver cancer.<sup>105,106</sup>

## ADVANCES IN CAR-T CELL THERAPY FOR HEPATOCELLULAR CARCINOMA

CAR-T cell therapy is being actively explored for HCC, with recent innovations addressing key challenges, such as immune evasion, tumour heterogeneity, and a suppressive microenvironment. Glypican-3 (GPC3), highly expressed in HCC and rarely in normal tissues, is a leading CAR-T target. Early clinical results show anti-tumour activity, though efficacy is limited by immunosuppression (Figure 3). To counter this, GPC3-CAR-T cells are engineered to release PD-1 or TGF $\beta$  inhibitors, enhancing persistence and function. Co-administering checkpoint inhibitors (e.g., anti-PD-1) also helps reduce T cell exhaustion.<sup>97,104,107</sup> Tandem CARs (TanCARs), recognising GPC3 and AFP, boost tumour detection and reduce immune escape. SynNotch CARs use

a two-step activation model, first sensing one antigen, then triggering CAR expression via another, ensuring tumour specificity and minimising off-target effects. Both methods improve tumour targeting in HCC models.<sup>44,98,108</sup> HCC's immunosuppressive environment hinders CAR-T activity. New CAR-Ts are modified to secrete cytokines like IL-15, supporting T cell survival. Targeting vascular endothelial growth factor improves infiltration, while combining with TGF $\beta$  inhibitors reprogrammes the microenvironment to be more immune-permissive.<sup>108</sup> Additionally, Universal CARs employ a modular 'lock-and-key' system, allowing redirection to multiple tumour antigens. This approach addresses tumour heterogeneity and lowers production costs, improving feasibility for personalised treatments.<sup>99,109</sup> CAR-T efficacy is improved by blocking inhibitory receptors such as PD-1, LAG-3, and TIM-3. Inhibiting CD39,

**Figure 3: Schematic representation of CAR-T cell therapy in liver cancer.**



Peripheral blood T cells are isolated and genetically engineered to express CAR. The modified CAR-T cells are expanded and re-infused into the patient, where they recognise and kill tumour cells in the liver through antigen-specific cytotoxicity.

CAR: chimeric antigen receptor.

a driver of adenosine-mediated suppression, restores function in HCC. Combining CAR-T with anti-LAG-3 antibodies further revives exhausted T cells.<sup>110-112</sup> Engineered cytokine receptors like 4/7 ICR convert suppressive IL-4 signals into stimulatory IL-7-like effects. Advanced 4/21 ICR CAR-T cells adopt a Th17-like phenotype under IL-4, enhancing anti-tumour activity and persistence in HCC.<sup>113,114</sup> To prevent recurrence, CAR-T cells are engineered to produce IL-7 or IL-21, promoting memory cell development. Small-molecule agents like T-lymphokine-activated killer cell-originated protein kinase inhibitors and CAR signalling modifications improve survival. Combining CAR-T with chemo, radiation, or oncolytic viruses further boosts immune response and tumour antigen visibility.<sup>115,116</sup>

## CONCLUSION

Recent evidence has revealed that cancer progression is not only driven by local immune and molecular alterations but also by systemic neuroendocrine regulation. Tumours can hijack neuroendocrine signalling pathways to manipulate host homeostasis, thereby creating conditions favourable for their growth and immune evasion. Through the release of stress-related hormones and neuropeptides, the tumour can modulate immune cell activity, suppress cytotoxic CD8<sup>+</sup> T lymphocytes, and promote Treg expansion within the tumour microenvironment. This neuroendocrine-immune interaction leads to impaired anti-tumour immunity and contributes to T cell exhaustion, a key obstacle to effective immunotherapy. Understanding how HCC exploits

neuroendocrine mechanisms to suppress immune surveillance may open new avenues for combination therapies that integrate immunomodulation with neuroendocrine targeting to restore both immune balance and systemic homeostasis.<sup>100</sup> Recent advances in immunotherapy have rekindled hope for HCC treatment, with T cell exhaustion receiving more attention. This pathological condition is one of the driving forces for HCC development, but its causative mechanisms are only half understood and require more clinical investigations. Clarification of the molecular mechanisms of T cell exhaustion is essential for unravelling the immune microenvironment of HCC and developing new therapeutic interventions. Both TILs and CAR-T cells possess double-edged roles, both fighting cancer and potentially supporting tumour growth, via a complex network of signalling interactions within the tumour microenvironment. Further research is necessary to unveil the occult functions and mechanisms of TILs, particularly how they mould and are moulded by the evolving HCC environment. Longitudinal studies of the functions of TILs and CAR-T cells in HCC development will provide additional information on immune-tumour interactions and guide the development of next-generation immunotherapeutic approaches. As a narrative review, this study is limited by the lack of a systematic search strategy, which may introduce selection bias. Additionally, due to the rapidly evolving nature of immunotherapy research, some recent data may not have been captured at the time of writing. Finally, many referenced findings originate from early-phase or preclinical studies, which limits the strength and generalisability of the conclusions.

## References

- Samant H et al. Addressing the worldwide hepatocellular carcinoma: epidemiology, prevention and management. *J Gastrointest Oncol*. 2021;12(Suppl 2):S361-S373.
- Sachdeva M, Arora SK. Prognostic role of immune cells in hepatocellular carcinoma. *EXCLI J*. 2020;19:718-33.
- Jelic; ESMO Guidelines Working Group. Hepatocellular carcinoma: ESMO clinical recommendations for diagnosis, treatment and follow-up. *Ann Oncol*. 2009;DOI: 10.1093/annonc/mdp124.
- Zhou G et al. Immune suppressive checkpoint interactions in the tumour microenvironment of primary liver cancers. *Br J Cancer*. 2022;126(1):10-23.
- Fujita T, Narumiya S. Roles of hepatic stellate cells in liver inflammation: a new perspective. *Inflamm Regen*. 2016;36(1):1.
- Song Q et al. A study on the efficacy and Safety Evaluation of a novel PD-1/CTLA-4 bispecific antibody. *Immunobiology*. 2024;229(6):152844.
- Wang K et al. Combination anti-PD-1 and anti-CTLA-4 therapy generates waves of clonal responses that include progenitor-exhausted CD8<sup>+</sup> T cells. *Cancer Cell*. 2024;42(9):1582-97.
- Zhu H-D et al. Transarterial chemoembolization with PD-(L) 1 inhibitors plus molecular targeted therapies for hepatocellular carcinoma

- (CHANCE001). *Signal Transduct Target Ther.* 2023;8(1):58.
9. Catakovic K et al. T cell exhaustion: from pathophysiological basics to tumor immunotherapy. *Cell Commun Signal.* 2017;15(1):1.
  10. Zhu Y et al. Molecular insight into T cell exhaustion in hepatocellular carcinoma. *Pharmacol Res.* 2024;203:107161.
  11. Muthiah KA et al. Prospective evaluation of procalcitonin in sepsis in the Illawarra area of Australia: PEPSIA study. *Crit Care Resusc.* 2007;9(2):137-42.
  12. Dolina JS et al. CD8+ T cell exhaustion in cancer. *Front Immunol.* 2021;12:715234.
  13. Hu Y et al. Reversal of T-cell exhaustion: mechanisms and synergistic approaches. *Int Immunopharmacol.* 2024;138:112571.
  14. Sharafi F et al. A comprehensive review about the utilization of immune checkpoint inhibitors and combination therapy in hepatocellular carcinoma: an updated review. *Cancer Cell Int.* 2022;22(1):269.
  15. Aggeletopoulou I et al. Chimeric antigen receptor T cell therapy for hepatocellular carcinoma: where do we stand? *Int J Mol Sci.* 2024;25(5):2631.
  16. Gumber D, Wang LD. Improving CAR-T immunotherapy: overcoming the challenges of T cell exhaustion. *EBioMedicine.* 2022;77:103941.
  17. Kang K et al. T cell exhaustion in human cancers. *Biochim Biophys Acta Rev Cancer.* 2024;1879(5):189162.
  18. Li R et al. Cellular kinetics and biodistribution of adoptive t cell therapies: from biological principles to effects on patient outcomes. *AAPS J.* 2025;27(2):55.
  19. Abdou AG et al. Role of CD8 cytotoxic T lymphocytes in hepatocellular carcinoma: an immunohistochemical study. *Medical Journal of Cairo University.* 2019;87:4061-9.
  20. Annibaldi A, Meier P. Checkpoints in TNF-induced cell death: implications in inflammation and cancer. *Trends Mol Med.* 2018;24(1):49-65.
  21. Volpedo G et al. Chapter 4 - The Fas/ FasL pathway as a target for enhancing anticancer adoptive cell therapy. *Immunotherapy in Resistant Cancer: From the Lab Bench Work to Its Clinical Perspectives.* Elsevier. 2021;2:47-68.
  22. Metkar SS et al. Cytotoxic cell granule-mediated apoptosis: perforin delivers granzyme B-serglycin complexes into target cells without plasma membrane pore formation. *Immunity.* 2002;16(3):417-28.
  23. Chen Z et al. AI-based tumor-infiltrating lymphocyte scoring system for assessing HCC prognosis in patients undergoing liver resection. *JHEP Rep.* 2024;7(2):101270.
  24. Xin H et al. The CD68+ macrophages to CD8+ T-cell ratio is associated with clinical outcomes in hepatitis B virus (HBV)-related hepatocellular carcinoma. *HPB (Oxford).* 2021;23(7):1061-71.
  25. Zhang L et al. Immunotherapy for advanced hepatocellular carcinoma, where are we? *Biochim Biophys Acta Rev Cancer.* 2020;1874(2):188441.
  26. Mellman I et al. The cancer-immunity cycle: Indication, genotype, and immunotype. *Immunity.* 2023;56(10):2188-205.
  27. Ikeguchi M et al. CD8+ lymphocyte infiltration and apoptosis in hepatocellular carcinoma. *Eur J Surg Oncol.* 2004;30(1):53-7.
  28. Alim LF et al. Molecular mechanisms of tumour necrosis factor signalling via TNF receptor 1 and TNF receptor 2 in the tumour microenvironment. *Curr Opin Immunol.* 2024;86:102409.
  29. Huan L et al. Targeting tumor metabolism to augment CD8+ T cell anti-tumor immunity. *J Pharm Anal.* 2025;15(5):101150.
  30. Fu C et al. Manganese improves CD8+ T cell recruitment via cGAS-STING in hepatocellular carcinoma. *International Immunopharmacology.* 2024;143:113591.
  31. González-Navajas JM et al. The impact of Tregs on the anticancer immunity and the efficacy of immune checkpoint inhibitor therapies. *Front Immunol.* 2021;12:625783.
  32. Peterson RA. Regulatory T-cells: diverse phenotypes integral to immune homeostasis and suppression. *Toxicol Pathol.* 2012;40(2):186-204.
  33. Valencia X, Lipsky PE. CD4+ CD25+ FoxP3+ regulatory T cells in autoimmune diseases. *Nat Clin Pract Rheumatol.* 2007;3(11):619-26.
  34. Tanaka A, Sakaguchi S. Regulatory T cells in cancer immunotherapy. *Cell Res.* 2017;27(1):109-18.
  35. Vignali DA et al. How regulatory T cells work. *Nat Rev Immunol.* 2008;8(7):523-32.
  36. Sharma MD et al. Indoleamine 2, 3-dioxygenase controls conversion of Foxp3+ Tregs to TH17-like cells in tumor-draining lymph nodes. *Blood, The Journal of the American Society of Hematology.* 2009;113(24):6102-11.
  37. Schmidt A et al. Molecular mechanisms of treg-mediated T cell suppression. *Frontiers in immunology.* 2012;3:51.
  38. Zong Y et al. Regulation of Treg cells by cytokine signaling and co-stimulatory molecules. *Front Immunol.* 2024;15:1387975.
  39. Shouse AN et al. Interleukin-2 receptor signaling acts as a checkpoint that influences the distribution of regulatory T cell subsets. *iScience.* 2024;27(12):111248.
  40. Chen X et al. Tumor-derived CD4+ CD25+ regulatory T cells inhibit dendritic cells function by CTLA-4. *Pathology-Research and Practice.* 2017;213(3):245-9.
  41. Tekguc M et al. Treg-expressed CTLA-4 depletes CD80/CD86 by trogocytosis, releasing free PD-L1 on antigen-presenting cells. *Proceedings of the National Academy of Sciences.* 2021;118(30):e2023739118.
  42. Yan W et al. CXCL10 mediates CD8+ T cells to facilitate vessel normalization and improve the efficacy of cetuximab combined with PD-1 checkpoint inhibitors in colorectal cancer. *Cancer Letters.* 2023;567:216263.
  43. Zhu Y, Qin L-X. Strategies for improving the efficacy of immunotherapy in hepatocellular carcinoma. *Hepatobiliary & Pancreatic Diseases International.* 2022;21(5):420-9.
  44. Muhammed TM al. T lymphocyte-based immune response and therapy in hepatocellular carcinoma: focus on TILs and CAR-T cells. *Naunyn-Schmiedeberg's Archives of Pharmacology.* 2025:1-18.
  45. Romagnani S. Th1 and Th2 in human diseases. *Clinical immunology and immunopathology.* 1996;80(3):225-35.
  46. Eagar TN, Miller SD. Helper T-cell subsets and control of the inflammatory response. *Clinical immunology: Elsevier;* 2019. p. 235-45. e1.
  47. Pesce B et al. TNF- $\alpha$  affects signature cytokines of Th1 and Th17 T cell subsets through differential actions on TNFR1 and TNFR2. *Int J Mol Sci.* 2022;23(16):9306.
  48. Kaiko GE et al. Immunological decision-making: how does the immune system decide to mount a helper T-cell response? *Immunology.* 2008;123(3):326-38.
  49. Aleebrahim-Dehkordi E et al. T helper type (Th1/Th2) responses to SARS-CoV-2 and influenza A (H1N1) virus: from cytokines produced to immune responses. *Transplant immunology.* 2022;70:101495.
  50. Zhang Y et al. Th1/Th2 cell's function in immune system. *T helper cell differentiation and their function.* 2014:45-65.
  51. Biedermann T et al. TH1 and TH2 Lymphocyte development and



- regulation of TH cell-mediated immune responses of the skin. *Journal of Investigative Dermatology Symposium Proceedings*. 2004; Elsevier.
52. Ryba-Stanisławowska M. Unraveling Th subsets: insights into their role in immune checkpoint inhibitor therapy. *Cellular Oncology*. 2024;1-18.
53. Lin Y et al. New insights on anti-tumor immunity of CD8<sup>+</sup> T cells: cancer stem cells, tumor immune microenvironment and immunotherapy. *Journal of Translational Medicine*. 2025;23(1):341.
54. Maggi E et al. T cell landscape in the microenvironment of human solid tumors. *Immunology Letters*. 2024;106942.
55. Lin J et al. LncRNA MEG3 suppresses hepatocellular carcinoma by stimulating macrophage M1 polarization and modulating immune system via inhibiting CSF-1 in vivo/vitro studies. *International Journal of Biological Macromolecules*. 2024;281:136459.
56. Qin L-X. Inflammatory immune responses in tumor microenvironment and metastasis of hepatocellular carcinoma. *Cancer Microenvironment*. 2012;5(3):203-9.
57. Budhu A, Wang XW. The role of cytokines in hepatocellular carcinoma. *Journal of leukocyte biology*. 2006;80(6):1197-213.
58. Song Y, Yang JM. Role of interleukin (IL)-17 and T-helper (Th) 17 cells in cancer. *Biochemical and biophysical research communications*. 2017;493(1):1-8.
59. Wu MY, Hill CS. TGF- $\beta$  superfamily signaling in embryonic development and homeostasis. *Developmental cell*. 2009;16(3):329-43.
60. Liu S et al. Molecular evolution of transforming growth factor- $\beta$  (TGF- $\beta$ ) gene family and the functional characterization of lamprey TGF- $\beta$ 2. *Frontiers in Immunology*. 2022;13:836226.
61. Chen J, Gingold JA, Su X. Immunomodulatory TGF- $\beta$  signaling in hepatocellular carcinoma. *Trends in molecular medicine*. 2019;25(11):1010-23.
62. Thangaraj JL et al. Disruption of TGF- $\beta$  signaling pathway is required to mediate effective killing of hepatocellular carcinoma by human iPSC-derived NK cells. *Cell Stem Cell*. 2024;31(9):1327-43. e5.
63. Qin D et al. Targeting tumor-infiltrating tregs for improved antitumor responses. *Frontiers in Immunology*. 2024;15:1325946.
64. Fu S et al. TGF- $\beta$  induces Foxp3<sup>+</sup> T-regulatory cells from CD4<sup>+</sup> CD25<sup>-</sup> precursors. *American Journal of Transplantation*. 2004;4(10):1614-27.
65. Babaei G et al. STAT protein family and cardiovascular diseases: overview of pathological mechanisms and therapeutic implications. *Molecular Biology Reports*. 2024;51(1):440.
66. Shi D et al. Structure, function, signaling pathways and clinical therapeutics: The translational potential of STAT3 as a target for cancer therapy. *Biochimica et Biophysica Acta (BBA)-Reviews on Cancer*. 2024:189207.
67. Thuya WL et al. Insights into IL-6/ JAK/STAT3 Signaling in the Tumor Microenvironment: Implications for Cancer Therapy. *Cytokine Growth Factor Rev*. 2025;85:26-42.
68. Songbai H et al. STAT3 Orchestrates Immune Dynamics in Hepatocellular Carcinoma: A Pivotal Nexus in Tumor Progression. *Critical Reviews in Oncology/Hematology*. 2025:104620.
69. Makino Y et al. STAT3 is activated by CTGF-mediated tumor-stroma cross talk to promote HCC progression. *Cellular and molecular gastroenterology and hepatology*. 2023;15(1):99-119.
70. Xu J et al. IL-6/STAT3 is a promising therapeutic target for hepatocellular carcinoma. *Frontiers in oncology*. 2021;11:760971.
71. Xing S, Ferrari de Andrade L. NKG2D and MICA/B shedding: a 'tag game' between NK cells and malignant cells. *Clinical & translational immunology*. 2020;9(12):e1230.
72. Tian Z et al. Macrophages and hepatocellular carcinoma. *Cell & bioscience*. 2019;9:1-10.
73. Hu R et al. STAT3: A key signaling molecule for converting cold to hot tumors. *Cancer letters*. 2020;489:29-40.
74. Zhang Z et al. Wnt/ $\beta$ -catenin signaling in the development and therapeutic resistance of non-small cell lung cancer. *Journal of Translational Medicine*. 2024;22(1):565.
75. Duchartre Y et al. The Wnt signaling pathway in cancer. *Critical reviews in oncology/hematology*. 2016;99:141-9.
76. Morita M et al. Role of  $\beta$ -catenin activation in the tumor immune microenvironment and immunotherapy of hepatocellular carcinoma. *Cancers*. 2023;15(8):2311.
77. Huang Y et al. Activation of Wnt/ $\beta$ -catenin signaling promotes immune evasion via the  $\beta$ -catenin/IKZF1/CCL5 axis in hepatocellular carcinoma. *International Immunopharmacology*. 2024;138:112534.
78. Zhao Z et al. Wnt/ $\beta$ -catenin signaling pathway in hepatocellular carcinoma: pathogenic role and therapeutic target. *Frontiers in Oncology*. 2024;14:1367364.
79. König D et al. Adoptive cell therapy with tumor-infiltrating lymphocytes in combination with nivolumab in patients with advanced melanoma. *Immuno-Oncology and Technology*. 2024;24:100728.
80. Lickefett B et al. Lymphodepletion—an essential but undervalued part of the chimeric antigen receptor T-cell therapy cycle. *Frontiers in immunology*. 2023;14:1303935.
81. Tennant MD et al. Efficient T cell adoptive transfer in lymphoreplete hosts mediated by transient activation of Stat5 signaling. *Molecular Therapy*. 2023;31(9):2591-9.
82. Shen W et al. Immunotherapeutic approaches for treating hepatocellular carcinoma. *Cancers*. 2022;14(20):5013.
83. Matsueda S et al. Recent clinical researches and technological development in TIL therapy. *Cancer Immunology, Immunotherapy*. 2024;73(11):232.
84. Amaria RN et al. Entering a new era of tumor-infiltrating lymphocyte cell therapy innovation. *Cytotherapy*. 2025;27(7):864-73.
85. Poschke IC et al. The outcome of ex vivo TIL expansion is highly influenced by spatial heterogeneity of the tumor T-cell repertoire and differences in intrinsic in vitro growth capacity between T-cell clones. *Clinical cancer research*. 2020;26(16):4289-301.
86. Dougé A et al. Adoptive T cell therapy in solid tumors: state-of-the art, current challenges, and upcoming improvements. *Molecular Cancer Therapeutics*. 2024;23(3):272-84.
87. Dong Y et al. Recent advances and future prospects in immune checkpoint (ICI)-based combination therapy for advanced HCC. *Cancers*. 2021;13(8):1949.
88. Chen X et al. Unveiling the role of tumor-infiltrating T cells and immunotherapy in hepatocellular carcinoma: a comprehensive review. *Cancers*. 2023;15(20):5046.
89. Li Q et al. PD-1/PD-L1 checkpoint inhibitors in advanced hepatocellular carcinoma immunotherapy. *Frontiers in immunology*. 2022;13:1070961.
90. Castet F et al. Atezolizumab plus bevacizumab: a novel breakthrough in hepatocellular carcinoma. *Clinical Cancer Research*. 2021;27(7):1827-9.
91. Shi D et al. Chimeric antigen receptor-glypican-3 T-cell therapy for advanced hepatocellular carcinoma: results of phase I trials. *Clinical Cancer Research*. 2020;26(15):3979-89.



92. Zhou Y et al. CAR-T cell therapy for hepatocellular carcinoma: current trends and challenges. *Frontiers in Immunology*. 2024;15:1489649.
93. Lin T-Y, Su T-H. Progression of portal hypertension after atezolizumab plus bevacizumab for hepatocellular carcinoma-report a case and literature review. *Journal of the Formosan Medical Association*. 2024;123(8):916-9.
94. Ramadan A et al. Personalized treatment approaches in hepatocellular carcinoma. *Arab Journal of Gastroenterology*. 2025;26(1):122-8.
95. Pelizzaro F, Farinati F, Trevisani F. Immune checkpoint inhibitors in hepatocellular carcinoma: current strategies and biomarkers predicting response and/or resistance. *Biomedicines*. 2023;11(4):1020.
96. Ji JH et al. Predictive biomarkers for immune-checkpoint inhibitor treatment response in patients with hepatocellular carcinoma. *International Journal of Molecular Sciences*. 2023;24(8):7640.
97. Ozer M et al. Adoptive cell therapy in hepatocellular carcinoma: a review of clinical trials. *Cancers*. 2023;15(6):1808.
98. Yang Y et al. The predictive value of PD-L1 expression in patients with advanced hepatocellular carcinoma treated with PD-1/PD-L1 inhibitors: A systematic review and meta-analysis. *Cancer Medicine*. 2023;12(8):9282-92.
99. Chen Y et al. The current advances and future directions of PD-1/PD-L1 blockade in head and neck squamous cell carcinoma (HNSCC) in the era of immunotherapy. *International Immunopharmacology*. 2023;120:110329.
100. Liu X et al. The immunosuppressive role of MDSCs in HCC: mechanisms and therapeutic opportunities. *Cell Communication and Signaling*. 2025;23(1):155.
101. Arvanitakis K, Germanidis G. Immunity revealed: Stratifying hepatocellular carcinoma for precision immunotherapy. *Molecular Therapy Oncology*. 2025;33(1).
102. Devan AR et al. The role of glypican-3 in hepatocellular carcinoma: Insights into diagnosis and therapeutic potential. *European Journal of Medical Research*. 2024;29(1):490.
103. Eghbali S, Heumann TR. Next-Generation Immunotherapy for Hepatocellular Carcinoma: Mechanisms of Resistance and Novel Treatment Approaches. *Cancers*. 2025;17(2):236.
104. Tehrani HA et al. GPC-3 in Hepatocellular Carcinoma; A Novel Biomarker and Molecular Target. *Experimental Cell Research*. 2024;114391.
105. Batra SA et al. Glypican-3-specific CAR T cells coexpressing IL15 and IL21 have superior expansion and antitumor activity against hepatocellular carcinoma. *Cancer Immunol Res*. 2020;8(3):309-20.
106. Li X, Hu D. Ligand-restricted synNotch switches enable precision cell therapy. *Trends Immunol*. 2025;46(2):91-3.
107. Wu Z et al. Universal CAR cell therapy: challenges and expanding applications. *Translational Oncology*. 2025;51:102147.
108. Chen T et al. The construction of modular universal chimeric antigen receptor T (MU-CAR-T) cells by covalent linkage of allogeneic T cells and various antibody fragments. *Molecular Cancer*. 2024;23(1):53.
109. Cherkassky L et al. Human CAR T cells with cell-intrinsic PD-1 checkpoint blockade resist tumor-mediated inhibition. *J Clin Invest*. 2016;126(8):3130-44.
110. Cai L et al. Targeting LAG-3, TIM-3, and TIGIT for cancer immunotherapy. *J Hematol Oncol*. 2023;16(1):101.
111. Liu Y et al. Review immune response of targeting CD39 in cancer. *Biomarker Research*. 2023;11(1):63.
112. Wu Y et al. Strategies for the enhancement of IL-21 mediated antitumor activity in solid tumors. *Cytokine*. 2024;184:156787.
113. Wang Y et al. An IL-4/21 inverted cytokine receptor improving CAR-T cell potency in immunosuppressive solid-tumor microenvironment. *Frontiers Immunol*. 2019;10:1691.
114. Ponterio E et al. Oncolytic virus and CAR-T cell therapy in solid tumors. *Frontiers Immunol*. 2024;15:1455163.
115. Štach M et al. Interleukin 21 enhances survival and expansion of CAR T cells Via inhibition of their terminal differentiation during interaction with tumor target cells. *Blood*. 2018;132(Supplement 1):4545.
116. Slominski RM et al. How cancer hijacks the body's homeostasis through the neuroendocrine system. *Trends in neurosciences*. 2023;46(4):263-75.